

Serological Evidence of Avian Mycoplasmosis and Salmonellosis in Different Poultry Birds at the Live Bird Market and Free Range Management System in Taraba State, Nigeria

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Abstract

Both avian mycoplasmosis due to *Mycoplasma gallisepticum* and *Mycoplasma synoviae* and salmonellosis due to *Salmonella pullorum* are responsible for economic losses to the poultry industry worldwide. This study was conducted between April to July, 2017, to determine the Seroprevalence of *Mycoplasma gallisepticum* (MG), *Mycoplasma synoviae* (MS) and *Salmonella Pullorum* (SP) in broiler chickens, layer chickens, indigenous chickens, guinea fowl, indigenous cross - breed chickens (Lacroilers) sampled at the Jalingo live bird market and indigenous chickens on free range. Out of the total of 792 serum samples tested, 60.22% were positive to MG. Also, out of the 275 sera samples tested for MS, 66.18% were positive for MS. *Salmonella pullorum* antibodies were detected in 53.9% of the 920 whole blood samples tested. Highest seropositive rates were detected in the indigenous chickens for both MG and MS with 71.93% and 90% seropositive rates, respectively. The highest antibody presence for SP were found in the layer chickens. The study found that all bird types had detectable antibodies for MG, MS and SP leading to the conclusion that MG, MS and SP antibodies are prevalent in chickens of different types as well as guinea fowl in the study area. The significance of these findings are discussed.

Keywords: Avian Mycoplasmosis; Salmonellosis; Mycoplasma gallisepticum; Mycoplasma synoviae; Salmonella pullorum; Serology; Poultry

Abbreviations

MG: Mycoplasma gallisepticum; MS: Mycoplasma synoviae; SP: Salmonella pullorum

Introduction

Both avian mycoplasmosis caused by *Mycoplasma gallisepticum (MG) and Mycoplasma synoviae (MS)* and salmonellosis due to *Salmo-nella pullorum (SP)* are responsible for economic losses to the poultry industry especially in chickens and turkeys worldwide [1,2]. Avian mycoplasmosis due to *Mycoplasma gallisepticum* and *Mycoplasma synoviae* are the species responsible for greater losses in the poultry industry. Infections are characterized by obstinate hacking cough, sneezing, tracheal rales, decreased feed efficacy, poor carcass quality, and suboptimal egg production in laying birds and reduced quality of day old chicks [3-6].

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Salmonella gallinarum and Salmonella pullorum remain a serious economic problem in countries where measures of control are not efficient and in those where climatic conditions favour the spread of these microorganisms [7]. Salmonella pullorum causes pullorum disease which is a severe septicaemic disease that causes high morbidity and mortality in young birds, especially newly hatched chicks, causing high losses to poultry producers [2]. Clinical signs in chicks includes; anorexia, diarrhoea, dehydration, weakness and high mortality while in adult birds decreased egg production, fertility, hatchability and anorexia and increased mortality are observed [6].

In Nigeria, disease remains a major setback to the poultry industry. Live bird markets could be a major source for the spread of poultry diseases as well as the type of management system. Though serological evidence of *Avian mycoplasma* infection has been established in some regions of Nigeria there is no such study in the live bird market where there is a convergence of birds brought from different villages and localities in Taraba state and indigenous chickens on free range. This calls for concern and that is why the study was undertaken to assess the antibody status of MG, MS and S.P to enable stakeholders to wake up to their responsibilities for strategic control measures.

Materials and Methods

The study was conducted in Jalingo metropolis the capital of Taraba state which is sub-sudan type of climate located between Latitude 8°54'N and Longitude 11°22'E in the North-eastern part of Nigeria. A total of 792 blood samples from broiler chickens, layer chickens, indigenous chickens, a cross-breed (lacroilers) of indigenous and exotic chickens and guinea fowl were collected from the slaughter and dressing section of the Jalingo live bird market (LBM) and from indigenous chickens on free range and the serum extracted and tested for MG antibodies using the MG antigen obtained from Charles River Laboratories, Inc. to perform the serum plate agglutination(SPA) test. Out of the 792 serum samples obtained, only 275 were tested for MS antibodies using *Mycoplasma synoviae* antigen from Charles River Laboratories, Inc. The whole blood test was deployed to test another 920 blood samples also obtained from the slaughter and dressing section of the Jalingo live bird market and indigenous chickens on free range using the *Salmonella pullorum* polyvalent antigen manufactured by Lohmann Animal Health Int., Winslow, Maine 04901 and distributed by Charles River Laboratories, Inc.

All prevalence results and 95% confidence intervals were estimated by the estimated true prevalence with an imperfect test [8]. The apparent prevalence was chosen for this study [9,10].

The analysis of variance (2 factor without replication) was used to determine significant differences in seroprevalence among different bird types.

Results and Discussion

In this study, 792 serum samples were tested for *Mycoplasma gallisepticum* antibodies. Out of the number 477 representing 60.22% were positive. Out of the 374 indigenous chickens tested for MG antibodies at the live bird market (LBM), 269 representing 71.93% were positive and was the highest prevalence. This was followed by the Indigenous chickens on free range (59.09%).

Broiler chickens were next with a Seroprevalence rate of 55.13%. Broilers, layer chickens, indigenous chickens and guinea fowl were all positive for MG antibodies. Details of results are presented in table 1.

	No of positives/no.	Percentages			
	sampled	Apparent prevalence	Estimated true prevalence	95% Confidence interval	
Broiler chicken (LBM)	129/234	55.13	60.82	54 - 69	
Layer Chickens (LBM)	54/113	47.79	52.57	42 - 63	
Indigenous chicken (LBM)	269/374	71.93	79.69	74 - 85	
Guinea fowl (LBM)	8/19	50.00	55.06	29 - 81	
Indigenous (Crossbreed)	3/13	9.68	9.75	2 - 27	
Indigenous chicken (FR)	13/22	59.09	65.27	42 - 86	
Total	477/792	60.22	66.55	62 - 70	

 Table 1: Seroprevalence of Mycoplasma gallisepticum in various bird types in Jalingo live bird market

 and free range indigenous chickens.

 LBM: Live Bird Market; FR: Free Range.

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The results for Seroprevalence of MS antibodies showed that highest Seroprevalence rates were found in indigenous chickens and broilers at the live bird market (LBM) with Seroprevalence rate of 90% each. This was followed by layer chickens (LBM) with 71.68% Seroprevalence and the guinea fowl with 33.33%. The least Seroprevalence rates was recorded in Lacroilers a cross - breed between indigenous and exotic chickens with 6.45% Seroprevalence. Details of the results are as presented in table 2.

	No of	Percentages					
	positives/no. sampled	Apparent prevalence	Estimated true prevalence	95% Confidence interval			
Broiler chicken (LBM)	45/50	90.00	1.00	87 - 100			
Layer Chickens (LBM)	81/113	71.68	79.42	69 - 88			
Indigenous chicken (LBM)	45/50	90.00	1.00	87 - 100			
Guinea fowl (LBM)	3/9	33.33	36.33	09 - 74			
Indigenous - exotic crossbreed (crossbreed)	2/31	6.45	6.13	0.18 - 22			
Indigenous chicken (FR)	6/22	27.27	29.52	13 - 55			
Total	182/275	66.18	73.24	66 - 80			

 Table 2: Seroprevalence of Mycoplasma synoviae in various bird types in Jalingo live bird

 market and free range indigenous chickens.

 LBM: Live Bird Market; FR: Free Range.

Salmonella pullorum antibodies were detected in all types of birds with Layer chickens (LBM) having the highest Seroprevalence (66.37%), followed by guinea fowl (61.11%); Lacroilers (58.62%), Indigenous birds on free scavenging (54.54%), and the least rate (48.71%) recorded in broilers (LBM). Details of the results are as presented in table 3.

	No of	Percentages		
	positives/no. sampled	Apparent prevalence	Estimated true prevalence	95% Confidence interval
Broiler chicken (LBM)	114/234	48.72	53.62	46 - 60
Layer Chickens (LBM)	75/113	66.37	73.45	63 - 83
Indigenous chicken (LBM)	264/504	52.38	57.73	53 - 64
Guinea fowl (LBM)	11/18	61.11	67.54	41 - 90
Indigenous - exotic cross breed (LBM)	17/29	58.62	64.74	43 - 84
Indigenous chicken (FR)	12/22	54.55	60.16	37 - 82
Total	493/920	53.59	59.09	55 - 63

 Table 3: Seroprevalence of Salmonella pullorum in different bird types in Jalingo live bird market

 and free range indigenous chickens. LBM: Live Bird Market; FR: Free Range.

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There were no significant differences (P > 0.05) in Seroprevalence rates among the different bird types for MG, MS and SP.

In the present study the highest (71.93%) MG sero-prevalence which was observed in the indigenous chicken sampled at the live bird market and those indigenous chickens on free range (59.09) is similar to the findings by Abdu., *et al.* [11] who reported Seroprevalence in indigenous chicken of 66.3%. The researchers reported MG Seroprevalence of 38.0 and 38.1 in backyard and intensively managed

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chickens, respectively; figures that are lower than our study. Ahmed., *et al.* [12] however reported higher MG and MS Seroprevalence in indigenous chickens on free range in Niger state, Nigeria. Both MG and MS have been reported in indigenous and exotic chickens elsewhere in Nigeria Orajaka., *et al* [13]. The Seroprevalence of MG antibodies in guinea fowl which was 50.00% is lower to the figure reportedly by Adesiyun and Abdu, [14]. The Seroprevalence of SP which was higher in all bird types is similar to other studies in Nigeria [15,16] and elsewhere in Africa [17-19].

Conclusion

The present study has confirmed that antibodies to MG, MS and SP are prevalent in all types of chickens and guinea fowl in Taraba state, north-eastern Nigeria. It also concluded that the live birds market and birds on free range pose a great danger to the epidemiology of MG, MS and SP in Taraba State, North-eastern, Nigeria.

Conflict of Interest Statement

The authors do not have any conflict of interest.

Bibliography

- 1. Gharaibeh S and Al Roussan D. "The use of molecular techniques in commercial chicken in Jordan". *International Journal of Poultry Science* 7.1 (2008): 28-35.
- 2. Shivaprasad HL. "Fowl typhoid and pullorum disease" 19.2 (2000): 405-424.
- 3. Carpenter TR., *et al.* "Vaccination with F strain *Mycoplasma gallisepticum* to reduce production losses in Layer chickens". *Avian Diseases* 25 (1981): 404-409.
- Chakrabarti A. "Chronic respiratory disease". In: Test Book of Clinical medicine. Kalyani publishers. New Delhi: 110.002 (1993): 360-419.
- Ley DH and Yoder HW. "Mycoplasma gallisepticum infection". In: Diseases of poultry, 10th Edition (BW Calnek with HJ Barnes, CW Beard, LR McDougal and YM Sait, Editions). Moshy Wolfer London (1997): 194-207.
- 6. Hossain KMM., *et al.* "Seroprevalence of *Mycoplasma gallisepticum* infection in chicken in the greater rajshahi District of Bangladesh". *Bangladesh medical and Veterinary Journal* 5 (2007): 09-14.
- Barrow PA and Freitas Neto OC. "Pullorum disease and fowl typhoid New thoughts on old disease: A review". Avian Pathology 40.1 (2011): 1-13.
- 8. Sergeant ESG. "Epitools epidemiological calculators". Australian Veterinary Animal Health Services and Australian Biosecurity Cooperative Research Centre for Emerging Infectious Disease (2016).
- 9. Brown LD., et al. "Interval estimation for a binomial proportion". Statistical Science 16 (2001): 101-133.
- 10. Michiels T., et al. "Prevalence of Mycoplasma gallisepticum and Mycoplasma synoviae in commercial poultry, racing pigeons and wild birds in Belgium". Avian Pathology 45.2 (2016): 244 252.
- 11. Abdu PA., et al. "Survey of Mycoplasma gallisepticum and Mycoplasma synoviae antibodies in chickens in Zaria". Nigerian Journal of Animal Production Research 3.1 (1983): 63-66.

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- Ahmed JS., et al. "Isolation and serological detection of Mycoplasma gallisepticum and Mycoplasma synoviae using a combined MG/ MS enzyme linked immune sorbent assay (ELISA) in indigenous chickens in Niger State, Nigeria". African Journal of Cellular Pathology 4 (2015): 70-73.
- 13. Orajaka LJE., *et al.* "Seroepidemiologic survey of *Mycoplasma* infections in native and exotic chickens in Okulelea District of South Eastern Nigeria". *Journal Sustainable Agricultural Environment* 4.1 (2002): 77-82.
- 14. Adesiyun AA and Abdu PA. "Serological survey of *Mycoplasma gallisepticum* antibody in four domestic poultry species around Zaria, Nigeria". *Bulletin Animal Health Production Africa* 33 (1985): 171-172.
- 15. Onunkwo O and Onoviran O. "Serological survey of mycoplasmosis and pullorum disease in Plateau State of Nigeria". *Kenya Veterinarian* 2 (1978): 33-34.
- 16. Obi OJ and Ike AC. "Prevalence and Antibiogram profile of Salmonellae in intensively reared and Backyard chickens in Nsukka, Area, Nigeria". *Nigeria Journal Biotechnology* 30 (2015): 18-30.
- 17. Endris M., *et al.* "Sero and media culture prevalence of salmonellosis in local and exotic chicken, Debre Zect, Ethiopia". *African Journal of Microbiology Research* 7.12 (2013): 1041-1044.
- 18. Butcher GD. This document is VM 130, one of a series of the Veterinary medicine large Animal Clinical Sciences Department, Florida Cooperative Extension service, Institute of food and Agriculture Sciences, University of Florida (2002).
- 19. Mera UM and Haruna M. "Prevalence of *Mycoplasma gallisepticum* (MG) antibodies in chickens in Sokoto, Nigeria". *East Journal of Veterinary Medicinal Science* 1.5 (2019): 58-59.

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