

# Aspects of Radiotherapy and its Impact on Radio Resistance and Cellular Metabolic Activity

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## Abstract

**Purpose:** Metabolic diseases or disorders are widespread, and the causes are numerous. Radiation therapy is one of the popular modalities in current cancer treatment. Although, radiation has some beneficial effects in biological systems, but scientific evidence suggesting that it has a number of harmful effects in our body. The aim of this review is to sketch a current scenario on radio resistance and the effects on radio therapy on metabolic activity.

Materials and Methods: Evidence from the databases PUBMED and SCIENCE DIRECT were considered.

**Results:** Findings suggest that radiation has a dose-dependent effect in our body. Abnormal ROS effects, diverse cellular metabolic activity, apoptosis suppression, alteration of the DNA damage response, and adaptation to hypoxic stress are the most common causes of radio resistance in our body. At high dose radiation can induce excessive ROS and inflammatory phenomena. Moreover, radiation has direct or indirect effects on biological regulators such as signaling proteins, enzymes, hormones, which might be the consequences of radiation-induce harmful effects on the cellular metabolic activities.

**Conclusion:** Radio resistance is a prominent case in radiotherapy. Radiation has beneficial as well as harmful impacts on biological systems.

Keywords: Radiotherapy; Cancer; Combination Therapy; Metabolic Effects

## Abbreviations

ABL1: Abelson Murine Leukemia Viral Oncogene Homolog 1; AMPK: AMP-Activated Kinase; CSF1: Colony Stimulating Factor 1; DSBs: Double-Strand Breaks; GSH: Reduced Glutathione; HIF-1: Hypoxia-Inducible Factor 1; iNOS: Inducible Nitric Oxide Synthase; IR: Ionizing Radiation; MAPK: Mitogen-Activated Protein Kinase; NF- $\kappa$ B: Nuclear Factor Kappa B; NO: Nitric Oxide; O<sub>2</sub><sup>•</sup>: Superoxide Radical; RNS: Reactive Nitrogen Species; ROS: Reactive Oxygen Species; TBI: Total-Body Irradiation; TIGAR: TP53 Induced Glycolysis and Apoptosis Regulator; VEGF: Vascular Endothelial Growth Factor.

## Introduction

Radiotherapy is one of the common strategies of cancer management. Now-a-days, more than 50% of the cancer patients have undergone radiation treatment. However, anatomical changes and tumor regression during radiotherapy may affect in the tumor therapy [1], this leading the understanding of this treatment modality.

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Radiation source is a kind of physical stress which may cause environmental pollution, increases the oxidative pressure and induces further damages such as DNA lesions, cell death, cancer and other diseases [2]. Therefore, radioprotective agents are administered either before or after radiation exposure to minimize radiotoxicity [3]. However, radiation affects biological system within a narrow range. For an example, in a recent study, the TBI at 6 Gy reduced signs of radiation sickness and improved survival rates, while TBI at 7 Gy decreased the rate of survival of the experimental animals [4]. 5,7-bis(methylaminosulfonyl)-8HQ and 8-methoxyquinoline derivatives were evident to show low cytotoxic activity against MOLT-4 cells [5], while some aminothiol derivatives were found to exert systemic toxicities [6]. On the other hand, radioiodine 131 is associated with side effects such as salivary gland dysfunction and an increased risk of leukemia [7].

Radiotherapy, depending upon the dose, has been reported to have different effects on tumor infiltrating myelomonocytic cells [8,9]. Ionizing radiation (IR) at high dose may cause elicit regenerative responses, thus the tumor regrowth. The CXCR4-CXCL12 chemokine axis has been implicated in the recruitment of tumor promoting myeloid cells after local irradiation [10]. Irradiation caused nuclear translocation of the damage-activated kinase Abelson murine leukemia viral oncogene homolog 1 (ABL1), which enhanced colony stimulating factor 1 (CSF1) gene transcription [11]. This review focuses on the occurrence of radioresistance and its impacts on metabolic activity.

#### **Radiotherapy and radio resistance**

Radiotherapy is also evident to activate new subnetworks, resulting in network flexibility [12]. IR exerts effects on whole cell components [13], thus, a dilution effect of the non-hub elements by IR. Chronic exposure to this low level of IR may cause greater damage [14]. Hubs that remain undamaged could eventually activate other radiation-responsive signaling networks, reintegrate network topologies and establish networks more resistant to radioresistance [15]. Irradiation-induced reactive oxygen species (ROS) can upregulate nuclear factor kappa B (NF- $\kappa$ B)-mediated inflammatory signaling cascade in cancer cells [16]. Possible pathways of the occurrence of radioresistance have been shown in figure 1. Moreover, activated inflammatory network (e.g. chemokines and cytokines) may result cancer cells survival by overcoming IR-induced inflammatory stress. It may result in acquired radioresistance and consequently reduced efficacy of radiotherapy in patients. However, radioresistance is more prominent in older patients than the youngs [17]. It may be due to higher ROS (e.g. NO and  $O_2^{\bullet}$ ) production rate in the former category patients.





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**Figure 1:** Mechanism of radioresistance in mammalian cells. [(a) Ionizing radiation (IR)-induced reactive oxygen species (ROS) and mitochondrial membrane potential (MMP) dependent radioresistance pathway. (b) IR-induced miRs, especially miR21 and cyclin D1 dependent cell survival. (c) IR-induced hypoxia inducible factor 1 and miR451 dependent radioresistance. (d) Irradiation effects in tumor cell growth. Other abbreviations  $\rightarrow$  SOD: superoxide dismutase; Cyt-c: Cytochrome c; GSK3 $\beta$ : Glycogen Synthase Kinase 3 Beta; NHEJ: Non-Homologous End Joining; HRR: Homologous End Joining; DSBs: Double Strand Breaks.

Conventional radiotherapy doses up to 70 Gy is evident to show biochemical failure rates of 30% or more in localized disease [18]. In a recent study, resveratrol (an antioxidant) was founded to promote IR effects in prostate cancer (PrCa) cells, possibly *via* mitogen-activated protein kinase (MAPK)-Akt-dependent pathway [19]. Antioxidants such as polyphenol chemical class, xanthine derivatives, tocopherol, sucralfate, and ascorbate have been evident to alter cytokine release affecting cutaneous and systemic changes in experimental animals [20]. In a recent study, it has been demonstrated that medicinal plants and their active constituents, especially plant-derived antioxidants may alleviate radiation-induced damage through antioxidant, anti-inflammatory, wound healing and immunostimulatory properties [21].

Certain chemical can cause mutation which is one of the major consequences of radioresistance. For an example, in a study trichloroethylene was found to cause mutation in P81S VHL, thus leading to diversify metabolic activity, apoptosis suppression, and alteration of the DNA damage response by IR therapy [22]. Moreover, exposure of tumor cells to radiation could impact a large number of proteins simultaneously, leading to modify chemotherapeutic agent's action [23].

In the development of tumor cancer cells partially undergo a hypoxic condition due to insufficient vasculature systems. These cells show hyper-activation of hypoxic-inducible factor (HIF)-1 network for adaptation to hypoxic stress. This also leads an IR resistance [24], possibly *via* increasing in vascular endothelial growth factor (VEGF) expression [25]. Early reports suggest that bevacizumab, one of the best-known VEGF inhibitor can normalize tumor vasculature, overcome resistance to radiation, inhibit repopulation after radiation and blockade of radiation-induced increased VEGF expression [26], therefore should be effective with an acceptable toxicity profile [27]. However, autophagy activation can promote bevacizumab resistance in some cancer cells [28].

Moreover, protected tumor vasculature can supply required oxygen and nutrients to the tumor cells, which may lead to grow tumor radioresistance and tumor growth progression. However, Sharma and Jain [29] suggested that radioresistance of hypoxic cells can be overcome by the combined therapy with the radio modifiers such as hematoporphyrin derivative (Hpd) and 2-deoxy-D-glucose (2-DG).

#### Radiation impacts on metabolic activity

Exposure of IR has some beneficial impacts, especially in the synthesis of vitamin  $D_3$  and its various biological roles in our body [30] (Figure 2).



ray, which is helpful for bone mineralization, neuromuscular function and varieties of metabolic activities.

However, it is also evident that IR has a variety of adverse effects on the immune system as it can affect the key metabolic processes such as glucose uptake, glycolysis, and energy metabolism [31]. Uptake of the radiopharmaceuticals is increased by the tumor cells than the normal cells [32], which may be a reason of effectivity of iodine-125 and IR combination therapy of certain malignant diseases [33].

At low doses ( $\leq$  1.25 Gy) IR inhibits the inducible nitric oxide synthase (iNOS) pathways (e.g. translational and post-translational). However, an increased degradation of heme is essential for iNOS activity, which demonstrating other post-translational modifications may be involved in the proteasome degradation pathway [34]. However, Hildebrandt., *et al.* [35] reported that IR up to 10 Gy did not stimulate macrophages, changes metabolic activity, cell proliferation, and reproductive integrity other than an increased in nitric oxide (NO) pro-

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duction. Furthermore, irradiation of autologous blood by UV rays in 81 patients with proinflammatory diseases is evident to modulate indices of antimicrobial protection, increase in the intensity of the histochemical reaction to peroxidase, reduction of pH in the neutrophil phagosomes, restore the serotonin, increased of the intensity of metabolic processes, changes in membrane phospholipid composition, and juvenile platelet forms [36]. Low dose irradiation (0.5-6 Gy) was also found to reprogram macrophage function, normalization of the tumor vasculature and more efficient recruitment of specific antitumor T cells, possibly *via* endothelial cell activation and down-regulation of immunosuppressive tumor promoting functions [37].

On the other hand, IR at high dose (4-8 Gy) induces cell impairment. Irradiation within a short period may upregulate TP53 induced glycolysis and apoptosis regulator (TIGAR) that lowers fructose-2,6-bisphosphate levels in cells, consequently decreases glycolysis and lead to the scavenge of ROS. The ultimate result is the highest resistance to cell death [38]. Generally, the high glycolytic rate protects cancer cells from ROS-induced DNA damage by supplying large amounts of reducing equivalents such as pyruvate, lactate, glutathione, and NADPH that scavenge ROS [39]. Ketone bodies and fatty acids may inhibit glycolysis [40]. An impairment of glycolysis causes mito-chondrial dysfunction, which increases excessive ROS production, thus leading cell death [41]. Moreover, oxidation of ketone bodies in peripheral tissue decreases the NADP<sup>+</sup>/NADPH ratio, thus increases the levels of the important physiological antioxidant enzyme, reduced glutathione (GSH) [42]. Vlashi., *et al.* [43] suggests that cancer stem cells possess high metabolic flexibility, which is the major cause of radioresistance in these types of cells. Effects of various doses of radiation in test systems have been shown in table 1.

Doses	Test systems	Health effects	Mechanisms	References
100 mGy	Mice	Brain damage and impaired cognition	DNA damage, inflammation, vascular damage, white matter injury and coagulation necrosis	Lowe and Wyrobek [44]
0-0.20 Gy	Human	Breast cancer	DNA damage, oxidative stress, chromosomal aberrations	Ozasa. <i>, et al</i> . [45]
50 mGy	Huamn	Leukemia	DNA damage	Pearce., <i>et al</i> . [46]
0.05 or 0.30 Gy	Mice	Malformation	DNA damage, global genome DNA methylation, chromosomal aberrations	Wang. <i>, et al</i> . [47]
> 2 Gy	Human	Cataracts	DNA damage, inflammation	Fujimichi and Hamada [48]
1 - 3 Gy	Rats	CVD	Endothelial dysfunction, inflammation, oxidative stress, alterations in coagulation and platelet activity, DNA damage, senescence and cell death	Baselet. <i>, et al</i> . [49]
60 mGy	Huaman	Brain tumors	DNA damage, chromosomal aberrations	Kutanzi., <i>et al</i> . [50]
-	Human	Thyroid cancer	DNA damage, chromosomal aberrations	Philchenkov and Balcer- Kubiczek [51]

#### Table 1: Effects of radiation in some test systems.

IR at 0 to 8 Gy is evident to phosphorylate and activate the metabolic sensor and tumor suppressor AMP-activated kinase (AMPK), possibly *via* p53 and cyclin-dependent kinase inhibitors p21<sup>cip1</sup> and p27<sup>kip1</sup> inhibit induction of nuclear fragmentation and cleavage of caspase 3 [52]. On the other hand, IR at 50 to 70 Gy may cause acute and late grade > or = 2 xerostomia and mucositis. In a study, amifostine (WR-2721), a cytoprotective agent was found to show protective effects of advanced ovarian cancer cells by conversion and uptake of the active metabolite, WR-1065 [53]. Effects of chronic and/or high IR dose in biological system has been shown in figure 3.

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Total-body irradiation (TBI) is evident to cause morphological changes in different organs in rats with differential expression of 53% (765 genes) that were mainly involved in a total of 21 pathways, including metabolic, cancer, and MAPK pathways [4].

IR generates intermediate free radicals and ROS leading to DNA double-strand breaks (DSBs) [54]. Delay or an impairment of the cellular repair system of this type of injury directly or indirectly causes mutation or even cell death. Furthermore, tumor cells can survive through activation of the DSB repair pathway, including homologous recombination and nonhomologous endjoining [55,56], this may lead to growing radioresistance [57]. Mainly the exposure of IR modulates numerous cellular networks, including DNA repair, survival, apoptosis, cell cycle, cell migration, protein localization, RNA processing, antioxidant defense, inflammation and cell proliferation, and so on. For example, p53-related genes and DNA-damage response genes are generally activated by irradiation in lung cancer cells [58].

In a study, a significant reduction of Cho/Cr and NAA/Cr ratios was observed with gamma knife radiosurgery performed in 18 metastatic brain tumor patients [59]. A decrease in Cho/Cr ratio reflects inhibition of proliferative activity and early apoptotic cell loss, while the reduction of NAA/Cr is associated with the radiation-induced modulation of neuronal activity. Petersen., *et al.* [60] suggests that cyclooxygenase-2 and its products may act as protectors against cell damage by ionizing radiation. In another study, IR at 25 cGy was found to modulate neurotransmission processes *via* metabotropic muscarinic ChR and gamma amino butyric acid receptor pathways [61]. Radioimmunotherapy using 131I-UJ13A or 131I-3F8 monoclonal antibodies have been evident to show moderate response and considerable side effects in IV neuroblastoma patients, probably *via* enhancing tumor uptake by modulation of antigen expression or by increasing the tumor perfusion/vascularity/permeability [62]. IR-induced damaging effects on our body have been shown in figure 4.



**Figure 4:** IR exposure and changes of biological phenomena. [IR-induced Ca=2 efflux and excessive production of reactive oxygen species (ROS) alters normal biological activities. ROS increased stress kinases alters the secretion of basic fibroblast growth factor (bFGF), which causes infertility in male by the destruction of blood-testis barrier.

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### Conclusion

The biological effects of radiation are dose-dependent. At low dose it is beneficial for us, however, chronic exposure to radiation at any dose is evident to produce various abnormalities in our body. Metabolic abnormalities are one of them. Radiation can act by the induction of ROS, inflammatory phenomena, regulation of cell signalling cascades, and so on. Radioresistance is evident through a number of ways, including alteration of cellular metabolic activities, suppression of apoptosis, alteration of the DNA damage response, and adaptation of the hypoxic stress. Chronic and at high IR doses directly and/or indirectly can induce oxidative stress and inflammatory responses. Furthermore, radiation can alter cell signalling cascades and other biochemical activities, thus the harmful effects of it on the metabolic activities in a biological system.

## Bibliography

- 1. Bibault J-E., et al. "Adaptive radiation therapy for non-small cell lung cancer". Cancer/Radiothérapie 19.6-7 (2015): 458-462.
- Moores BM and Regulla D. "A review of the scientific basis for radiation protection of the patient". *Radiation Protection Dosimetry* 147.1-2 (2011): 22-29.
- 3. Hosseinimehr SJ. "Trends in the development of radioprotective agents". Drug Discovery Today 12.19-20 (2007): 794-805.
- Li J., et al. "MASM, a Matrine Derivative, Offers Radioprotection by Modulating Lethal Total-Body Irradiation-Induced Multiple Signaling Pathways in Wistar Rats". Molecules 21.5 (2016): E649.
- Ariyasu S., et al. "Design and synthesis of 8-hydroxyquinoline-based radioprotective agents". Bioorganic and Medicinal Chemistry 22.15 (2014): 3891-3905.
- Copp RR., et al. "Radioprotective efficacy and toxicity of a new family of aminothiol analogs". International Journal of Radiation Biology 89.7 (2013): 485-442.
- Noaparast Z and Hosseinimehr SJ. "Radioprotective agents for the prevention of side effects induced by radioiodine-131 therapy". *Future Oncology* 9.8 (2013): 1145-1159.
- 8. De Palma M., *et al.* "A new twist on radiation oncology: low-dose irradiation elicits immunostimulatory macrophages that unlock barriers to tumor immunotherapy". *Cancer Cell* 24.5 (2013): 559-561.
- Russell JS and Brown JM. "The irradiated tumor microenvironment: role of tumor-associated macrophages in vascular recovery". *Frontiers in Physiology* 4 (2013): 157.
- 10. Kozin SV., et al. "Recruitment of myeloid but not endothelial precursor cells facilitates tumor regrowth after local irradiation". Cancer Research 70.14 (2010): 5679-5685.
- 11. Xu J., et al. "CSF1R signaling blockade stanches tumor-infiltrating myeloid cells and improves the efficacy of radiotherapy in prostate cancer". Cancer Research 73.9 (2013): 2782-2794.
- Rashi-Elkeles S., *et al.* "Transcriptional modulation induced by ionizing radiation: p53 remains a central player". *Molecular Oncology* 5.4 (2011): 336-348.
- 13. Somosy Z. "Radiation response of cell organelles". Micron 31.2 (2000): 165-181.
- 14. Albert R., et al. "Error and attack tolerance of complex networks". Nature 406 (2000): 378-382.
- 15. Lee SY., et al. "Identifying genes related to radiation resistance in oral squamous cell carcinoma cell lines". International Journal of Oral and Maxillofacial Surgery 42.2 (2013): 169-176.

#### Aspects of Radiotherapy and its Impact on Radio Resistance and Cellular Metabolic Activity

- 16. Multhoff G and Radons J. "Radiation, inflammation, and immune responses in cancer". Frontiers in Oncology 2 (2012): 58.
- 17. Tasat DR., et al. "Radiation effects on oxidative metabolism in young and aged rat alveolar macrophages". Cellular and Molecular Biology 48.5 (2002): 529-535.
- Zietman AL., *et al.* "Randomized trial comparing conventional-dose with high-dose conformal radiation therapy in early-stage adenocarcinoma of the prostate: long-term results from proton radiation oncology group/American college of radiology 95-09". *Journal of Clinical Oncology* 28.7 (2010): 1106-1111.
- 19. Rashid A., *et al.* "Resveratrol enhances prostate cancer cell response to ionizing radiation. Modulation of the AMPK, Akt and mTOR pathways". *Radiation Oncology* 6 (2011): 144.
- 20. Amber KT., et al. "The use of antioxidants in radiotherapy-induced skin toxicity". Integrative Cancer Therapies 13.1 (2014): 38-45.
- Kim W., et al. "Effects of traditional oriental medicines as anti-cytotoxic agents in radiotherapy". Oncology Letters 13.6 (2017): 4593-4601.
- 22. DeSimone MC., *et al.* "Pleiotropic Effects of the Trichloroethylene-Associated P81S VHL Mutation on Metabolism, Apoptosis, and ATM-Mediated DNA Damage Response". *Journal of the National Cancer Institute* 105.18 (2013): 1355-1364.
- 23. Seo H., et al. "Network-based approaches for anticancer therapy (Review)". International Journal of Oncology 43.6 (2013): 1737-1744.
- 24. Brown JM and Wilson WR. "Exploiting tumour hypoxia in cancer treatment". Nature Reviews Cancer 4.6 (2004): 437-447.
- 25. Moeller BJ and Dewhirst MW. "HIF-1 and tumour radiosensitivity". British Journal of Cancer 95.1 (2006): 1-5.
- Zhuang HQ., et al. "Research progress on the mechanisms of combined bevacizumab and radiotherapy". Recent Patents on Anti-Cancer Drug Discovery 9 (2014): 129-134.
- 27. Akiyama Y., *et al.* "Advantages and Disadvantages of Combined Chemotherapy with Carmustine Wafer and bevacizumab in Patients with Newly Diagnosed Glioblastoma: A Single-Institutional Experience". *World Neurosurgery* 113 (2018): e508-e514.
- Huang H., et al. "Autophagy activation promotes bevacizumab resistance in glioblastoma by suppressing Akt/mTOR signaling pathway". Oncology Letters 15.2 (2018): 1487-1494.
- Sharma RK and Jain V. "Tackling radioresistance of hypoxic cells by metabolic modulation of bioenergetics--a 31P MRS study on perfused Ehrlich ascites tumor cells". Indian Journal of Physiology and Pharmacology 46.1 (2002): 51-60.
- 30. Islam MT. "Beneficial aspects of ultraviolet rays in protective and sound health". EC Pharmacology and Toxicology 6.2 (2018): 57-65.
- Li H-H., et al. "Ionizing Radiation Impairs T Cell Activation by Affecting Metabolic Reprogramming". International Journal of Biological Sciences 11.7 (2015): 726-736.
- Bodei L., et al. "Radionuclide Therapy with Iodine-125 and Other Auger-Electron-Emitting Radionuclides: Experimental Models and Clinical Applications". Cancer Biotherapy and Radiopharmaceuticals 18.6 (2003): 861-877.
- Kassis AI., et al. "In vivo therapy of neoplastic meningitis with methotrexate and 5-[1251]iodo-29-deoxyuridine". Acta Oncologica 39.6 (2000): 731-737.
- Hildebrandt G., et al. "Inhibition of the iNOS Pathway in Inflammatory Macrophages by Low-Dose X-Irradiation In Vitro". Strahlentherapie und Onkologie 179.3 (2003): 158-166.

- 35. Hildebrandt G., *et al.* "Mechanisms of the anti-inflammatory activity of low-dose radiation therapy". *International Journal of Radiation Biology* 74.3 (1998): 367-378.
- 36. Piksin IN., et al. "Ultraviolet irradiation of blood in surgery". Khirurgiia (Mosk) 11 (1990): 100-104.
- 37. Klug F., *et al.* "Low-dose irradiation programs macrophage differentiation to an iNOS(+)/M1 phenotype that orchestrates effective T cell immunotherapy". *Cancer Cell* 24.5 (2013): 589-602.
- 38. Peña-Rico MA., *et al.* "TP53 induced glycolysis and apoptosis regulator (TIGAR) knockdown results in radiosensitization of glioma cells". *Radiotherapy and Oncology* 101.1 (2011): 132-139.
- 39. Meijer TWH., *et al.* "Targeting hypoxia, HIF-1, and tumor glucose metabolism to improve radiotherapy efficacy". *Clinical Cancer Research* 18.20 (2012): 5585-5594.
- 40. Cullingford TE. "The ketogenic diet fatty acids, fatty acid activated receptors and neurological disorders". *Prostaglandins, Leukotrienes and Essential Fatty Acids* 70.3 (2004): 253-264.
- 41. Allen BG., *et al.* "Ketogenic diets enhance oxidative stress and radio-chemo-therapy responses in lung cancer xenografts". *Clinical Cancer Research* 19.14 (2013): 3905-3913.
- Veech RL. "The therapeutic implications of ketone bodies: the effects of ketone bodies in pathological conditions: ketosis, ketogenic diet, redox states, insulin resistance, and mitochondrial metabolism". *Prostaglandins, Leukotrienes and Essential Fatty Acids* 70.3 (2004): 309-319.
- 43. Vlashi E., et al. "Metabolic state of glioma stem cells and nontumorigenic cells". Proceedings of the National Academy of Sciences 108.38 (2011): 16062-16067.
- 44. Lowe X and Wyrobek A. "Characterization of the early CNS stress biomarkers and profiles associated with neuropsychiatric diseases". *Current Genomics* 13.6 (2012): 489-497.
- 45. Ozasa K., *et al.* "Studies of the mortality of atomic bomb survivors, report 14, 1950–2003: an overview of cancer and noncancer disease". *Radiation Research* 177.3 (2012): 229-243.
- 46. Pearce MS., et al. "Radiation exposure from CT scans in childhood and subsequent risk of leukaemia and brain tumours: a retrospective cohort study". *Lancet* 380.9840 (2012): 499-505.
- 47. Wang B., *et al.* "Adaptive response of low linear energy transfer Xrays for protection against high linear energy transfer accelerated heavy ion-induced teratogenesis". *Birth Defects Research Part B Developmental and Reproductive Toxicology* 95.6 (2012): 379-385.
- 48. Fujimichi Y and Hamada N. "Ionizing irradiation not only inactivates clonogenic potential in primary normal human diploid lens epithelial cells but also stimulates cell proliferation in a subset of this population". *PLoS One* 9.5 (2014): e98154.
- 49. Baselet B., et al. "Cardiovascular diseases related to ionizing radiation: the risk of low-dose exposure (review)". International Journal of Molecular Medicine 38.6 (2016): 1623-1641.
- 50. Kutanzi KR., *et al.* "Pediatric exposures to ionizing radiation: carcinogenic considerations". *International Journal of Environmental Research and Public Health* 13.11 (2016): E1057.
- Philchenkov AA and Balcer-Kubiczek EK. "Molecular markers of apoptosis in cancer patients exposed to ionizing radiation: the post-Chornobyl view". Experimental Oncology 38.4 (2016): 224-237.
- 52. Sanli T., et al. "Lovastatin Sensitizes Lung Cancer Cells to Ionizing Radiation". Journal of Thoracic Oncology 6.3 (2011): 439-450.

## Aspects of Radiotherapy and its Impact on Radio Resistance and Cellular Metabolic Activity

- 53. Culy CR and Spencer CM. "Amifostine: an update on its clinical status as a cytoprotectant in patients with cancer receiving chemotherapy or radiotherapy and its potential therapeutic application in myelodysplastic syndrome". *Drugs* 61.5 (2001): 641-684.
- 54. Li L., *et al.* "Cellular responses to ionizing radiation damage". *International Journal of Radiation Oncology* \* *Biology* \* *Physics* 49.4 (2001): 1157-1162.
- 55. Sonoda E., *et al.* "Differential usage of non-homologous end-joining and homologous recombination in double strand break repair". *DNA Repair* 5.9-10 (2006): 1021-1029.
- 56. Li YH., *et al.* "Inhibition of non-homologous end joining repair impairs pancreatic cancer growth and enhances radiation response". *PLoS One* 7.6 (2012): e39588.
- 57. Lee YS., *et al.* "Differential gene expression profiles of radioresistant non-small-cell lung cancer cell lines established by fractionated irradiation: tumor protein p53-inducible protein 3 confers sensitivity to ionizing radiation". *International Journal of Radiation Oncology \* Biology \* Physics* 77.3 (2010): 858-866.
- 58. Xu QY., *et al.* "Identification of differential gene expression profiles of radioresistant lung cancer cell line established by fractionated ionizing radiation in vitro". *Chinese Medical Journal* 121.18 (2008): 1830-1837.
- 59. Chernov MF, et al. "Early metabolic changes in metastatic brain tumors after Gamma Knife radiosurgery: 1H-MRS study". Brain Tumor Pathology 21.2 (2004): 63-67.
- 60. Petersen C., *et al.* "New targets for the modulation of radiation response--selective inhibition of the enzyme cyclooxygenase 2". *Current Medicinal Chemistry Anti-Cancer Agents* 3.5 (2003): 354-359.
- 61. Anan'eva TV and Dvoretskiĭ AI. "Pharmacological approaches to study of choline- and GABA-receptor states in neuronal membranes after low doses irradiation". *Radiatsionnaia Biologiia, Radioecologiia* 40.1 (2000): 76-80.
- 62. Hoefnagel CA. "Nuclear medicine therapy of neuroblastoma". Quarterly Journal of Nuclear Medicine 43.4 (1999): 336-343.

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