

## Feed Additives to Increase Plant Protein Substitution in Aquafeeds

**Mahmoud A O Dawood\***

*Department of Animal Production, Faculty of Agriculture, Kafrelsheikh University, Kafrelsheikh, Egypt*

**\*Corresponding Author:** Mahmoud A O Dawood, Department of Animal Production, Faculty of Agriculture, Kafrelsheikh University, Kafrelsheikh, Egypt.

**Received:** May 31, 2017; **Published:** June 30, 2017

### Abstract

Aquaculture is one of the fastest growing sectors as an alternative option in animal protein sources. Therefore, it is an important task to provide farmed fish with high quality diets to obtain the best performance. However, high quality fish diets should contain not only optimal quality and quantity of nutrients but also additives to keep fish healthy and optimal growth. Fishmeal (FM) is one of the most important components in practical diets for aquafeeds, but its increasing demand, unstable supply and high price have made it necessary to search for alternative protein sources. The increased utilization of soybean meal (SBM) has been embraced as a sustainable alternative to FM. Recently, functional feed additives is a natural way to transform complex feed components into absorbable nutrients. Therefore, the addition of feed additives in fish feeds would improve nutrients utilization in plant protein sources, reducing feed cost and the excretion of nutrients into the environment.

**Keywords:** Soy Bean Meal; Fish Meal; Feed Additives; Aquafeeds

Fish meal (FM) represents an ideal nutritional source of dietary protein for fish [1]. Increasing demand, unstable supply, and high prices of FM along with the continuous expansion of aquaculture are reasons for many nutritionists to realize that soon they will no longer be able to afford it as a major protein source in aquafeeds [2]. Currently one of the challenges that fish nutritionists face is the need to partially or totally replace FM with less expensive, non-traditional animal or plant protein sources [2-5].

Soybean meal (SBM) has been recognized as one of the most appropriate alternative protein sources for FM in aquafeed because of their consistent nutritional composition, comparatively balanced amino acid profile, availability, and reasonable price [6-8]. Methods for increasing SBM's inclusion rates in soy-sensitive species are required, and one of the methods is to apply dietary supplementation of functional feed additives [1,9]. Substituting SBM for FM at low levels has been relatively easy but substituting at higher levels is difficult. There are a number of challenges that must be overcome to maintain acceptable growth rates and feed efficiency values at higher levels of substitution of fish meal. The first is cost of plant protein concentrates. Until recently, fish meal protein was much less expensive than protein from soy or wheat concentrates, available as soy protein concentrate or wheat gluten meal.

It is well known that, SBM contains many anti-nutritive compounds including trypsin inhibitor, lectin, saponins, phytate, flatulence-producing oligosaccharides and allergenic protein [2]. Because the aquatic animals' intestinal tracts are not fully developed and easily damaged by the anti-nutritive compounds, several methods have been developed to eliminate the anti-nutritive compounds. As has been done previously, we can effectively decrease trypsin inhibitor and phytate content in soybeans, increase the dry matter content and increase the digestion rate of nitrogen and amino acid to improve the feed conversion ratio by heating [3]. Still, some allergenic proteins, such as glycinin,  $\alpha$ -conglycinin and  $\beta$ -conglycinin, are not deactivated by heating, and will harm young animals. Some reports indicate that the fermenting process or enzyme treatment with protease and carbohydrase can decrease the content of flatulence-producing oligosaccharides and allergenic proteins, and increase the digestibility of SBM [9-11]. However, the selection of bacteria used for fermentation is very important.

To optimize the use of the plant protein in aquafeeds, there is an ongoing search to find dietary additives that could protect the intestine from the effects of SBM. Traditionally, additives have been incorporated into fish diets to control diseases, increase health, and improve the stress response [12-19]. However, little focus has been given to the use of additives for controlling intestinal inflammation [20-22]. At present, there are only two studies that evaluate the effects of additives, specifically mannan-oligosaccharide and  $\beta$ -glucans, on soybean meal-triggered intestinal inflammation in farmed fish [17,23-25]. These investigations indicate that only mannan-oligosaccharide is able to decrease, to varying degrees, the altered intestinal histology observed in fish fed with diets including low amounts of soybean meal [16,17].

FM contained abundant nucleic acids and nucleotides, while little was found in SBM [2]. Nucleotides as feed additives play an important role in gut development and repair, skeletal muscle development, heart function and immune response [26,27], and gained wide attention as potential immunomodulators. Research related to nucleotide nutrition in fish was stimulated by the reports of Burrells, *et al.* [28,29] who demonstrated that dietary nucleotide supplementation improved growth, immune responses and disease resistance of Atlantic salmon. Similar results have also been reported in tilapia (*Oreochromis mossambicus*) [30], Pacific white shrimp (*Litopenaeus vannamei*) [31], grouper (*Epinephelus malabaricus*) [32] and rainbow trout [33]. However, nucleotide supplementation did not significantly influence the growth of turbot (*Scophthalmus maximus* L.) fed diets with 30% to 50% SBM but could be helpful to improve the non-specific immune response and the intestinal histological structure of turbot [2].

Probiotics may be beneficial for aquatic animals and could result in improved performance and health [33-35]. Probiotics were selected for fermented soybean meal production with different amounts of solution [36]. Probiotics could play a role in the ability to utilize high levels of soybean in rainbow trout, as already observed by Sealey, *et al.* [37] and Ramos, *et al.* [38]. Also, probiotics can play a role in soybean tolerance since lactic acid fermentation was able to eliminate indigestible carbohydrates and anti-nutritional factors in soybean meal fed to Atlantic salmon [39] and the addition of *Lactobacillus plantarum* has improved the ability of amberjack (*Seriola dumerili*) to deal with high levels of soybean meal [1]. Further, prebiotics increased protein digestibility of the soybean-meal-based experimental diets of red drum (*Sciaenops ocellatus*) (Linnaeus) [40].

## Conclusion

In conclusion, the use of feed additive is recommended to improve the performance and health condition of cultured fish species [41]. Also, increased replacement levels of plant protein sources was observed in several cultured fish species upon the supplementation of functional feed additives which result in reducing the cost of feeding.

## Bibliography

1. Dawood MAO., *et al.* "Effects of partial substitution of fish meal by soybean meal with or without heat-killed *Lactobacillus plantarum* (LP20) on growth performance, digestibility, and immune response of Amberjack, *Seriola dumerili* Juveniles". *BioMed Research International* (2015): 514196.
2. Peng M., *et al.* "Effects of nucleotide supplementation on growth, immune responses and intestinal morphology in juvenile turbot fed diets with graded levels of soybean meal (*Scophthalmus maximus* L.)". *Aquaculture* 392 (2013): 51-58.
3. Lunger AN., *et al.* "The effects of organic protein supplementation upon growth, feed conversion and texture quality parameters of juvenile cobia (*Rachycentron canadum*)". *Aquaculture* 264.1 (2007): 342-352.
4. Wang L., *et al.* "Effects of soybean meal fermentation by *Lactobacillus plantarum* P8 on growth, immune responses, and intestinal morphology in juvenile turbot (*Scophthalmus maximus* L.)". *Aquaculture* 464 (2016): 87-94.
5. Yan J., *et al.* "Effects of dietary chitosan on growth, lipid metabolism, immune response and antioxidant-related gene expression in *Misgurnus anguillicaudatus*". *Beneficial Microbes* 8.3 (2017): 439-449.

6. Chen W, *et al.* "Effects of dietary soybean saponins on feed intake, growth performance, digestibility and intestinal structure in juvenile Japanese flounder (*Paralichthys olivaceus*)". *Aquaculture* 318.1 (2011): 95-100.
7. Shimeno S, *et al.* "The growth performance and body composition of young yellowtail fed with diets containing defatted soybean meal for a long period". *Bulletin of the Japanese Society of Scientific Fisheries (Japan)* 59.3 (1993): 821-825.
8. Storebakken T, *et al.* "Soy products as fat and protein sources in fish feeds for intensive aquaculture". *Soy in Animal Nutrition* (2000): 127-170.
9. Dawood MAO, *et al.* "Effects of Using Exogenous Digestive Enzymes or Natural Enhancer Mixture on Growth, Feed Utilization, and Body Composition of Rabbitfish, *Siganus rivulatus*". *Journal of Agricultural Science and Technology B* 4 (2014): 180-187.
10. Goda A, *et al.* "Effect of Using Baker's Yeast and Exogenous Digestive Enzymes as Growth Promoters on Growth, Feed Utilization and Hematological Indices of Nile tilapia, *Oreochromis niloticus* Fingerlings". *Journal of Agricultural Science and Technology B* 2 (2012): 15-28.
11. Yigit NO and Keser E. "Effect of cellulase, phytase and pectinase supplementation on growth performance and nutrient digestibility of rainbow trout (*Oncorhynchus mykiss*, Walbaum 1792) fry fed diets containing canola meal". *Journal of Applied Ichthyology* 32.5 (2016): 938-942.
12. Adel M, *et al.* "Effects of *Pediococcus pentosaceus* supplementation on growth performance, intestinal microflora and disease resistance of white shrimp, *Litopenaeus vannamei*". *Aquaculture Nutrition* (2017).
13. Adel M, *et al.* "Effects of dietary *Spirulina platensis* on growth performance, humoral and mucosal immune responses and disease resistance in juvenile great sturgeon (*Huso huso* Linnaeus, 1754)". *Fish and Shellfish Immunology* 56 (2016): 436-444.
14. Dawood MAO and Koshio S "Recent advances in the role of probiotics and prebiotics in carp aquaculture: A review". *Aquaculture* 454 (2016): 243-251.
15. Dawood MAO and Koshio S "Vitamin C supplementation to optimize growth, health and stress resistance in aquatic animals". *Reviews in Aquaculture* (2016).
16. Dimitroglou A, *et al.* "Effects of mannan oligosaccharide (MOS) supplementation on growth performance, feed utilisation, intestinal histology and gut microbiota of gilthead sea bream (*Sparus aurata*)". *Aquaculture* 300 (2010): 182-188.
17. Refstie S, *et al.* "Effects of dietary yeast cell wall  $\beta$ -glucans and MOS on performance, gut health, and salmon lice resistance in Atlantic salmon (*Salmo salar*) fed sunflower and soybean meal". *Aquaculture* 305 (2010): 109-116.
18. Tahmasebi-Kohyani A, *et al.* "Effects of dietary nucleotides supplementation on rainbow trout (*Oncorhynchus mykiss*) performance and acute stress response". *Fish Physiology and Biochemistry* 38.2 (2012): 431-440.
19. Yan J, *et al.* "Effects of dietary protein and lipid levels on growth performance, fatty acid composition and antioxidant-related gene expressions in juvenile loach *Misgurnus anguillicaudatus*". *Aquaculture Research* (2017).
20. Dawood MAO, *et al.* "Interaction effects of dietary supplementation of heat-killed *Lactobacillus plantarum* and  $\beta$ -glucan on growth performance, digestibility and immune response of juvenile red sea bream, *Pagrus major*". *Fish and Shellfish Immunology* 45.1 (2015): 33-42.

21. Dawood MAO., *et al.* "Probiotics as an environment-friendly approach to enhance red sea bream, *Pagrus major* growth, immune response and oxidative status". *Fish and Shellfish Immunology* 57 (2016): 170-178.
22. Dawood MAO., *et al.* "Immune responses and stress resistance in red sea bream, *Pagrus major*, after oral administration of heat-killed *Lactobacillus plantarum* and vitamin C". *Fish and Shellfish Immunology* 54 (2016): 266-275.
23. Dawood MAO., *et al.* "Changes in the growth, humoral and mucosal immune responses following  $\beta$ -glucan and vitamin C administration in red sea bream, *Pagrus major*". *Aquaculture* 470 (2017): 214-222.
24. Dawood MAO., *et al.* "Physiological response, blood chemistry profile and mucus secretion of red sea bream (*Pagrus major*) fed diets supplemented with *Lactobacillus rhamnosus* under low salinity stress". *Fish Physiology and Biochemistry* 43.1 (2017): 179-192.
25. Dawood MAO., *et al.* "Dietary supplementation of  $\beta$ -glucan improves growth performance, the innate immune response and stress resistance of red sea bream, *Pagrus major*". *Aquaculture Nutrition* 23.1 (2017): 148-159.
26. Hossain MS., *et al.* "Efficacy of nucleotide related products on growth, blood chemistry, oxidative stress and growth factor gene expression of juvenile red sea bream, *Pagrus major*". *Aquaculture* 464 (2016): 8-16.
27. Li P and Gatlin DM. "Nucleotide nutrition in fish: current knowledge and future applications". *Aquaculture* 251 (2006): 141-152.
28. Burrells C., *et al.* "Dietary nucleotides: a novel supplement in fish feeds. 1. Effects on resistance to disease in salmonids". *Aquaculture* 199 (2001): 159-169.
29. Burrells C., *et al.* "Dietary nucleotides: a novel supplement in fish feeds. 2. Effects on vaccination, salt water transfer, growth rates and physiology of Atlantic salmon (*Salmo salar* L.)". *Aquaculture* 199 (2001): 171-184.
30. Ramadan A., *et al.* "The effect of ascogen on the immune response of tilapia fish to *Aeromonas hydrophila* vaccine". *Fish and Shellfish Immunology* 4 (1994): 159-165.
31. Murthy HS., *et al.* "Dietary  $\beta$ -glucan and nucleotide effects on growth, survival and immune responses of pacific white shrimp, *Litopenaeus vannamei*". *Journal of Applied Aquaculture* 21.3 (2009): 160-168.
32. Lin YH., *et al.* "Dietary nucleotide supplementation enhances growth and immune responses of grouper, *Epinephelus malabaricus*". *Aquaculture Nutrition* 15.2 (2009): 117-122.
33. Tahmasebi-Kohyani A., *et al.* "Dietary administration of nucleotides to enhance growth, humoral immune responses, and disease resistance of the rainbow trout (*Oncorhynchus mykiss*) fingerlings". *Fish and Shellfish Immunology* 30.1 (2011): 189-193.
34. Dawood MAO., *et al.* "Effects of dietary inactivated *Pediococcus pentosaceus* on growth performance, feed utilization and blood characteristics of red sea bream, *Pagrus major* juvenile". *Aquaculture Nutrition* 22.4 (2016b): 923-932.
35. Dawood MAO., *et al.* "Effects of dietary supplementation of *Lactobacillus rhamnosus* or/and *Lactococcus lactis* on the growth, gut microbiota and immune responses of red sea bream, *Pagrus major*". *Fish and Shellfish Immunology* 49 (2016): 275-285.
36. Rivero-Ramirez F., *et al.* "Combined effects of dietary mannan oligosaccharides and *Pediococcus acidilactici* and their combination in low fish meal and fish oil diets for European sea bass, *Dicentrarchus labrax*, juveniles". *Fish and Shellfish Immunology* (2016): 53-69.

37. Sealey WM., *et al.* "Soybean meal level and probiotics in first feeding fry diets alter the ability of rainbow trout *Oncorhynchus mykiss* to utilize high levels of soybean meal during grow-out". *Aquaculture* 293 (2009): 195-203.
38. Ramos MA., *et al.* "Commercial *Bacillus* probiotic supplementation of rainbow trout (*Oncorhynchus mykiss*) and brown trout (*Salmo trutta*): growth, immune responses and intestinal morphology". *Aquaculture Research* (2016).
39. Refstie S., *et al.* "Lactic acid fermentation eliminates indigestible carbohydrates and antinutritional factors in soybean meal for Atlantic salmon (*Salmo salar*)". *Aquaculture* 246 (2005): 331-345.
40. Burr G., *et al.* "Effects of prebiotics on nutrient digestibility of soybean-meal-based diets by red drum (*Sciaenops ocellatus*)". *Aquaculture Research* 39.15 (2008): 1680-1686.
41. Dawood MAO., *et al.* "Effects of heat killed *Lactobacillus plantarum* (LP20) supplemental diets on growth performance, stress resistance and immune response of red sea bream, *Pagrus major*". *Aquaculture* 442 (2015): 29-36.

**Volume 9 Issue 5 June 2017**

**© All rights reserved by Mahmoud A O Dawood.**