

Antigenic Determinants of Utrophin from *Dracunculus Medinensis*: New Approach for Synthetic Peptide Vaccines

Sonu Mishra* and Virendra Gomase

Department of Biotechnology, Mewar University, Chittorgarh, Rajasthan, India

*Corresponding Author: Sonu Mishra, Department of Biotechnology, Mewar University, Chittorgarh, Rajasthan, India.

Received: December 09, 2016; Published: December 14, 2016

Abstract

The synthetic peptide vaccines idea is usually based on identification and chemical synthesis of B-cell and T-cell epitopes which are immunodominant and can induce specific immune responses. Deeper knowledge of antigens of utrophin from *Dracunculus medinensis*, mechanisms of immune response and the development of effective and safe adjuvants give hope that the effective peptide vaccines will be developed in the future. These disadvantages led to the development of subunit vaccines, including synthetic peptides as antigen, which consist of a specific part of the whole antigen which has been demonstrated to stimulate an immune response by eliciting antibodies that neutralize the biological activity of proteins.

Keywords: Utrophin; Peptide; Antigenicity

Introduction

Utrophin from *Dracunculus medinensis* are characterized to envision the antigenicity and solvent accessible region that allows potential drug targets to identify active sites against various reactions. Prediction of antigenicity predicts those segments within utrophin that are antigenic by eliciting an antibody response [1-10]. Antigenic peptides should be located in solvent accessible regions and contain each hydrophobic and hydrophilic residue which believed that majority surface exposed regions of a protein are potential antigenic. Prediction of peptides those are in the N- and C-terminal region of the protein, because the N- and C-terminal regions of proteins are usually solvent accessible and unstructured hence Antibodies against those regions are also likely to recognize the native protein that can help to design of synthetic peptide vaccine and immuno-diagnostic reagents [10-25].

Material and Methods

Antigenic epitopes are determined by exploitation the Hopp and Woods, Welling, Parker, Kolaskar and Tongaonkar antigenicity method and Bepipred Epitope Prediction [16,26-29]. Predictions are on the basis of supported plots that ensure the prevalence of amino acid residues in experimentation notable segmental epitopes [30-34].

Results and Interpretation

Protein Sequence

utrophin, partial [*Dracunculus medinensis*]

DVEVVKAQFKEHEQFMQSLTESQDSVGRVLHRGNVICQLDDEQNMSLLSQLKLV
NAKWERVREIAMNRQ NLLLEKLNLSLQIQLKKL

Theoretical pI: 8.11

Molecular weight: 10330.9

Number of amino acids: 88

Hopp and Woods antigenicity methods

Hopp and woods method predicts antigenic determinants by searching protein sequences of utrophin from *Dracunculus medinensis* to find the area of greatest local hydrophilicity and the hydrophilic regions in the protein are located on the surface and are potentially antigenic. The point of highest local average hydrophilicity is located in or adjacent to an antigenic determinant. In this scale the amino acid value is starting from -3 (most hydrophobic) to 3 (most hydrophilic).

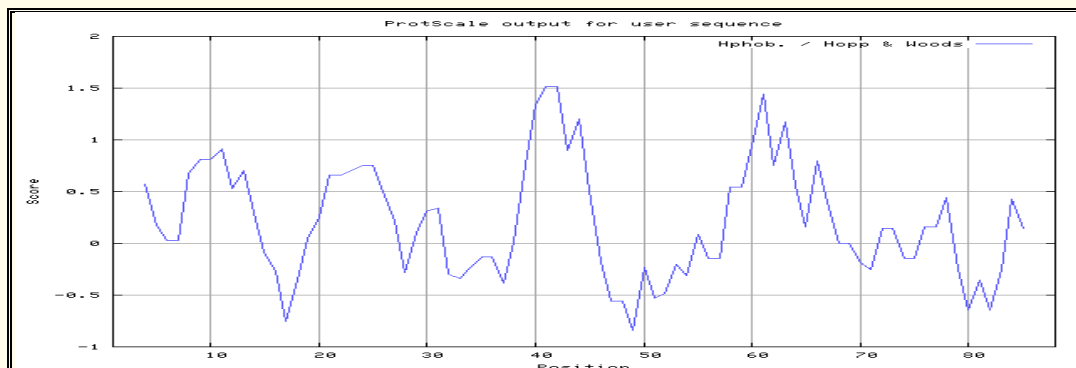


Figure 1: Hopp and Woods antigenicity plot of utrophin from *Dracunculus medinensis*. In this scale the amino acid value is starting from -3 are consider (most hydrophobic) to 3 (most hydrophilic). The values greater than 0 are consider to be hydrophilic that are exposed on the surface of the folded protein.

Min: -0.843, Max: 1.514 score at (position 48-50, 40-42) window-7

Welling antigenicity methods

Welling antigenicity method is based on the percentage of each amino acid present in known antigenic regions (utrophin from *Dracunculus medinensis*) compared to the percentage of the amino acids in the average composition of a protein. Previous strategies are based on the assumption that antigenic regions are primarily hydrophilic at the surface of the protein. This method is better than the Hopp-Woods scale of hydrophobicity which is also used to identify antigenic regions.

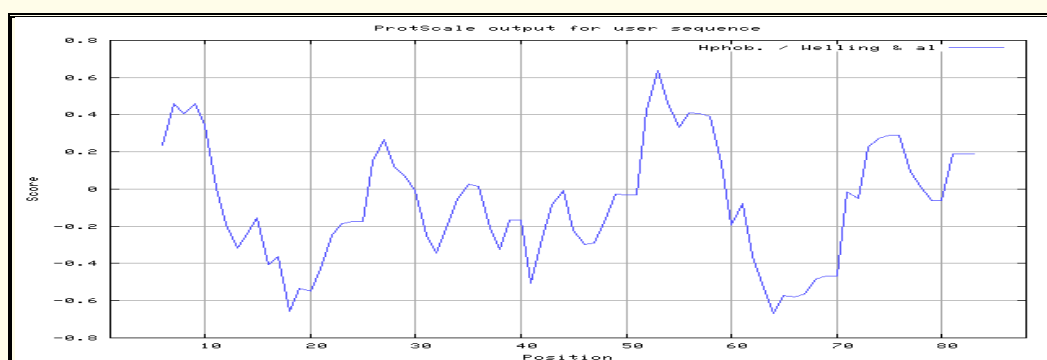


Figure 2: Welling antigenicity plot of utrophin from *Dracunculus medinensis*. The Max: 0.636 score which was shown at the Position: 53.

Min: -0.667, Max: Score: 0.636 (max) score at (position 63-65, 51-55) window-11

Parker Hydrophilicity Prediction

Parker scale predicts antigenicity by identifying regions of greatest native hydrophilicity of utrophin from *Dracunculus medinensis*. It was derived from the Hopp-Woods scale however, these uses the HPLC retention times of model peptides to determine hydrophilicity. Parker hydrophilicity scale is sequence-based method that has been shown recently to perform prediction of linear epitopes of utrophin from *Dracunculus medinensis*.

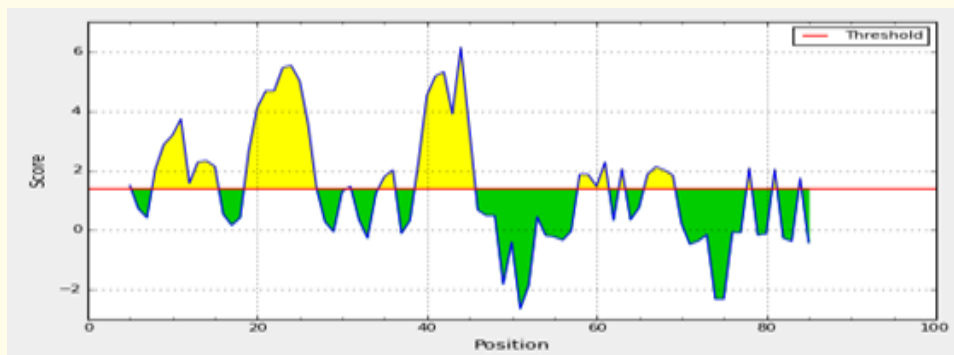


Figure 3: Parker antigenicity plot of utrophin from *Dracunculus medinensis* hydrophilic scale based (threshold setting = 1.396). Parker antigenicity scale predicted 88 length peptides this scale predicted maximum score at position 41- DDEQNMS -47, under the threshold value 1.396.

Center position: 4; Window size: 7; Threshold: 1.396; Average: 1.396, Minimum: -2.657, Maximum: 6.157

| Position | Residue | Start | End | Peptide | Score |
|----------|---------|-------|-----|---------|-----------------|
| 36 | I | 33 | 39 | GNVICQK | 2.014 |
| 37 | C | 34 | 40 | NVICQKL | -0.114 |
| 38 | Q | 35 | 41 | VICQKLD | 0.314 |
| 39 | K | 36 | 42 | ICQKLDD | 2.271 |
| 40 | L | 37 | 43 | CQKLDDE | 4.529 |
| 41 | D | 38 | 44 | QKLDDEQ | 5.186 |
| 42 | D | 39 | 45 | KLDDEQN | 5.329 |
| 43 | E | 40 | 46 | LDDEQNM | 3.914 |
| 44 | Q | 41 | 47 | DDEQNMS | 6.157(Maximum) |
| 45 | N | 42 | 48 | DEQNMSL | 3.414 |
| 46 | M | 43 | 49 | EQNMSLL | 0.671 |
| 47 | S | 44 | 50 | QNMSLLS | 0.486 |
| 48 | L | 45 | 51 | NMSLLSQ | 0.486 |
| 49 | L | 46 | 52 | MSLLSQL | -1.829 |
| 50 | S | 47 | 53 | SLLSQLK | -0.414 |
| 51 | Q | 48 | 54 | LLSQLKL | -2.657(Minimum) |
| 52 | L | 49 | 55 | LSQLKLV | -1.871 |

| | | | | | |
|----|---|----|----|---------|--------|
| 53 | K | 50 | 56 | SQLKLVN | 0.443 |
| 54 | L | 51 | 57 | QLKLVNA | -0.186 |
| 55 | V | 52 | 58 | LKLVNAK | -0.229 |

Table 1: Parker Hydrophilicity Prediction Result Data.

Average: 1.396 Minimum: -2.657 Maximum: 6.157

Predicted residue scores

Kolaskar & Tongaonkar Antigenicity

Kolaskar & Tongaonkar Antigenicity is a semi-empirical method for the prediction of antigenic regions including information of surface accessibility and flexibility. The method was able to predict antigenic determinants with an accuracy of 75%.

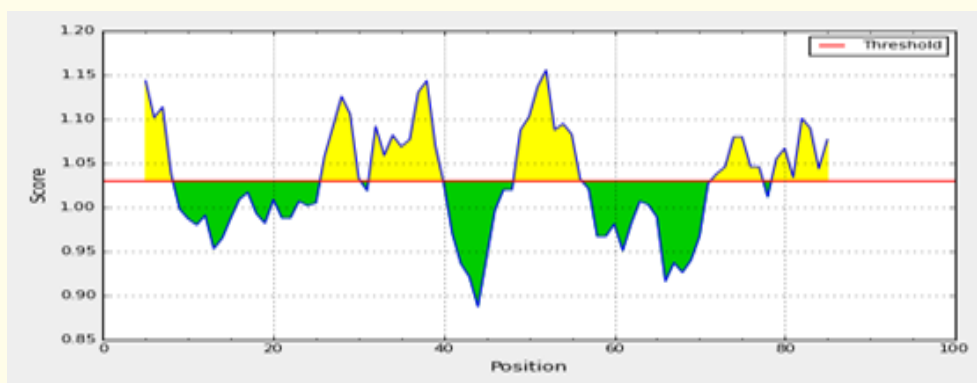


Figure 4: Kolaskar and Tongaonkar antigenicity prediction plot of utrophin from *Dracunculus medinensis* predicts those segments among a protein sequence that are to be antigenic by eliciting an antibody response (threshold setting = 1.030). This scale predicted peptide in the positions 49-LSQLKLV-55.

Center position: 4; Window size: 7; Threshold: 1.030.

| No. | Start Position | End Position | Peptide | Peptide Length |
|-----|----------------|--------------|---------|----------------|
| 1 | 49 | 55 | LSQLKLV | 7 |

Table 2: Predicted peptides of utrophin from *Dracunculus medinensis*.

| Position | Residue | Start | End | Peptide | Score |
|----------|---------|-------|-----|---------|----------------|
| 38 | Q | 35 | 41 | VICQKLD | 1.144 |
| 39 | K | 36 | 42 | ICQKLDD | 1.07 |
| 40 | L | 37 | 43 | CQKLDDE | 1.027 |
| 41 | D | 38 | 44 | QKLDDEQ | 0.97 |
| 42 | D | 39 | 45 | KLDDEQN | 0.936 |
| 43 | E | 40 | 46 | LDDEQNM | 0.921 |
| 44 | Q | 41 | 47 | DDEQNMS | 0.887(Minimum) |
| 45 | N | 42 | 48 | DEQNMSL | 0.942 |
| 46 | M | 43 | 49 | EQNMSLL | 0.997 |

| | | | | | |
|----|---|----|----|---------|----------------|
| 47 | S | 44 | 50 | QNMSLLS | 1.02 |
| 48 | L | 45 | 51 | NMSLLSQ | 1.02 |
| 49 | L | 46 | 52 | MSLLSQL | 1.088 |
| 50 | S | 47 | 53 | SLLSQLK | 1.103 |
| 51 | Q | 48 | 54 | LLSQLKL | 1.137 |
| 52 | L | 49 | 55 | LSQLKLV | 1.156(Maximum) |
| 53 | K | 50 | 56 | SQLKLVN | 1.088 |
| 54 | L | 51 | 57 | QLKLVNA | 1.095 |
| 55 | V | 52 | 58 | LKLVNAK | 1.083 |

Table 3: Kolaskar & Tongaonkar Antigenicity Result Data.

Average: 1.030 Minimum: 0.887 Maximum: 1.156

Discussion

Antigenic determinants of utrophin from *Dracunculus medinensis* are determined by finding the area of greatest local hydrophilicity using the Hopp-Woods method. This method has a high success rate than other methods. The success of this method is its cautious approach to charge interactions that gives equal weight to positive and negative charged residues, whereas other methods tend to favor one or the other. The sites chosen by this method is to be highly exposed and charged regions of the protein's surface therefore, have ample opportunity to contact other proteins. Here we found high peaks at position: 41 with max score: 1.514 and the minimum found at the position: 49 with minimum score: -0.843 using window-7. Welling Method used to locate hydrophilic regions in a protein since, it is assumed that antigenic determinants are located on surface which contain charged and polar residues. These methods are used to obtain a rough idea for estimation of potentially antigenic regions. However, as shown by Hopp and Woods not all antigenic regions are hydrophilic and not all hydrophilic regions are antigenic. Therefore, welling developed a method based on the percentage of each amino acid present in known antigenic determinants compared with the percentage of the amino acids in the average composition of a protein. Here we found the Position: 53 with Score: 0.636 (max) and at Position: 64 Score: -0.667 (min) by using window-11. Parker used three parameters - hydrophilicity, accessibility and flexibility to calculate the antigenic propensity using a composite plot. This method has improved to predict antigenic determinants as compared to Hopp and Woods' method. Parker antigenicity plot is based on (threshold setting = 1.396) and this scale predicted maximum score at position: 44(residue: Q) 41-DDEQNMS-47 with the score: 6.157 (Maximum) under the threshold value 1.396. Kolaskar Tongaonkar antigenicity methods and predict location of antigenic determinants within utrophin from *Dracunculus medinensis* that are antigenic by eliciting an antibody response. This plot predicts those segments among a protein sequence that are to be antigenic by eliciting an antibody response (threshold setting = 1.030). This scale predicted a 7 length peptide in the position: 52 (Residue: L) and the sequence: 49- LSQLKLV-55 with score:1.156 (Maximum) . A typical profile show characteristic peaks and troughs, corresponding to the most hydrophobic and most hydrophilic parts of the protein respectively. Different residues which are rankings are commonly used hydrophobicity scales. While the scales differ in detail, there is a general consensus regarding the types of residue that appear at the most hydrophobic end (S, L, V and M) and those that appear at the most hydrophilic end (N, Q, E, D and K) (Figure 1 to Figure 4). We also find the location in solvent accessible regions in protein by using the hydrophobic scale Emini surface accessibility. This prediction revealed an epitope with 6 amino acid residues maximum (2.755) in the sequence positions:40 i.e 38- QKLDDE-43 and at position:41 with sequence 39- KLDDEQ-44 of utrophin from *Dracunculus medinensis* (Table 3). Hydropathy scale is a physiochemical property that quantifies the hydrophobicity of an amino acid. A window size is suggested to be 7-9 residues for predicting surface sites. The most of used scales are hydrophobicity scales which are derived on the basis of experimental studies on partitioning of peptides in apolar and polar solvents to predict membrane-spanning segments that are highly hydrophobic and secondary structure conformational parameter scales. The maximum region of hydrophilicity is to be considered as an antigenic site having hydrophobic characteristics.

Conclusion

Peptide fragments of utrophin from *Dracunculus medinensis* involved multiple antigenic components to direct and empower the immune system to protect the host. From the above result, it is concluded that antigenicity methods predict the location of antigenic determinants utrophin from *Dracunculus medinensis* that are antigenic by eliciting an antibody response. Hence, the region spanning the sequence positions will be of greater importance for epitope-based vaccine design. The amino acids making up the epitope are usually charged and hydrophilic in nature. From the study of physicochemical properties, it was found that, the region of maximal hydrophilicity is likely to be antigenic site, having hydrophobic characteristics because c- terminal region of utrophin from *Dracunculus medinensis* is solvent accessible. The mobility of protein segments those are located on the surface of a protein due to an entropic energy potential which seem to correlate well with known antigenic determinants. These antigenic peptides can be used as their identifiers. Therefore, these antigenic determinants are also important for synthetic peptide vaccine production.

Conflict of Interest

None.

Bibliography

1. Sweet RM and Eisenberg D. "Correlation of sequence hydrophobicities measures similarity in three-dimensional protein structure". *Journal of Molecular Biology* 171.4 (1983): 479-488.
2. Kyte J and Doolittle RF. "A Simple Method for Displaying the Hydrophobic Character of a Protein". *Journal of Molecular Biology* 157.1 (1982): 105-132.
3. Abraham DJ and Leo AJ. "Extension of the fragment method to calculate amino acid zwitterions and side chain partition coefficients". *Proteins* 2.2 (1987): 130-152.
4. Bull HB and Breese K. "Surface tension of amino acid solutions: A hydrophobicity scale of the amino acid residues". *Archives of Biochemistry and Biophysics* 161.2 (1974): 665-670.
5. Guy HR. "Amino acid side chain partition energies and distributions of residues in soluble proteins". *Biophysical Journal* 47.1 (1985): 61-70.
6. Miyazawa S and Jernigen RL. "Estimation of Effective Interresidue Contact Energies from Protein Crystal Structures: Quasi-Chemical Approximation". *Macromolecules* 18.3 (1985): 534-552.
7. Roseman MA. "Hydrophilicity of polar amino acid side-chains is markedly reduced by flanking peptide bonds". *Journal of Molecular Biology* 200.3 (1988): 513-522.
8. Chothia C. "The nature of accessible and buried surfaces in proteins". *Journal of Molecular Biology* 105.1 (1976): 1-14.
9. Eisenberg D., et al. "Analysis of membrane and surface protein sequences with the hydrophobic moment plot". *Journal of Molecular Biology* 179.1 (1984): 125-142.
10. Manavalan P and Ponnuswamy PK. "Hydrophobic character of amino acid residues in globular proteins". *Nature* 275.5681 (1978): 673-674.
11. Black SD and Mould DR. "Development of Hydrophobicity Parameters to Analyze Proteins Which Bear Post- or Cotranslational Modifications". *Analytical Biochemistry* 193.1 (1991): 72-82.

12. Fauchere JL and Plisk VE. "Hydrophobic parameters of amino-acid side-chains from the partitioning of N-acetyl-amino-acid amide". *European Journal of Medicinal Chemistry* 18.4 (1983): 369-375.
13. Janin J. "Surface and inside volumes in globular proteins". *Nature* 277.5696 (1979): 491-492.
14. Rao MJK and Argos P. "A conformational preference parameter to predict helices in integral membrane proteins". *Biochimica et Biophysica Acta* 869.2 (1986): 197-214.
15. Tanford C. "Contribution of hydrophobic interactions to the stability of the globular conformation of proteins". *Journal of the American Chemical Society* 84.22 (1962): 4240-4274.
16. Welling GW, et al. "Prediction of sequential antigenic regions in proteins". *FEBS Letters* 188.2 (1985): 215-218.
17. Rose GD, et al. "Hydrophobicity of amino acid residues in globular proteins". *Science* 229.4716 (1985): 834-838.
18. Wolfenden RV, et al. "Affinities of amino-acid side-chains for solvent water". *Biochemistry* 20.4 (1985): 849-855.
19. Wilson KJ, et al. "The behaviour of peptides on reverse-phase supports during high-pressure liquid chromatography". *Biochemical Journal* 199.1 (1981): 31-41.
20. Aguilar JC and Rodríguez EG. "Vaccine adjuvants revisited". *Vaccine* 25.19 (2007): 3752-3762.
21. Gomase VS, et al. "Prediction of MHC binding peptides and epitopes from alfalfa mosaic virus". *Current Drug Discovery Technologies* 4.2 (2007): 117-215.
22. Gomase VS. "Prediction of antigenic epitopes of utrophin Bmbktx1 from *Mesobuthus martensii*". *Current Drug Discovery Technologies* 3.3 (2006): 225-229.
23. Mishra S and Gomase VS. "Study Hydrophobicity and Antigenicity of Cytochrome C Oxidase Subunit II from *D. medinensis*: New Prototype of Synthetic Vaccine Development". *Immunochemistry Immunopathology* 2 (2016): 113.
24. Mishra S and Gomase VS. "Identification of Antigenic Determinants, Solvent Accessibility and MHC Binders of Antigen NADH Dehydrogenase Subunit 5 from a Little Dragon from Medina (*Dracunculus medinensis*)". *Drug Designing* 5 (2016): 126.
25. Mishra S and Gomase VS. "Study of Hydrophobicity and Prediction of Antigenic Epitope of NADH dehydrogenase subunit 5 from *D. medinensis*". *Drug Designing* 5 (2016): 127.
26. Hopp TP and Woods KR. "Prediction of protein antigenic determinants from amino acid sequences". *Proceedings of the National Academy of Sciences USA* 78.6 (1980): 3824-3828.
27. Parker JMR, et al. "New hydrophilicity scale derived from high-performance liquid chromatography retention data: correlation of predicted surface residues with antigenicity and X-ray derived accessible sites". *Biochemistry* 25.19 (1986): 5425-5432.
28. Kolaskar AS and Tongaonkar PC. "A semi-empirical method for prediction of antigenic determinants on protein antigens". *FEBS Letters* 276.1-2 (1990): 172-174.
29. Jens Erik Pontoppidan Larsen, et al. "Improved method for predicting linear B-cell epitopes". *Immunome Research* 2 (2006): 2.

30. Senior K. "Taking the bite out of snake venom". *Lancet* 353.9168 (1999): 1946.
31. Ahmad S., *et al.* "Real value prediction of solvent accessibility from amino acid sequence". *Proteins* 50.4 (2003): 629-635.
32. Emini EA., *et al.* "Induction of hepatitis a virus-neutralizing antibody by a virus-specific synthetic peptide". *Journal of Virology* 55.3 (1985): 836-839.
33. Karplus PA and Schulz GE. "Prediction of chain flexibility in proteins: a tool for the selection of peptide antigen". *Naturwissenschaften* 72 (1985): 212-213.
34. Dill KA. "Dominant forces in protein folding". *Biochemistry* 29.31 (1990): 7133-7155.

Volume 2 Issue 5 December 2016

© All rights reserved by Sonu Mishra and Virendra Gomase.