

Antifungal Potential of Polyalthia longifolia against Rhizoctonia solani

Poonam Meena*, Tripta Jain and Kanika Sharma

Microbial Research Laboratory, Department of Botany, University College of Science, MLSU, Udaipur Rajasthan, India

*Corresponding Author: Poonam Meena, Microbial Research Laboratory, Department of Botany, University College of Science, MLSU, Udaipur Rajasthan, India.

Received: January 07, 2022; Published: March 28, 2022

Abstract

Use of synthetic fungicides in inappropriate manner have negative effects on whole ecosystem with a possible carcinogenic risk on human health so there is a strong need to search a strategy which are environmentally safe as well as an economically viable. In this continuation present study reveals the antifungal activity of partially purified extracts of *Polyalthia longifolia* leaf against *Rhizoctonia solani* which is responsible for black scurf disease of potato. Partially purified extracts of *P. longifolia* leaf were prepared by hot extraction method in different organic solvents. Among all the organic solvents used for extraction, maximum percent extractive value was obtained with petroleum ether fraction which was 13.25%. Acetone were used for solubility of extract because acetone did not show any toxic effect on test pathogen. All partially purified fractions were subjected for phytochemical screening by standard methods. Maximum metabolites were found in petroleum ether, acetone and aqueous fractions. Poison food technique was used for determination of antifungal activity. Among all fractions, maximum inhibition of test pathogen was showed by petroleum ether fraction which is 81.25%. MIC and MFC were performed by two fold serial dilution method. 0.625 mg/ml was found as MIC and 1.25 mg/ml was MFC. Mancozeb and Bavistin were used as standard fungicides. Results suggest that Petroleum ether fraction have maximum secondary metabolites which are the reason of maximum inhibition of pathogen so we can conclude that *Polyalthia longifolia* leaf either in the form of extract or powder can be used as ecofriendly herbal biocontrol agent against *Rhizoctonia solani*.

Keywords: Antifungal Activity; Hot Extraction; Poison Food Technique; MIC; MFC

Introduction

The today's need to control plant diseases because most of the synthetic fungicides leave toxic substance which impact adverse effect on environment [1,2] and increases the resistant pathogen populations. So, to overcome this global safer measures have to develop which will be economically viable and free from the dependency on the synthetic agrochemicals for plant disease control [3-6]. Secondary metabolites present in the plants are well known source of natural antifungal agents because they are truly natural so do not leave any toxic substance on plants, animals, soil and on complete ecosystem. They are novel environmentally safer compounds which are a treasure of nature which can be used separately or can be mixed all together to form an herbal bioformulation as they are plant based bioproducts.

Secondary metabolites have been isolated in the form of plant extracts, essential oil and studied antifungal activity by several workers [7,8] and people are using these strategy for biological control of plant diseases and arrange biological control programs [9-11]. Biological

Citation: Poonam Meena., et al. "Antifungal Potential of Polyalthia longifolia against Rhizoctonia solani". EC Pharmacology and Toxicology 10.4 (2022): 18-27.

control has the potential to manage plant disease which associated with various phenomenon Plant extracts is usually prepared with cold and hot extraction methods [12-15]. Ethanol or Methanol is universal solvents which use for preliminary screening of active principles of plants. Crude extract is used for screening of antifungal activity of plant material which is followed by hot extract which is prepared by successive extraction method [16].

Partially purified extracts are used for studies of various phytochemicals, responsible for antimicrobial activity. The medicinal and antimicrobial activities of plant species are due to the phytochemical constituents present in it. Plants are full of secondary metabolites viz. tannins, terpenoids, alkaloids, and flavonoids [17,18] and they have well known antimicrobial properties. Antifungal agents are natural products which have always been found a better treatment control of fungi [19]. Antimicrobial properties are very well known factor of plants since ancient time. Many literatures have reported about the inhibitory effect of plants and their control of plant diseases. In the present study antifungal *Polyalthia longifolia* leaves were detected against *Rhizoctonia solani*.

Polyalthia longifolia is a lofty evergreen plant, native to India, commonly planted due to its effectiveness in alleviating noise pollution. Many common names are given to *P. longifolia* but most common name is ashoka, "the buddha tree" [20]. Its growth is symmetrical pyramidal with pendulous branches and long narrow lanceolate leaves. The tree is known to grow over 30 ft in height. This plant is known for its antibacterial, antioxidant, anti-inflammatory, anti-cancer, hepato-protective and anti-hyperglycemic activity [21] but there is little or no information about the antifungal activity of *P. longifolia* against plant pathogenic fungi *R. solani* which is our test pathogen therefore, the main object of our study is to determine the antifungal activities of *P. longifolia* partially purified extracts using *in vitro* antimicrobial screening methods.

Materials and Methods

Collection and identification of plant material

The healthy infection free leaves of *Polyalthia longifolia* were collected from the campus of University College of science, Mohanlal Sukhadia University, Udaipur, Rajasthan, India. The herbarium specimen was identified by Dr. Amit Kotia, In charge Herbarium, Department of Botany, and University college of Rajasthan, Jaipur, India.

Preparation of extract

Test Plant material leaves were dried in shade at room temperature and grounded with electric grinder. The ground material was passed through sieve of mesh size 60 to obtain a fine powder which was used to prepare partially purified extract by hot extraction method.

Hot extraction

In this extraction method continuous extraction of powdered plant material in soxhlet apparatus with various organic solvents were used for separation of different phytochemical constituents [22,23]. Solvent series used for successive separation was non-polar to polar i.e.

Petroleum ether \rightarrow Benzene \rightarrow Chloroform \rightarrow Acetone \rightarrow Alcohol \rightarrow Methanol \rightarrow Water.

In this method the dried extract and fractions were weighed and their percent extractive value was also calculated percentage by the following formula given below.

Citation: Poonam Meena, et al. "Antifungal Potential of Polyalthia longifolia against Rhizoctonia solani". EC Pharmacology and Toxicology 10.4 (2022): 18-27.

Percent extractive value

Percent extractive = $\frac{\text{Weight } \mathbf{\delta} \quad \text{dried extract}}{\text{Weight } \mathbf{\delta} \quad \text{dried plant material}} \times 100$

Phytochemical study of Polyalthia longifolia leaf extract

Various fractions obtained by hot extraction of leaves of test plant were than subjected to qualitative test for the identification of phytochemical constituents.

Inhibitory activity of solvent against test pathogen

Besides the solubility, inhibitory activity of solvent is also an important requirement for the selection of suitable solvent. Best solvent should have good solubility as well as less toxicity. To ascertain less toxic solvent all the solvents examined in solubility testing were assayed for finding inhibitory effect on test pathogen using poison food technique.

Assay of antifungal activity of partially purified extracts

The inhibitory activity of partially purified extracts of *Polyalthia longifolia* was assayed by poison food technique [24,25]. 10 mg/ ml concentration of stock solution was used for antifungal activity. 2 ml of stock solution was mixed with 18 ml molten sterile culture medium and this mixture was poured into pre-sterilized petri-plates (9 cm diameter) and allowed to solidify at room temperature. In the control set no extract was used. The plates were then incubated at 28 ± 2 °C for seven days. Culture control and acetone control were also maintained along with test samples. Antifungal activity was measured as a function of increase in growth of 6 mm disc of inoculum.

The average diameter of the fungal colonies was measured on the seventh day of incubation and percentage of mycelial growth inhibition was calculated by the following formula given below:

Percent Mycelial growth inhibition = gc-gt/gc×100

Where:

gc = Growth of mycelia colony after incubation period in control set subtracting the diameter of inoculum's disc

gt = Growth of mycelia colony after incubation period in treatment set subtracting the diameter of inoculum's disc.

Estimation of minimum inhibitory concentration (MIC) of selected plant extract

Minimum inhibitory concentration (MIC) was determined by broth dilution method [26,27]. Potato dextrose broth (PDB) was used for determining inhibitory activity. 20 mg/ml concentration of extract was prepared for stock solution. Two fold serial dilution method was used for the preparation of 10 mg/ml to 0.019 mg/ml concentration from the stock solution and subsequently autoclaved. The final concentration was serially diluted with sterile potato dextrose broth medium to attain final concentrations. All these tubes were than respectively inoculated with 10 μ l of spore suspension (1 × 10⁶ spores/ml) and incubated at 27 ± 2°C for 72h. One tube containing extract free autoclaved medium was used as control. Three replicates of each concentration were maintained and experiment was repeated thrice.

Estimation of MFC of selected plant extract

A loopful of fungal biomass from all of the above mentioned tubes containing 9 ml broth medium and MIC other concentrations respectively were streaked onto the extract free PDA slants and incubated at $27 \pm 2^{\circ}$ C for 72h. Presence or absence of growth was observed after respective incubation time (7th day). Appearance of growth indicates that the extract concentration is just fungistatic and absence of growth indicates that extract concentration is fungicidal.

Statistical analysis

All experiments were performed in triplicates (n = 3) and the data were presented as the mean \pm standard deviation.

Results and Observations

The percent extractive values of partially purified leaf fractions in various organic solvents are given in table 1. Petroleum ether fraction gave maximum percent extractive value of 13.25% whereas alcohol, acetone, benzene, chloroform, acetone and aqueous fraction of *Polyalthia longifolia* gave 4.8, 4.5, 4.2 and 0.7 extract value respectively.

S. No.	Type of Extract	Weight of Dried extract (gm)	% Extractive Value
1.	Petroleum ether fraction	2.65	13.25
2.	Benzene fraction	0.84	4.2
3.	Chloroform fraction	0.4	2
4.	Acetone fraction	0.9	4.5
5.	Alcohol fraction	0.96	4.8
6.	Aqueous fraction	0.14	0.7

Table 1: Percent extractive value of different partially purified fractions of Polyalthia longifolia leaf extract.

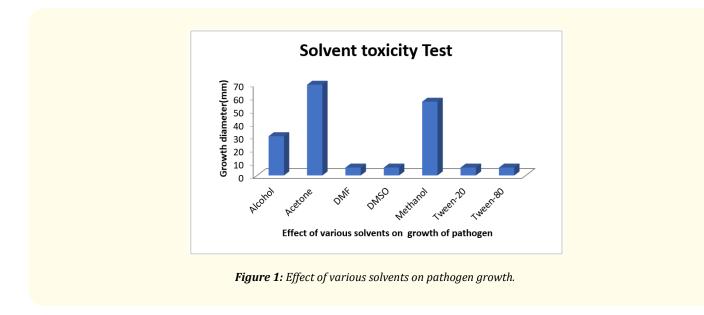
Results of phytochemical testing of *Polyalthia longifolia* leaves extract obtained in different solvents are given in table 2. Alkaloids, phytosterols, carbohydrate, saponins and flavanoids are present on petroleum ether fraction. In benzene only saponins and flavanoids are present. In chloroform fraction only saponins are present and rest of all absent. In acetone fraction, alkaloids, phytosterols and volatile oil are absent and remaining metabolites are present. In alcohol fraction only phytosterol and volatile oil are present. In aqueous fraction, phytosterols and tannins are absent and remaining metabolites are present.

S. No.	Fractions	Alkaloids	Phytosterols	Volatile oil	Tannins	Carbohydrate	Saponins	Flavonoids
1.	PE	+	+	-	-	+	+	+
2.	Benzene	-	-	-	-	-	+	+
3.	Chloroform	-	-	-	-	-	+	-
4.	Acetone	-	-	-	+	+	+	+
5.	Alcohol	-	+	+	-	-	-	-
6.	Aqueous	+	-	+	-	+	+	+

Table 2: Phytochemical screening of various fractions of Polyalthia longifolia leaf extract.

+ ve: Present, - ve: Absent.

22



Antifungal activity of partially purified extract

Results of antifungal activity of partially purified extract fraction of *Polyalthia longifolia* leaf and standard chemical fungicides against *R. solani* are given in figure 2. Amongst comparative inhibitory activity of mancozeb and bavistin, mancozeb was significantly active against *R. solani*. Significant inhibition of *R. solani* was observed with all partially purified fractions but petroleum ether fraction showed maximum inhibition with 81.25% followed by acetone (73.33%) Chloroform fraction (65.83%), benzene fraction (64.92%), Ab.alcohol (62.92%) and aqueous fraction (30.83%).

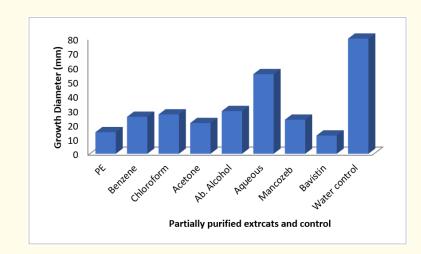


Figure 2: Antifungal activity of partially purified extract of Polyalthia longifolia.

MIC and MFC of petroleum ether fraction

Petroleum ether fraction was used for further studies as this fraction was found to be most effective. MIC and MFC are presented in table 3. 0.019 mg/ml to 10 mg/ml of petroleum ether extract was assayed and MIC was found to be 0.625 mg/ml for *Alternaria solani* and MFC for this fungus was observed at 1.25 mg/ml.

S. No.	Test pathogen	MIC (mg/ml)	MFC (mg/ml)
1.	Rhizoctonia solani	0.625	1.25

Table 3: MIC and MFC of petroleum ether fraction of Polyalthia longifolia leaf extract.

Discussion

Chemical fungicides impose the adverse effect on all living and non living objectives present on earth. Hence, there is a great need for natural antifungal belonging to plants which can be acted on new targets with fewer side effects [28].

The traditional use of plants as medicines provides the basic knowledge for selecting plant ingredients either in the form of essential oils/extracts/powder. It is important to investigate scientifically validate and investigate the plants which have been used in traditional medicines as potential sources of novel antimicrobial compounds [29]. Also, the resurgence of interest in natural therapies and increasing consumer demand for effective, safe, natural products means that quantitative data on plant oils and extracts is required [30,31]. Various secondary metabolites are responsible for antimicrobial potential [32-36].

Secondary metabolites were found effective against many pathogenic bacteria, fungi and viruses and even found active against drug resistant microorganisms [37]. Plant extracts and pure compounds have shown antimicrobial activity against pathogens can be used as nutraceuticals and in food preservation [38-40].

The poison food technique was used for the assay of antifungal activity depends on the inhibition of microorganism growth as an indication of activity which is measured as a function of diameter of the growth zone [41]. The dilution methods indicate the activity in terms of MIC and MFC also. Hence, in the present study MIC of plant extracts and essential oils against the test fungi was done by macro broth two fold serial dilution techniques [42].

Rhizoctonia solani is the common soil inhibiting plant pathogenic fungus which causes black scruff disease in potato [43]. Natural chemicals and their use for integrated plant protection is one of the focuses of research workers all over the world. The results of the present investigation are clear indication for plant extracts to control fungal pathogen [44].

Antimicrobial screening with various organic solvents because the primary metabolites are soluble in water whereas secondary metabolites are soluble in organic solvents [45]. These extracts are studied to search for various compounds, responsible for antimicrobial activity.

Results of the present study suggested that petroleum ether extract gave best activity against *R. solani*. It can be due to better solubility of the active molecules like terpenoids, flavanoids, tannins in petroleum ether as compared to water. [46] reported that extract prepared with an organic solvent exhibits better antimicrobial activity. Stronger extraction capacity directly proportion to produce greater number of active constituents responsible for antimicrobial activity [47-49].

Conclusion

Antifungal activity of partially purified extracts of *P. longifolia* leaf against *R. solani* was studied by poison food technique and broth dilution method. Petroleum ether fraction showed maximum inhibition against *R. solani*. This indicates the probably active compound is present in this fraction. On the basis of results obtained it can be concluded that partially purified petroleum ether fraction of *Polyalthia longifolia* leaf showed maximum inhibition against *Rhizoctonia solani* as compare to standard fungicides. This was used to develop an eco-friendly natural fungicide which will not leave any toxic substance in the environment

Conflicts of Interest

There are no conflicts of interest. As this is my original research work.

Research Involving Human Participants and/or Animals

In this study no use of any kind of animals for the experimental purpose.

Authors Contribution Statement

Ms. Poonam Meena perceived the idea, carried out the research study, evaluated the results and drafted the manuscript. Dr. Tripta Jain guided to Ms. Meena in conducting this research study and also reviewed and approved the manuscript.

Funding Support

There is no funding for the work.

Bibliography

- 1. Gurjar MS., et al. "Efficacy of plant extracts in plant disease management". Agricultural Sciences 3.3 (2012): 425-433.
- 2. Barupal T., *et al.* "Inhibitory effects of leaf extract of *Lawsonia inermis* on *Curvularia* lunata and characterization of novel inhibitory compounds by GC-MS analysis". *Biotechnology Reports* 23 (2019) e00335.
- Camps VRE., et al. "Use of botanical insecticides for sustainable agriculture: Future Perspectives". Ecological Indicators 105 (2019): 483-495.
- 4. Karthic Rajamanickam., et al. "In-vitro antimicrobial activity and In-vivo toxicity of Moringa oleifera and Allamanda cathartica against multiple drug resistant clinical pathogens". International Journal of Pharma and Bio Sciences 4 (2013): 768-775.
- 5. Maya C and Thippanna M. "*In vitro* evaluation of ethano-botanically important plant extracts against early blight disease (*Alternaria Solani*) of tomato". *G.J.B.B* 2.2 (2013): 248-252.
- 6. Hada D and Sharma K. "Assessment of antifungal activity of *cassia fistula* l. fruit pulp against *Alternaria solani*". *International Journal of Pharmacy and Pharmaceutical Sciences* 7.1 (2014): 438-441.
- 7. Audipudi AV and Chakicherla BVS. "Antioxidative and antimicrobial activity of methanol and chloroform extracts of *Gmelina arborea* Roxb". *International Journal of Biochemistry and Biotechnology* 6 (2010): 139-144.
- 8. Ashraf Z., *et al.* "*In Vitro* Antibacterial and Antifungal Activity of Methanol, Chloroform and Aqueous Extracts of Origanum vulgare and Their Comparative Analysis". *International Journal of Organic Chemistry* 1.4 (2011): 257-261.

Citation: Poonam Meena., et al. "Antifungal Potential of Polyalthia longifolia against Rhizoctonia solani". EC Pharmacology and Toxicology 10.4 (2022): 18-27.

- 9. Narayan DP and Purohit SS. "Agro's colour atlas of medicinal plants". India Agrobios 3 (2004).
- 10. Mehta S and Sharma K. "Detection of herbal biocontrol Agents by qualitative methods". *International Journal of Pharma Research and Review* 5 (2016): 12-16.
- 11. Meena P., *et al.* "Antifungal activity of leaf extracts of *polyalthia longifolia* (sonn.) benth. And hook. f. against *rhizoctonia solani*". *Plant Archives* 21.1 (2021): 1366-1371.
- 12. Afrouzan H., *et al.* "Chemical Composition and Antimicrobial Activities of Iranian Propolis". *Iranian Biomedical Journal* 22.1 (2018): 50-65.
- Zaker M. "Natural plant products as eco-friendly fungicides for plant diseases control-A review". *The Agriculturists* 14.1 (2016): 134-141.
- 14. Mehta S and Sharma K. "Control of *Alternaria solani* by herbal Formulation". *Journal of Biologically Active Products from Nature* 8.5 (2018): 319-325.
- 15. Chingwaru C., et al. "Aqueous extracts of Flacourtia indica, Swartzia madagascariensis and Ximenia caffra are strong antibacterial agents against Shigella Spp., Salmonella typhi and Escherichia coli 0157". South African Journal of Botany 128 (2020): 119-127.
- 16. Mehta and Sharma. "Prevention of fungi by natural antimicrobial agents". *Journal of Medical Pharmaceutical and Allied Sciences* 9 (2020): 2428-2436.
- 17. Bhagwat MK and Datar AG. "Antifungal activity of herbal extracts against plant pathogenic fungi". *Archives of Phytopathology and Plant Protection* 47.8 (2013): 959-965.
- 18. Chiamaka R., *et al.* "Characterization and *In vitro* antifungal potential of *Rosmarinus officinalis* and *Eucalyptus globulus* essential oils on phytopathogen *Colletotrichum* sp". *World Journal of Microbiology* 3.1 (2016): 38-42.
- 19. Gavaric N., *et al.* "Chemical profile, antioxidant and antibacterial activity of thyme and oregano essential oils, thymol and carvacrol and their possible synergism". *Journal of Essential Oil-Bearing Plants* 18.4 (2015): 1013-1021.
- 20. Khatoon A., et al. "Studies on in vitro evaluation of antibacterial activities of Eucalyptus globules labill leaf". International Journal of Current Pharmaceutical Research 9.4 (2017): 140-142.
- 21. Yadava RN and Verma V. "A new biologically active flavone glycoside from the seeds of *Cassia Fistula* (Linn.)". *Journal of Asian Natural Products Research* 5.1 (2003): 57-61.
- 22. Harborne JB., et al. "Methods of plant analysis". In phytochemical methods. London, New York: Chapman and hill (1984): 05-06.
- 23. Kokate CK., et al. "Pharmacognogy". In: Analytic pharmacognosy (7th edition), Nirali Prakashan, Pune, (1990): 122-124.
- Groover RK and Moore JD. "Toxicometric studies of Fungicides against the brown root organisms *Sclerotinia fructicola* and *S. laxa*". *Phytopatho*logy 52 (1962): 516-519.
- 25. Collee FG., *et al.* "Test for identification of bacteria". In: Mackie and McCartney Practical Medical Microbiology. Singapore Longman Singapore publishers Ltd. (1996): 131-150.

- 26. Kumar Danendra Hardel and Sahoo Laxmidhar. "A review on phytochemical and Pharmacological of *Eucalyptus globules*. A Multipurpose Tree". *International Journal of Research in Ayurveda and Pharmacy* 2 (2011): 1527-1530.
- 27. Parajuli RR., et al. "Fungitoxicity of the essential oils of Some aromatic plants against Alternaria brassicicola". Scientific World 3 (2005).
- 28. Mahesh B and Satish S. "Antimicrobial activity of some important medicinal plant against plant and human pathogens". *World Journal of Agricultural Sciences* 4 (2008): 839-843.
- 29. Mariri A and Safi M. "*In vitro* antibacterial activity of several plant extracts and oils against some gram-negative bacteria". *Iranian Journal of Medical Sciences* 39.1 (2014): 36-43.
- 30. Bansod S and Rai M. "Antifungal activity of essential oils from Indian medicinal plants against human pathogenic *Aspergillus fumigatus* and *A. niger". World Journal of Modelling and Simulation* 3.2 (2008): 81-88.
- 31. Tyagi AK and Malik A. "Antimicrobial action of essential oil vapors and negative air ions against *pseudomonas fluorescens*". International Journal of Food Microbiology 143.3 (2010): 205-210.
- 32. Selvamohan T., *et al.* "Antimicrobial activity of selected medicinal plants against some selected human pathogenic bacteria". *Advances in Applied Science Research* 3.5 (2012): 3374-3381.
- 33. Goel A and Sharma K. "Effect of Euphorbia Pulcherrima Leaf and Inflorescence Extract on Various Cytomorphological Parameters Aspergillus fumigates: World Academy of Science, Engineering and Technology". International Journal of Biological, Biomolecular, Agricultural, Food and Biotechnological Engineering 7.9 (2011): 516-519.
- 34. Amenu D., *et al.* "Antimicrobial activity of medicinal plant extracts and their synergistic effect on some selected pathogens". *American Journal of Ethnomedicine* 1.1 (2014): 18-29.
- 35. Roozegar MA., et al. "Antimicrobial effect of Pistacia atlantica leaf extract". Bioinformation 12.1 (2016): 19-21.
- 36. Verma SK., et al. "In vitro cytotoxicity of Argemone Mexicana against different human cancer cell line". International Journal of Chemical, Environmental and Pharmaceutical Research 1 (2010): 37-39.
- 37. Saroglou V., *et al.* "Sesquiterpene lactone from *Centaurea spinosa* and their antibacterial and cytotoxic activities". *Journal of Natural Products* 68.9 (2005): 1404-1407.
- 38. Swamy MK., *et al.* "Antimicrobial properties of plant essential oils against human pathogens and their mode of action an updated review". *Evidence-Based Complementary and Alternative Medicine* (2016): 3012462.
- 39. Chouhan S., et al. "Antimicrobial activity of some essential oils present status and future perspectives". Medicines 4.3 (2017): 58.
- 40. Collee FG., *et al.* "Test for identification of bacteria". In: Mackie and McCartney Practical Medical Microbiology. Singapore Longman Singapore publishers Ltd. (1996): 131-150.
- 41. Digrak M., *et al.* "Antibacterial and antifungal effects of various commercial plants extract". *Journal of Pharmaceutical Biology* 37.3 (1999): 216-220.
- 42. Chouhan S., et al. "Antimicrobial activity of some essential oils present status and future perspectives". Medicines 4.3 (2017): 58.
- Altemimi A., et al. "Extraction, Isolation, and Identification of bioactive compounds from plant extracts". Phytochemicals Plants 6.4 (2017): 42.

Citation: Poonam Meena, et al. "Antifungal Potential of Polyalthia longifolia against Rhizoctonia solani". EC Pharmacology and Toxicology 10.4 (2022): 18-27.

- 44. Panda S K., *et al.* "Selective antifungal action of crude extracts of *Cassia fistula* L. A preliminary study on *Candida* and *Aspergillus* species". *Malaysian Journal of Microbiology* 6 (2010): 62-68.
- 45. Velavan S. "Phytochemical Technique-A Review". World Journal of Science and Research 1 (2015): 80-91.
- 46. Handa SS., *et al.* "Extraction Technologies for Medicinal and Aromatic Plants". International centre for science and high technology, Trieste (2008): 21-25.
- 47. Bimakr M., *et al.* "Comparison of different extraction methods for the extraction of major bioactive flavonoid compounds from spearmint (*Mentha spicata* L.) leaves". *Food and Bioproducts Processing* 89.1 (2010): 67-72.
- 48. Mute VM., et al. "Anthelmintic effect of Tamarind indica linn leaves juice extract on Pheretima posthuma". International Journal of Pharmaceutical Research and Development 7 (2009): 1-6.
- 49. Shafiq Y., *et al.* "Assessment of killing kinetics assay and bactericidal mechanism of crude methanolic bark extract of *Casuarina equisetifolia*". *Pakistan Journal of Pharmaceutical Sciences* 31.5 (2018): 2143-2148.

Volume 10 Issue 4 April 2022 © All rights reserved by Poonam Meena., *et al.*