

Pharmacological Significance of Marine Seaweed, *Halimeda opuntia*

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Abstract

Nowadays, looking for alternative medicine sources is a hot topic because they are readily available, safe, and, more particularly, come from sustainable sources. Seaweed-based medications are generated in numerous pharmaceutical and biotechnology businesses. Green algae known as *Halimeda opuntia* are particularly prevalent in tropical and subtropical coastal waters that are 1 - 2 meters deep. Numerous research point to this species as a key source of bioactivities and bioactive chemicals, both of which could serve as potential therapeutic targets in the pharmaceutical sector. Based on this hypothesis, a literature review that focused on the pharmacological significance of the seaweed *Halimeda opuntia* was conducted. Here, we report some of the species' drug-targeting effects, including its anticancer, antioxidant, cytotoxic, hypoglycemic, and antibacterial capabilities. This literature review will compel researchers to focus on this species in order to create new medications for human welfare.

Keywords: *Halimeda opuntia*; Green Seaweed; Pharmacology; Medicinal Value; Sustainable Drug Development; Bioactive Compounds

Introduction

Marine environment covers almost about 71% of the earth, which possesses versatile organisms from prokaryotes to eukaryotes and those ranges from primary food chain to tertiary level. Mostly, the primary food producers are green seaweeds and some cyanobacteria related organisms. The ecosystems of the marine origin are much tougher than the terrestrial zone and due to the harsh ecosystem [1], organisms in the marine food chain continuously develops strong defense system in their inner body to sustain [2]. Due to the sustainable growth of the marine organism such as protozoan, metazoan, mollusks, phytoplankton and some other marine grasses and seaweeds, they often produce lots of secondary metabolites, though they also have primary metabolites. These secondary metabolites are of great importance in mankind, especially in health [3]. Many drugs are originated from the marine sources and their efficacy are very high as compared to other terrestrial origin [4].

Peoples are dependent on sea for their food, feed, and drugs as well. Specially, seaweeds are promising in relation to human benefits as they are rich source of protein, carbohydrates, fatty acids, and bioactive compounds. Bioactive compounds are a treasure trove of new therapeutic drugs development, which would have anti-inflammatory, analgesics, hypoglycemic, anti-thrombotic, cytotoxic, anti-microbial, anti-diarrheal, and some other medicinal effects [5]. Seaweeds are rich in minerals, dietary fibers and vitamins [6]. So, they are rich in proximates and minerals [7]. Due to their rich nutritional values, seaweeds can lower the risk of cardiovascular diseases (CVDs); protect against metabolic syndromes, cancer and viral diseases; improve lipid status, reduce inflammations, attenuate weight gain, regulate blood

glucose in type 2 diabetic (T2D) patients and improve the health of bone and digestive tract [8]. However, there is a concern that seaweed consumption may increase exposure to toxic heavy metals such as arsenic, aluminum, cadmium, lead, rubidium, silicon, strontium, and tin although their contribution to total elemental intake does not pose any threat to the consumers [9]. Another possible risk of consuming seaweeds may be the intake of dangerously high concentration of iodine [5]. Nevertheless, considering the health benefits of seaweed consumption, they undoubtedly appear a promising component of the ever-growing market of functional foods or nutraceuticals [10].

Since ancient the time, seaweeds have been employed in herbal therapy [11]. Additionally, they are being investigated as potential sources of bifidobacterial right now [12]. Besides, many cancer therapeutic drugs have been evolved from marine macro- and microalgae. In addition, a recently licensed seaweed-based medication for the treatment of Alzheimer's disease (AD), sodium oligomannate (G V -971), modifies the gut microbiome [13]. Thus, marine algae would have emerging role in drug development.

Bioactive compounds from seaweed

Marine algae are the rich source of bioactive compounds and bioactive peptides that have direct human health benefits [14-19]. The bioactive compounds are found both as primary and secondary metabolites in marine algae [3,20]. Bioactive peptides, polysaccharides (laminarins, fucans, galactans, ulvan, alginates, carrageenan etc.), polyphenols, fatty acids (omega-3 fatty acids such as docosahexaenoic acid, eicosapentaenoic acid etc.), vitamins (both fat soluble vitamins such as vitamin A, vitamin D, vitamin E and water-soluble vitamins such as vitamin C, vitamin B12 etc.), carotenoids, terpenoids etc. have been extracted from seaweeds [2,5]. These compounds have potential diversified pharmaceutical activities [5,10]. Moreover, a database called Seaweed Metabolite Database (SWMD) has been created in order to remain focused on seaweed-derived bioactive chemicals that target the pharmaceutical sector [21].

Halimeda opuntia and its characteristics

Halimeda is a genus of warm temperate to tropical macroalgae whose chloroplasts contain the characteristic photosynthetic pigments of the Chlorophyta along with siphonoxanthin and siphonoin, and cell walls are composed of a xylose-based β -1-3 linked xylan. Holdfasts secure their upright, pendent, or sprawling thalli [22]. They have a calcium carbonate endoskeleton that is deposited inside the crevices between the segments formed by medullary filaments [22]. Since this process requires active photosynthesis and the intercellular space to be segregated from the external media by tightly compressed peripheral utricles, the calcification rate in *Halimeda* is enhanced in the light and hindered in the dark [23]. A dense mat of *H. opuntia*, sometimes known as watercress alga, can cover greater areas (about 1 meter) and obscure individual plants. It includes thick, extensively branching clusters of rounded, three-lobed or ribbed leaf-like segments (Figure 1) [22]. It can grow between 10 and 25 cm tall, and its branches are in various planes. It uses its holdfast to cling to hard surfaces. The filament that makes up the holdfast and segments has a diameter of between 0.05 and 0.1 mm and resembles a fungal hypha [24]. *H. opuntia* can be found in all warm seawater on rock about mean sea level down to 2 meters [25]. This species is distributed in tropical and sub-tropical waters, mainly in the Indian Ocean, the Pacific Ocean and also in the Atlantic Ocean [24]. Mostly, it is found in the areas of high predator activity, so that very often it produces a new segment that contains a high concentration of bioactive compounds for their self-defense [26,27].

Bioactive compounds in *Halimeda opuntia*

Generally, the *Halimeda* genus contains various types of bioactive compounds such as alkaloids, halimeditrasetat, halimeditrial, halitunal, udoteal, rhipocephalin, rhipocephenal, some steroids and polyphenols [14,28]. The chemical composition of the extract contains different bioactive metabolites such as Stigmast-5-en-3-ol, (3β), a phytosterol compound found in plants, was abundant in *H. opuntia*. According to Nazarudin, *et al.* [29], phytosterol functions chemically as a compound with high antioxidant activity and a moderate radical scavenger. The extracts also contains a high concentration of neophytadiene, a member of the sesquiterpenoids class of compounds

[30]. It is said to have powerful antioxidant and anti-inflammatory properties [31]. Meanwhile, 1-dodecanol, 3,7,11-trimethyl found in *H. opuntia* methanolic extract may be responsible for biological effects such as anti-tumor and antimicrobial effects [32]. Cholest-4-en-3-one, a metabolite found in *H. opuntia* methanolic extracts, has been reported to have antitumor activity, decreasing breast cancer cell viability [33]. Hexadecanoic acid is a fatty acid found in high concentrations in *Halimeda* species [29,34]. It has been reported to have antioxidant properties and to be cytotoxic to human colorectal carcinoma cells (HCT-116) [30,35]. Meanwhile, heptadecane and 1-eicosanol have previously been studied for bactericidal activity and have proven to be effective antibacterial agents against a variety of Gram positive and Gram-negative bacteria [36,37].



Figure 1: *Halimeda opuntia* species.

Pharmacological prospects of *Halimeda opuntia*

Based on the literature search, it is known that *H. opuntia* contains potent bioactive compounds that have various pharmacological activities such as antimicrobial, antioxidant, cytotoxic, antileishmanial, antityrosinase, anti-plasmodium activities and many more. These bioactivities of *H. opuntia* are due to the presence of phytochemicals in its extracts. Phenolic compounds, flavonoids, alkaloids, steroids, tannins and triterpenes have been detected in its crude extracts [38,39]. A glimpse of pharmacological properties of *Halimeda opuntia* are highlighted below, based on literature tracking and a summary of its pharmacological relevance was mentioned in table 1.

Antioxidant and cytotoxic properties

H. opuntia extracts are known to have antioxidant properties. DPPH scavenging activities have been exhibited by its aqueous extract [40], methanolic extract [41], ethanolic extract [14] and ethyl acetate extract [42]. Polar *H. opuntia* fraction enriched in phenolic compounds was reported to have strong DPPH scavenging activity [43]. One study found that its free phenolic acid (FPA) fractions can increase the expression of superoxide dismutase (SOD) and catalase (CAT) genes and lower the level of thiobarbituric acid reactive substance (TBARS) (a marker of lipid peroxidation) and liver damage in CCl_4 -treated rat models [40]. Another study reported the ability of *H. opuntia* methanolic extract to inhibit lipid peroxidation in linoleic acid system [41]. The same study showed its antityrosinase activity [41], which can prevent and treat reactive oxygen species (ROS)-related diseases [44]. Salt of *H. opuntia* may also have strong antioxidant activity [45].

Bioactivity	Extracts/Compounds	Sample collection site	Assays/Methods/ Models	References
Antibacterial Activity against Gram Negative Bacteria				
<i>Escherichia coli</i>	70% ethanolic extracts	Kodiyaghat, South Andaman	Agar well diffusion	[49]
	50% methanolic extracts	Gulf of Lampung	Disc diffusion	[50]
	Ethanolic and methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[51]
	Methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
	Diethyl ether extracts	Mbudya Island, Tanzania	Agar well diffusion	[52]
<i>Serratia marcescens</i>	Dimethylformamide extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
<i>Pseudomonasaeruginosa</i>	Dimethylformamide extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
<i>Salmonella typhi</i>	50% methanolic extracts	Gulf of Lampung	Disc diffusion	[50]
	Ethanolic and methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[51]
<i>Shigella dysenteriae</i>	Ethanolic and methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[51]
<i>Klebsiella pneumoniae</i>	Ethanolic and methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[51]
	Methanolic extract	Jeddah Coast	Agar well diffusion	[41]
<i>Pseudoalteromonas bacteriolytica</i>	Lipophilic extracts	Atlantic islands, Bahamas	Growth inhibition assay (Kubaneck, <i>et al.</i> 2003)	[55]
<i>Staphylococcus aureus</i>	70% ethanolic extracts	Kodiyaghat, South Andaman	Agar well diffusion	[49]
	Methanolic extracts	Atlantic Coast, Panama	Cylinder plate method	[54]
	Lipid soluble extracts	Magueyes Island, La Parguera	Disc diffusion	[53]
	Diethyl phthalate or 1,2-Benzenedicarboxylic acid, dioctyl ester		Disc diffusion	[56]
	Isodecyl octyl phthalate or 1,2-Benzene dicarboxylic acid, isodecyl octyl ester		Disc diffusion	[56]
	50% methanolic extracts	Gulf of Lampung	Disc diffusion	[50]
	Methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
	Diethyl ether extracts	Mbudya Island, Tanzania	Agar well diffusion	[52]

<i>Bacillus subtilis</i>	Diocetyl phthalate or 1,2-Benzenedicarboxylic acid, dioctyl ester		Disc diffusion	[56]
	Isodecyl octyl phthalate or 1,2-Benzene dicarboxylic acid, isodecyl octyl ester		Disc diffusion	[56]
	Methanolic extracts	Atlantic Coast, Panama	Cylinder plate method	[54]
	50% methanolic extracts	Gulf of Lampung	Disc diffusion	[50]
	Methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
	Diethyl ether extracts	Mbudya Island, Tanzania	Agar well diffusion	[52]
	Lipid soluble extracts	Magueyes Island, La Parguera	Disc diffusion	[53]
<i>Bacillus cereus</i>	Dimethylformamide extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
<i>Enterococcus aerogenes</i>	Ethanollic and methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[51]
<i>Enterococcus faecalis</i>	Methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
Antifungal Activity				
<i>Candida albicans</i>	Methanolic extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
	Diethyl ether extracts	Mbudya Island, Tanzania	Agar well diffusion	[52]
<i>Candida utilis</i>	Dimethylformamide extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
<i>Saccharomyces cerevisiae</i>	Methanolic and dimethylformamide extracts	Red Sea Coast	Disc diffusion Broth dilution	[46]
<i>Halophytophthora spinosa</i>	Lipophilic and hydrophilic extracts	Atlantic islands, Bahamas	Growth inhibition assay (Kubaneck, et al. 2003)	[55]
<i>Schizochytrium gregatum</i>	Lipophilic extracts	Atlantic islands, Bahamas	Disc diffusion	[55]
<i>Aspergillus flavus</i>	Ethyl acetate extracts	Hurghada coast, Red Sea Coast, Egypt	Sabouraud agar well diffusion	[57]
<i>Aspergillus niger</i>	Ethyl acetate extracts	Hurghada coast, Red Sea Coast, Egypt	Sabouraud agar well diffusion	[57]
Antioxidant activity	Aqueous extract (free phenolic acid extracted with tetrahydrofuran)	Bajo de Santa Ana, Havana City, Cuba	DPPH assay TBARS assay	[40]
	Salt	Pramuka Island Kepulauan Seribu	Cuprac method	[45]
	Methanolic extract	Jeddah Coast	DPPH assay inhibition of lipid peroxidation in linoleic acid system	[41]
	Phenolic acids and their soluble and insoluble esters	Bajo de Santa Ana, Havana, Cuba	DPPH assay β -carotene-linoleic acid system	[43]

Hepatoprotective activity	Free phenolic acid fraction	Bajo de Santa Ana, Havana City, Cuba	CCl ₄ treated rats Reverse transcription/ Polymerase chain reaction	[40]
Cytotoxic activity	Methanolic extract	Red Sea Coast	Brine shrimp lethality assay	[46]
	Aqueous extract	Sri Lanka	Brine shrimp lethality assay	[39]
Anti-plasmid activity (on R plasmids of some clinical isolates)	Methanolic extract	Red Sea Coast	Method described by (Deshpande, Dhakephalkar and Kanekar, 2001)	[46]
Antityrosinase activity	Methanolic extract	Jeddah Coast	Modified dopachrome method	[41]
Wound healing activity	Aqueous extract	Coastal areas in Sri Lanka	Scratch would assay using mouse fibroblast (L929) cells	[39]
Interferon β production promoting activity	Methanolic extract	Japan	Poly(I:C) induced human osteosarcoma cell line, MG63	[48]
Antiviral activity against acyclovir-resistant HSV-1 and -2	Dichloromethane:methanol (1:1) extract	Southeastern Brazilian Coast	Vero cells	[58]
Antileishmanial activity	80% alcoholic extract	Bajo de Santa Ana	Promastigotes, amastigotes and peritoneal macrophages from BALB/c mice	[59]

Table 1: Pharmacological relevance of *Halimeda opuntia*.

Methanolic [46] and aqueous [39] extracts of *H. opuntia* may be considerably toxic to brine shrimp nauplii. Halimedatrial, a diterpenoid trialdehyde isolated from *Halimeda* sp., has cytotoxic and antimicrobial activity [26]. 4,9-diacetoxyudoteal, a linear diterpene aldehyde isolated from *H. opuntia*, is toxic to pomacentrid fishes [47]. Another study reported human interferon β production promoting activity of *H. opuntia* methanolic extract [48]. So, *H. opuntia* may be a good source of potent antitumor agents.

Antiviral and antimicrobial properties

Antibacterial activities of *H. opuntia* extracts are well documented. A study reported antibacterial activities of its 70% ethanolic, 70% methanolic and 100% hexane extracts against *E. coli* and *S. aureus* [49]. Another study showed the antibacterial activities of its methanolic extracts against *S. typhi*, *S. aureus*, *B. subtilis* and *E. coli* [50]. In a different study, antibacterial activities of its ethanolic, methanolic, petroleum ether and dimethyl formamide extracts were investigated against *E. coli*, *S. typhi*, *Shigella dysenteriae*, *K. pneumoniae* and *Entero-*

bacter aerogenes [51]. Most extracts, except petroleum ether extract, showed strong antibacterial activities against most of the selected bacteria [51]. In yet another study, antibacterial activities of methanolic, ethanolic, dimethylformamide, hexanic, chloroform, dimethyl sulfoxide and aqueous extracts of *H. opuntia* were assayed against four gram negative (*P. aeruginosa*, *E. coli*, *Proteus vulgaris*, *Serratia marcescens*), six gram positive (*S. aureus*, *Micrococcus luteus*, *Enterococcus faecalis*, *B. subtilis*, *B. cereus*, *B. megaterium*) bacterial species [46]. Methanolic and dimethylformamide extracts showed the highest activity, but no activity was found against *M. luteus*, *B. megaterium* and *P. vulgaris* [46]. Another study demonstrated antibacterial activities of diethyl ether extract of *H. opuntia* against *S. aureus*, *B. subtilis* and *E. coli* [52]. Activities against both *B. subtilis* and *S. aureus* were displayed by its lipid soluble [53] and methanolic extracts [54]. In another study, methanolic extract was found to be very effective against *K. pneumoniae* [41]. A different study reported the lipophilic, but not hydrophilic, constituents of dichloromethane: methanol (1:1) extract of *H. opuntia* to be active against *Pseudoalteromonas bacteriolytica* [55]. Two fatty acids (Dioctyl phthalate and Isodecyl octyl phthalate) isolated from *H. opuntia* have antibacterial activities. Both are active against *B. subtilis* and *S. aureus*, and Isodecyl octyl phthalate is also active against *Streptococcus faecalis* [56].

Antifungal activities of *H. opuntia* extracts have been tested against only a few fungal pathogens. So far, antifungal activities of its extracts have been found against *Candida albicans* [46], *C. utilis* [46], *Saccharomyces cerevisiae* [46], *Halophytophthora spinosa* [55], *Schizochytrium aggregatum* [55], *Aspergillus niger* [57] and *A. flavus* [57]. Its antiviral activity is even less studied. One study found that its dichloromethane: methanol (1:1) extract could be active against acyclovir-resistant Herpes simplex virus-1 and -2 [58]. *H. opuntia* extracts may have antiprotozoal activities. Promising anti-leishmanial activity of its 80% alcoholic extract has been reported [59].

Halimeda opuntia extract also possesses antiviral effect. In a study it was found that *H. opuntia* extract works like MHC class-1 in fish. Study reported that the extract of this species has the ability to activate the MHC class-1, limit the growth of Koi Herpes Virus (KHV) of *Cyprinus carpio* and also maintains the fish cell harmless [60].

Anticoagulant and wound healing properties

Aqueous extracts of *H. opuntia* may have wound-healing activity [39]. Its methanolic extract can cure R-plasmid from some clinical isolates of *E. coli* [46]. *H. opuntia* can be useful in bioremediation because of its ability to uptake heavy metals [61-63]. It can also improve crop production. Priming seeds of *Triticum aestivum* and *Eruca sativa* with aqueous extract of *H. opuntia*, can mitigate harmful effects under lead stress [64] and cadmium stress [38], respectively. Marine-derived sulfated polysaccharides have anti-coagulant, anti-tumor and anti-viral activities [65].

Endophytes

Endophytic microorganisms residing in marine algae and sponges are the chemical synthesizers, both primary and secondary metabolites, that are beneficial to them as well as the host or symbionts [66] Microorganisms associated with *H. opuntia* can also be a good source of bioactive compounds. The ethyl acetate extract and several compounds extracted from its endophytic fungus *Aspergillus versicolor* contains isorhodoptilometrin-1-methyl ether, emodin, 1-methyl emodin, evariquinone, 7-hydroxyemodin 6,8-methyl ether, siderin, arugosin and variculanol that possesses anticancer (in murine colon 38, human colon HCT-116, human lung H-125, HepG2, murine L1210, human CCRF-CEM, human CFU-GM cell line), antimicrobial activities and inhibitory activities against Hepatitis C virus (HCV) protease [67]. Epiphytic bacteria isolated from *H. opuntia* exhibited significant antibacterial activities [68].

Anticancer agent

Algal bioactive compounds can endanger cytotoxicity by various means such as inhibiting the cancer cell invasion, migration, or metastasis and or controlling the utilization of cell growth metabolites [69]. Several reports suggests that *Halimeda opuntia* possesses anti-

cancer properties due to the presence of lots of bioactive compounds. Studies with methanolic extract of *H. opuntia* in estrogen positive breast cancer cell line (MCF7) and mice fibroblast cell line (3T3) showed increased cytotoxicity with IC_{50} value of 25.14 ± 1.02 and 65.23 ± 0.25 $\mu\text{g/ml}$ respectively. The anticancer activity was also found in the same cell lines with colony formation assay and spheroid formation assay, and it was reported that methanolic extract of *H. opuntia* dramatically abrogated the colony formation and spheroid formation tendency and thus, demonstrated its strong anticancer activity [70]. Besides, methanolic extract of this species also increases apoptosis and necrosis in the MCF7 cancer cell lines [60]. It is known that antioxidant system provides increased defense support to human body for the prevention of cancer [69]. Due to the homeostatic conditions, our body is continuously generating reactive oxygen species (ROS), which are quince away by the active antioxidant system [71-73]. Various natural antioxidants are able to scavenge the ROS and *Halimeda opuntia* also possesses lots of antioxidants such as flavonoids, phenolic compounds, carotenoids, chlorophylls and their derivatives that are the prime bioactive compounds for impeding tumor development [74]. In some animal studies, it was reported that due to the presence of polyphenols from *H. opuntia*, lots of free radicals were scavenged leading to the inhibition of tumor development [75].

Hawas., *et al.* [67] investigated that the *Halimeda opuntia* endophyte, *Aspergillus versicolor* also possesses anticancer effect due to the presence of siderin, arugosin and variculanol bioactive compounds. These compounds were investigated in two leukemias (murine L1210 and human CCRF-CEM), four solid tumors (murine colon 38, human colon HCT-116, human lung H-125, human liver HEP-G2), as well as human normal cells (CFUGM) using the disk diffusion assay [67].

Hypoglycemic effect

Due to their bioactive secondary metabolites, marine seaweeds have been intensively investigated for their potential to combat diabetes through multiple pathways. Marine algae have been shown to have a hypoglycemic effect in addition to having the ability to reduce diabetic complications in several *in-vitro* and *in-vivo* investigations that have been completed so far.

Alwaleed., *et al.* [76] reported a pancreatoprotective function of *Halimeda opuntia* ethanol extract in alloxan-induced diabetic murine model. *H. opuntia* extract significantly reduced the pre- and post-prandial blood glucose level. It was also observed that the diabetic kidney functions and liver functions were also improved when *H. opuntia* extract was pre-administered to the alloxan-induced model animals. The hematological parameters were also significantly increased in the *H. opuntia* pre-administered groups compared to the negative control and positive control groups. Thus, Alwaleed., *et al.* suggested the *H. opuntia* extract the best extract for use as a preventative medicine (pre-treatment activity) since it improves diabetics by lowering insulin resistance, lowering blood glucose levels, and regenerating damaged beta cells in the pancreas [76].

Conclusion and Future Direction

Due to promising bioactive chemicals, marine seaweeds have recently undergone substantial research for their medicinal properties and *Halimeda opuntia* species would be considered similarly. The purpose of the current study was to evaluate the potential of *Halimeda opuntia* species as a new source for marine pharmaceuticals, which aims to create revolutionary medicines with natural ingredients and reduce the side effects brought on by synthetic substances. Due to the high cost of medications and the poor purchasing power of the people of many nations, access to vital medicines is currently a commodity that only fewer than 50% of the world's population enjoys.

Traditional medicine, which uses natural substances like plants, herbs, and seaweeds to treat diseases and disorders, is this population's first line of defense against health problems. Therefore, it is possible to combine the use of traditional medicine and contemporary treatment, as is currently the case in industrialized and developed nations like United States of America, Japan, Australia, Canada, and France. The lack of sufficient biochemical characterization of the extracts from seaweeds used in traditional medicine, however, might have serious consequences, such as the resurgence of allergies or the assimilation of poisonous substances that can harm our bodies.

Despite the large number of studies that have been done on the compounds and extracts of *Halimeda opuntia* species, more research needs to be done in order to find novel molecules for use in various biotechnological applications, directly and indirectly enhancing human wellbeing.

Author Contribution

MAA- did literature search and wrote the manuscript; TA- searched literature; UC- edited the manuscript; JA- edited the manuscript; MMA- conceived the idea, designed the research, surveyed the literatures, supervised the work, wrote and edited the manuscript.

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Conflict of Interest

Authors have no conflicts of interest to disclose.

Bibliography

1. Gupta V, et al. "Seaweed metabolomics: A new facet of functional genomics". *Advances in Botanical Research* (2014): 31-52.
2. Salehi B, et al. "Current trends on seaweeds: Looking at chemical composition, phytopharmacology, and cosmetic applications". *Molecules* (2019): 4128.
3. Kasanah N, et al. "Review antibacterial compounds from red seaweeds (Rhodophyta)". *Indonesian Journal of Chemistry* 15.2 (2015): 201-209.
4. Manach C, et al. "Polyphenols: Food sources and bioavailability". *The American Journal of Clinical Nutrition* (2004).
5. Cherry P, et al. "Risks and benefits of consuming edible seaweeds". *Nutrition Reviews* (2019).
6. MacArtain P, et al. "Nutritional value of edible seaweeds". *Nutrition Reviews* 65.12 (2007): 535-543.
7. Smitha JL, et al. "Nutrient and heavy metal content of edible seaweeds in New Zealand". *New Zealand Journal of Crop and Horticultural Science* 38 (2010): 19-28.
8. Brown EM, et al. "Seaweed and human health". *Nutrition Reviews* 72 (2014): 205-216.
9. Desideri D, et al. "Essential and toxic elements in seaweeds for human consumption". *Journal of Toxicology and Environmental Health* 79 (2016): 112-122.
10. Mohamed S, et al. "Seaweeds: A sustainable functional food for complementary and alternative therapy". *Trends in Food Science and Technology* 23 (2012): 83-96.
11. Chengkui Z, et al. "Chinese seaweeds in herbal medicine". *Hydrobiologia* 116.117 (1984): 152-154.
12. De Jesus Raposo MF, et al. "Emergent sources of prebiotics: Seaweeds and microalgae". *Marine Drugs* (2016).

13. Wang X., et al. "Sodium oligomannate therapeutically remodels gut microbiota and suppresses gut bacterial amino acids-shaped neuroinflammation to inhibit Alzheimer's disease progression". *Cell Research* (2019).
14. Ahsan T. et al. "Phytochemical screening and evaluation of antioxidant and cytotoxic activities of *Halimeda opuntia*". *Journal of Marine Biology and Aquaculture* 6.1 (2020): 1-6.
15. Islam T., et al. "Bioactive Compounds Screening and In Vitro Appraisal of Potential Antioxidant and Cytotoxicity of *Cladophoropsis* sp. Isolated from the Bay of Bengal". *EC Pharmacology and Toxicology* 8.10 (2020): 19-31.
16. Islam T., et al. "Pharmacological prospects of *Cladophoropsis* sp. seaweed". *Journal of Earth and Ocean Science* 1.1 (2021): 9-24.
17. Alam MM. "Prospect of marine bioactive peptides as DPP4 inhibitor". *Oceanography and Fisheries Open access Journal (OFOAJ)* 14.5 (2022): 555896.
18. Amin MA., et al. "Green seaweed *Ulva lactuca*, a potential source of bioactive peptides revealed by in silico analysis". *Informatics in Medicine Unlocked* 33 (2022): 101099.
19. Akter J. et al. "Pharmacological importance of *Gracilariopsis lemaniformis* seaweed". *Journal of Rangamati Science and Technology University* 1.1 (2022).
20. Alam MM. "Therapeutic potential of marine bioactive compounds against SARS-CoV2 infection". *CPQ Medicine* 11.1 (2020): 1-18.
21. Davis GDJ., et al. "Seaweed metabolite database (SWMD): A database of natural compounds from marine algae". *Bioinformatics* 5 (2011): 361-364.
22. Drew E. "Halimeda". *Encyclopedia of Modern Coral Reefs* (2011): 535-539.
23. Borowitzka MA., et al. "Calcification in the Green Alga *Halimeda*". *Journal of Experimental Botany* 27.5 (1976): 879-893.
24. Hillis-Colinvaux L. "Ecology and taxonomy of *Halimeda*: primary producer of coral reefs". *Advances in Marine Biology* 17 (1980): 0120261170.
25. Guiry MD. "AlgaeBase". World-wide electronic publication, National University of Ireland, Galway (2020).
26. Paul VJ., et al. "Isolation of Halimedatrial: Chemical Defense Adaptation in the Calcareous Reef-Building Alga *Halimeda*". *Science* 221.4612 (1983): 747-749.
27. Paul VJ., et al. "Use of ingested algal diterpenoids by *Elysia halimeda* Macnae (Opisthobranchia: Ascoglossa) as antipredator defenses". *The Journal of Experimental Marine Biology and Ecology* 119 (1988): 15-29.
28. Dini I., et al. "Alkaloid Caulerpin and cytotoxic activity against NCL-H460 lung cancer cells isolated along with beta-sitosterol from the *Halimeda cylindracea* Decaisne". *Sains Malaysiana* 50.9 (2021): 2663-2674.
29. Nazarudin MF., et al. "Metabolic variations in seaweed, *Sargassum polycystum* samples subjected to different drying methods via 1H NMR-based metabolomics and their bioactivity in diverse solvent extracts". *The Arabian Journal of Chemistry* 13.11 (2020): 7652-7664.
30. Mujeeb F., et al. "Phytochemical evaluation, antimicrobial activity, and determination of bioactive components from leaves of *Aegle marmelos*". *BioMed Research International* 2014 (2014): 1-11.

31. Venkataraman B., *et al.* "Antibacterial, antioxidant activity and GC-MS analysis of *Eupatorium odoratum*". *Asian Journal of Pharmaceutical and Clinical Research* 5.2 (2012): 99-106.
32. Esghaei M., *et al.* "Evaluation of anticancer activity of *Camellia sinensis* in the caco-2 colorectal cancer cell line". *Asian Pacific Journal of Cancer Prevention* 19.6 (2018): 1697-1701.
33. Elia J., *et al.* "4-cholesten-3-one decreases breast cancer cell viability and alters membrane raft-localized EGFR expression by reducing lipogenesis and enhancing LXR-dependent cholesterol transporters". *Lipids in Health and Disease* 18.168 (2019): 1-15.
34. Nazarudin MF, *et al.* "Chemical composition and evaluation of the α -glucosidase inhibitory and cytotoxic properties of marine algae *Ulva intestinalis*, *Halimeda macroloba*, and *Sargassum ilicifolium*". *Evidence-Based Complementary and Alternative Medicine* 2020 (2020): 1-13.
35. Ravi L., *et al.* "Cytotoxic potential of N-hexadecanoic acid extracted from *Kigelia pinnata* Leaves". *The Asian Journal of Cell Biology* 12.1 (2017): 20-27.
36. Kubo I., *et al.* "Antibacterial activity of long-chain alcohols: the role of hydrophobic alkyl groups". *Bioorganic and Medicinal Chemistry Letters* 3.6 (1993): 1305-1308.
37. Ozdemir G., *et al.* "Antibacterial activity of volatile extracts of *Spirulina plantensis*". *Phytotherapy Research* 18 (2004): 754-757.
38. Saad-Allah K., *et al.* "Protective Role of the Seaweed *Halimeda opuntia* Extract on Cadmium-Stressed *Eruca sativa* (Mill.)". *Egyptian Journal of Botany* 56.3 (2006): 863-881.
39. Premarathna AD. *et al.* "Preliminary screening of the aqueous extracts of twenty-three different seaweed species in Sri Lanka with *in-vitro* and *in-vivo* assays". *Heliyon* 6.6 (2020): e03918.
40. De Oliveira e Silva AM., *et al.* "*In vivo* and *in vitro* antioxidant activity and hepatoprotective properties of polyphenols from *Halimeda opuntia* (Linnaeus) Lamouroux". *Redox Report* 17.2 (2012): 47-53.
41. Hamza AH., *et al.* "Potential Antimicrobial, Antioxidant and Anityrosinase Activities achieved by Selected Species of Marine Macroalgae". *Journal of Pure and Applied Microbiology* 8 (2014): 257-265.
42. Gazali M., *et al.* "The Screening of Green Algae *Halimeda opuntia* (Linnaeus) as an Antioxidant from the Coast of West Aceh". *Jurnal Ilmu Pertanian Indonesia* 24.3 (2019): 267-272.
43. Vidal A., *et al.* "Antioxidant activity and polyphenols of seaweed species *Halimeda opuntia* and *Halimeda monile*". *ARS Pharmaceutica* 50 (2009): 24-31.
44. Zuo AR., *et al.* "The antityrosinase and antioxidant activities of flavonoids dominated by the number and location of phenolic hydroxyl groups". *Chinese Medicine* 13.1 (2018) 51.
45. Nufus C., *et al.* "Characteristics of green seaweed salt as alternative salt for hypertensive patients". *IOP Conference Series: Earth and Environmental Science* 278.1 (2019): 012050.
46. Selim SA. "Antimicrobial, Antiplasmid and Cytotoxicity Potentials of Marine Algae *Halimeda opuntia* and *Sarconema filiforme* collected from Red Sea Coast". *International Journal of Medical, Health, Biomedical, Bioengineering and Pharmaceutical Engineering* 6.1 (2012): 79-84.

47. Tillekeratne LMV, *et al.* "4,9-diacetoxyudoteal: A linear diterpene aldehyde from the green alga *Halimeda opuntia*". *Phytochemistry* 23.6 (1084) 1331-1333.
48. Nakano T, *et al.* "In vitro promoting activity of human interferon β production by extracts of marine algae from Japan". *Cytotechnology* 25.1-3 (1997): 239-241.
49. Mishra JK, *et al.* "Antibacterial Activity of Seaweed *Halimeda opuntia* From the Coasts of Souths Andaman". *Global Journal of Bio-Science and Biotechnology* 5.3 (2016): 345-348.
50. Hendri MH. "Antibacterial Potential Screening of Halimeda sp on Some Types of Pathogenic Bacteria". *International Journal of Marine Science* 53 (2015): 1-6.
51. Al-Judaibi A. "Antibacterial Effects of Extracts of Two Types of Red Sea Algae". *Journal of Biosciences and Medicines* 2.2 (2014): 74-82.
52. Mtolera MS, *et al.* "Antimicrobial activity of extracts from six green algae from Tanzania". *Current Trends in Marine Botanical research In East African Region* (1996): 211-217.
53. Ballantine DL, *et al.* "Antibiotic activity of lipid-soluble extracts from Caribbean marine algae". Twelfth International Seaweed Symposium". *Dordrecht: Springer Netherlands* (1987): 463-469.
54. Gupta M, *et al.* "Antimicrobial Activity of Various Algae of the Panamanian Atlantic Coast". *Revista Médica de Panamá* 16.1 (1991): 64-68.
55. Engel S, *et al.* "Antimicrobial activities of extracts from tropical Atlantic marine plants against marine pathogens and saprophytes". *Marine Biology* 149.5 (2006): 991-1002.
56. Anggadiredja JT. "Diversity of Antibacterial Compounds from *Eucheuma Serra*, *Halimeda Opuntia*". *Jurnal Teknologi Lingkungan* 12.2 (2011): 131-142.
57. Ahmed AE, *et al.* "Diversity of Toxigenic Molds and Mycotoxins Isolated from Dairy Products: Antifungal Activity of Egyptian Marine Algae on *Aspergillus* and *Candida* Species". *Journal of Pure and Applied Microbiology* 14.1 (2020): 215-232.
58. Soares AR, *et al.* "Antiviral activity of extracts from Brazilian seaweeds against herpes simplex virus". *Revista Brasileira de Farmacognosia* 22.4 (2012): 714-723.
59. Parra MG, *et al.* "Actividad antileishmanial de seis extractos de organismos marinos". *Revista Cubana de Medicina tropical* 64.1 (2012): 61-64.
60. Yanuhar U, *et al.* "The effect of crude protein *Halimeda* sp. On *Cyprinus carpio* infected Koi Harpes Virus on expression of major histocompatibility complex class-1". *Journal of Fisheries and Marine Research* 1.1 (2017): 15-19.
61. Kuyucak N, *et al.* "Accumulation of Cobalt by Marine Alga". *Biotechnology and Bioengineering* 33 (1989): 809-814.
62. Volesky B. "Advances in biosorption of metals: Selection of biomass types" *FEMS Microbiology Reviews* 14 (1994): 291-302.
63. Mutia G, *et al.* "Analysis of Bio-Accumulation of Heavy Metals in Seaweeds *Ulva rigida* and *Halimeda opuntia* in Validation of Their Safety for Use in Aquaculture Feeds in Kenya". *IOSR Journal of Environmental Science, Toxicology and Food Technology* 12.8 (2018): 56-63.

64. Nessim A., et al. "Mitigation of Lead Stress in *Triticum aestivum* by Seed Priming in Aqueous Extracts of The Macroalga *Halimeda opuntia* and *Codium fragile*". *Egyptian Journal of Botany* 58.2 (2018): 263-274.
65. Udayangani RMAC., et al. "Potential Health Benefits of Sulfated Polysaccharides from Marine Algae". *Encyclopedia of Marine Biotechnology* (2020): 629-635.
66. Chowdhury KR., et al. "Pharmaceutical potential of endophytes associated to marine sponge and algae from the Bay of Bengal and their contribution to the blue economy of Bangladesh". *International Journal of Pharmaceutical Sciences and Research* 13.5 (2022): 2013-2019.
67. Hawas UW., et al. "Bioactive anthraquinones from endophytic fungus *Aspergillus versicolor* isolated from red sea algae". *Archives of Pharmacal Research* 35.10 (2012): 1749-1756.
68. Basondwah SH., et al. "Epiphytic Bacteria Associated with the Green Algal *Halimeda Opuntia* as a Source of Antibacterial Agent". *IOSR Journal of Pharmacy and Biological Sciences* 14.3 (2019): 79-85.
69. Alam MM. "Essence of antioxidants in aging science: NRF2, a true fact". *CPQ Medicine* 5.5 (2019): 1-5.
70. Nazarudin MF., et al. "Preliminary screening of antioxidant and cytotoxic potential of green seaweed, *Halimeda opuntia* (Linnaeus) Lamouroux". *Saudi Journal of Biological Sciences* 29.4 (2022): 2698-2705.
71. Goto M., et al. "Alcohol dehydrogenase 3 contributes to the protection of liver from nonalcoholic steatohepatitis". *Genes to Cells* 20 (2015): 464-480.
72. Alam MM., et al. "Glucocorticoid receptor signaling represses the antioxidant response by inhibiting histone acetylation mediated by the transcriptional activator NRF2". *Journal of Biological Chemistry* 292.18 (2017): 7519-7530.
73. Okazaki K., et al. "Enhancer remodeling promotes tumor-initiating activity in NRF2-activated non-small cell lung cancers". *Nature Communications* 11 (2020): 5911.
74. Elia J., et al. "4-cholesten-3-one decreases breast cancer cell viability and alters membrane raft-localized EGFR expression by reducing lipogenesis and enhancing LXR-dependent cholesterol transporters". *Lipids in Health and Disease* 18.168 (2019): 1-15.
75. Yoshie Y., et al. "Compositional difference of phenolic compounds between two seaweeds, *Halimeda* spp". *Journal of the Tokyo University of Fisheries* 88 (2002): 21-24.
76. Alwaleed EA., et al. "Evaluation of the pancreatoprotective effect of algal extracts on alloxan-induced diabetic rat". *Bioactive Carbohydrates and Dietary Fibre* 24 (2020): 100237.

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