Afiukwa Ngozi¹, Nwuzo Agabus¹, Okonkwo Eucharia¹, Udu-Ibiam Onyinyechi¹, Emioye Abraham², Ejikeugwu Chika^{1*} and Iroha Ifeanyichukwu¹

¹Department of Applied Microbiology, Faculty of Science, Ebonyi State University, Abakaliki, Ebonyi State, Nigeria ²Department of Biological Science, Federal University Ndufu-Alike, Ikwo, Ebonyi State, Nigeria

*Corresponding Author: Ejikeugwu Chika, Department of Applied Microbiology, Faculty of Science, Ebonyi State University, Abakaliki, Ebonyi State, Nigeria.

Received: February 06, 2018; Published: May 02, 2018

Abstract

Antibiotic resistance is a natural biological phenomenon that is vital to microbial colonization and survival in the environment. But this process is of clinical significance because of the ability of antibiotic resistant bacteria to render inefficacious some available antimicrobial agents. This study evaluated the prevalence and antibiotic susceptibility pattern of Escherichia coli isolates from cloacal swabs of wild and domestic birds in Ebonyi state, Nigeria over a three (3) year period. Four different bird species including bats, ducks, parrots and pigeons were used for this study. A total of fifty (50) cloacae swabs of each wild and domestic bird species were collected using sterile swab sticks for each year. The samples were analyzed using standard microbiology techniques and the isolates were biochemically confirmed using API kits. Antibiotic susceptibility test on the isolates was conducted using modified Kirby-Bauer disc diffusion method. The results obtained from this study showed that E. coli were present at varying levels in the 4 bird species investigated. E. coli species were most prevalent in pigeons across the 3 years with the range of 30.43% - 65.63%. This was followed by ducks which recorded between 17.95% - 34.38% across the 3 years. Parrots and bats respectively had 28.26% and 19.57% in 2012; 17.95% and 15.38% in 2014. The result of the antimicrobial susceptibility testing revealed that more than 50% of the isolated E. coli was resistant or intermediately resistant to cefuroxime, ceftazidime and cefotaxime. However, the E. coli isolates were found to be most susceptible to imipenem, meropenem and ertapenem. Isolates from parrots and bats were generally more resistant to the antibiotics than the same isolates from pigeon and duck samples. Notably, resistance to imipenem was observed only in E. coli isolates recovered from parrots and bats at the rate of 22.22%. The carbapenems, imipenem and meropenem were the most active antibiotics tested in this study. Conclusively, this study has shown that E. coli isolates recovered from birds are antimicrobial resistant in nature. The frequency of antibiotic resistant E. coli in birds in the community is of public health importance because of their implications in community-acquired infections.

Keywords: Escherichia coli; Antibiotics; Resistance; Antibiogram; Birds; Nigeria

Introduction

From the time when the first sulfa and penicillin were introduced in the 1930s and 1940s, respectively, bacteria have continued to discover a remarkable ability to increase different types of resistance mechanisms to antimicrobials they were susceptible to [1]. Antibiotic resistant bacteria are exceedingly important to human health, but the wild reservoirs of resistant determinants are poorly understood [2]. Their origins in wild life is important to human health because of the increasing importance of zoonotic diseases as well as the need for predicting emerging resistant pathogens [2]. Wild animals provide a biological mechanism for the spread of antibiotic resistance genes and the antimicrobial-resistant Escherichia coli and Enterococcus species that originated from wild life species were reported for the first time from Japanese wild birds [3-5]. E. coli was also reported 5 years later in South Africa baboons that feed on human refuse [6,7]. E. coli is a facultative anaerobic Gram negative bacterium found in the Enterobacteriaceae; and they are found in the intestinal tract of warm blooded animals including mammals, poultry and wild birds [8,9]. Faecal E. coli is considered to be a key indicator for the selective pressure exerted by the use of antimicrobials on intestinal populations of bacteria [10]. The transfer of antibiotic resistance in enteric bacteria including E. coli, Salmonella spp., Campylobacter spp. and Enterococci spp. from birds to humans have been previously reported and this can occur through the food chain or by direct contact [11-13]. Therefore, the frequent occurrence of antimicrobial resistance in wildlife has some public health implications including the potential to serve as an environmental reservoir of resistance gene and zoonotic diseases [2]. The mobility and migration of avian species are remarkable biological phenomena. At some points in their yearly migrations, they can disperse microorganisms that pose a threat to the health of both animals and the public in diverse geographical areas [14,15]. Birds may also act as bio-indicators of the level of contamination of bacterial pathogens and genetic determinants of antibiotic resistance in the environment [16]. Because of their ease in picking up human and environmental bacteria and their diverse presence in different ecological niches, wild birds could act as transporters or reservoirs of resistant bacteria. They could probably have an important epidemiological role in the dissemination of resistant pathogenic microorganisms present in humans [2,13,17]. This study therefore evaluated the possible involvement of birds such as pigeons, ducks, parrots and bats in the dissemination of antibiotic resistant E. coli in Ebonyi State, Nigeria.

Materials and Methods

Sample collection: A total of 500 cloacal samples from four different wild and domestic birds' species (duck, pigeon, parrots, bat) were used for this study for a period of 3 years (2012 - 2014). Each bird from which each sample was collected was labeled appropriately to avoid repetition in sample collection.

Bacteriological analysis: Each sample collected was used to inoculate sterile nutrient broth in test tubes (Oxoid, UK) and incubated for 18 - 24h at 37°C. A loopful from each tube was inoculated onto MacConkey (MAC) and Eosin Methylene Blue (EMB) agar (Oxoid, UK), and incubated for 18 - 24h at 37°C. Suspected colonies of *E. coli* were subcultured onto freshly prepared MAC and EMB agar and incubated for 18-24 hrs at 37°C to obtain pure cultures. *E. coli* isolates were identified using API kit [18].

Antimicrobial susceptibility testing: All *E. coli* isolates were screened for their susceptibility to different antibiotics using the Kirby-Bauer disc diffusion technique as per the guidelines of Clinical Laboratory Standards Institute (CLSI). A 0.5 McFarland equivalent standard of each of the test organism was inoculated on the surface of Mueller-Hinton (MH) agar plates using a sterile swab stick for each test organism. Antibiotic disks of cefuroxime, CXM (30 µg); ceftazidime, CAZ (30 µg); cefotaxime, CTX (30 µg); ceftriaxone, CRO (30 µg); cefoxitin, FOX (30 µg) amoxicillin/clavulanic acid, AMC (20/10 µg); aztreonam, ATM (30 µg); imipenem, IMI (30 µg); meropenem, MEP (30 µg) and ertapenem, ETP (30 µg) [Oxoid, UK] were aseptically placed on the surface of the inoculated MH agar plates at a distance of 25 mm. The susceptibility plates were incubated for 18 - 24 hrs at 37°C. The diameters of the observed clear zones of inhibition were measured and recorded in line with the CLSI criteria, and as was previously described [18,19].

Results

The results obtained from this study showed that a total of 117 isolates of *E. coli* were obtained across the three years of study. Of the 117 isolates, 46 (39.32%), 39 (33.33%) and 32 (27.35%) were obtained in 2012, 2013 and 2014, respectively (Table 1).

Year	Sample source (Birds)	Cloacal swab sample	Percentage occurrence of E. coli
2012	Ducks	50	10 (21.74)
	Pigeons	50	14 (30.43)
	Weaver birds	50	13 (28.26)
	Bats	50	9 (19.57)
	Total	200	46
2013	Ducks	50	7 (17.95)
	Pigeons	50	19 (48.72)
	Weaver birds	50	7 (17.95)
	Bats	50	6 (15.38)
	Total	200	39
2014	Ducks	50	11 (34.38)
	Pigeons	50	21 (65.63)
	Weaver birds	-	-
	Bats	-	-
	Total	100	32

Table 1: Prevalence of E. coli obtained from four wild and domestic birds species in 2012, 2013 and 2014.

In 2012, the 10 E. coli species screened, were 100% susceptible to IMI and ETP; followed by 8 (80.0%) each to FOX and MEP; 7 (70.0%) to ATM; 6 (60.0%) each to CTX and CRO; 3 (30.0%) and 1 (10.0%) to CXM and CAZ, respectively (Figure 1). The highest resistance recorded was 90.0% and to CAZ; followed by 70.0% to CXM, 40.0% each to CXT and CRO; 30.0% to ATM; 20.0% each to FOX and MEP. None of the isolates was resistant to IMI and ETP (Figure 1).

Citation: Ejikeugwu Chika., *et al.* "A Three-Year Study on the Prevalence and Antibiotic Susceptibility Pattern of *Escherichia coli* Isolated from Cloacal Swabs of Wild and Domestic Birds in Ebonyi State, Nigeria". *EC Microbiology* 14.5 (2018): 266-273.

267

268

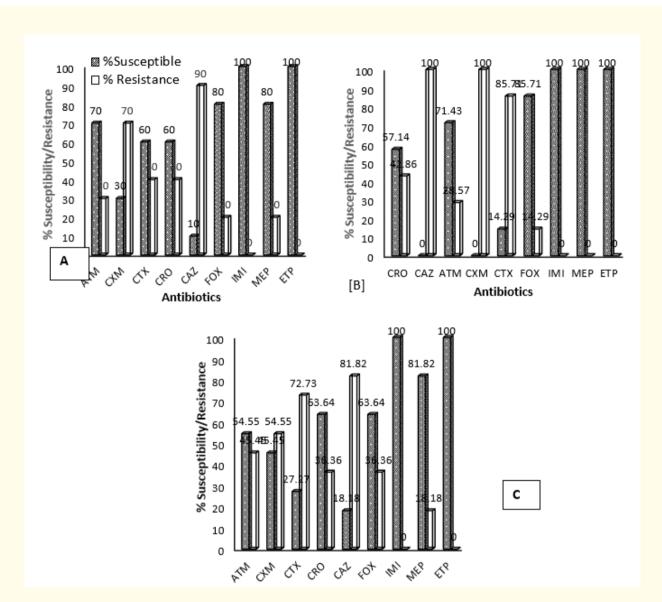
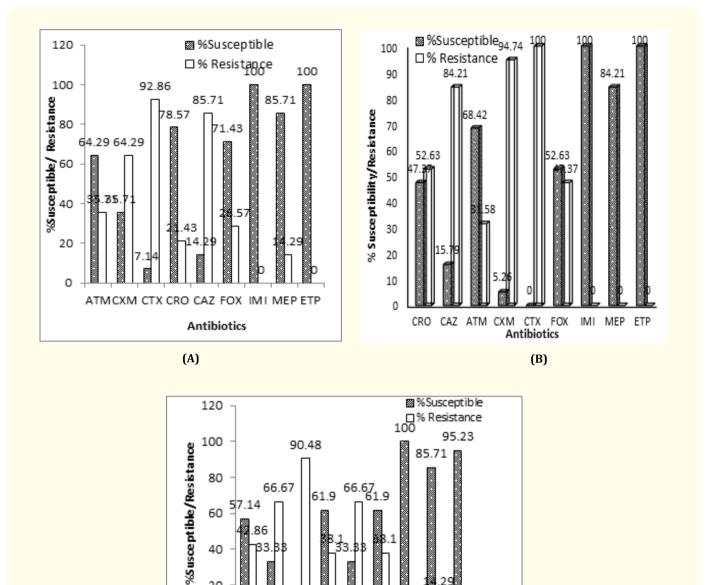


Figure 1: Antibiotic Susceptibility and Resistance Patterns of E. coli Isolated from Duck Cloacal Swabs in 2012 [A], 2013 [B] and 2014 [C].

In 2013, all the *E. coli* species screened were observed to be susceptible to IMI, MEP and ETP, 7 (100%). The isolates were totally resistant to CAZ and CXM, 7 (100%) each; 6 (85.71%) of the isolates (Figure 1). In 2014, the 11 *E. coli* species screened were absolutely (100%) susceptible to IMI and ETP. In 2012, it was observed that all the 14 *E. coli* species screened were susceptible to IMI and ETP (100%). In 2013, all 19 *E. coli* species screened were susceptible to IMI and ETP (100%). In 2013, all 19 *E. coli* species screened were susceptible to IMI and ETP (100%). In 2014, the result showed all the isolates of *E. coli* screened to be susceptible only to IMI, 21 (100%); 20 of the 21 isolates (95.23%) and 18 (85.71%) were respectively susceptible to ETP and MEP.



269



66.67 61.9

ATM CXM CTX CRO CAZ FOX IMI MEP ETP Antibiotics

(C)

Figure 2: Antibiotic Susceptibility and Resistance Pattern of E. coli Isolated from Pigeon Cloacal Swabs in Abakaliki in 2012 [A], 2013 [B] and 2014 [C].

76

61.9

66.67

57.14

86

60

40

20

0

270

In 2012, the result showed all the *E. coli* species screened to be totally susceptible to IMI and ETP, 13 (100%); 11 (84.62%) were susceptible to MEP; 8 (61.54%) each to CRO and ATM; 7 (53.85%), 3 (23.08%), 2 (15.38%) and 1 (7.69%) to FOX, CAZ, CXM and CTX, respectively (Figure 3). In 2013, all the isolates (7) screened were susceptible to IMI, MEP, FOX and ETP (100%); 5 (71.43%) and 4 (57.14%) were respectively susceptible to ATM and CRO.

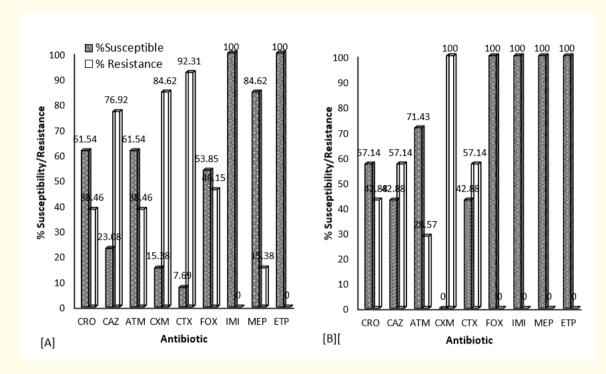


Figure 3: Antibiotic Susceptibility and Resistance Pattern of E. coli Isolated from Weaver bird Cloacal Swabs in Abakaliki in 2012 [A] and 2013 [B].

In 2012, 7 of the 9 isolates of E. coli screened (77.78%) were found to be susceptible to each of IMI, ETP and MEP; followed by 6 (66.67%) each to CRO and FOX; 5 (55.56%), 2 (22.22). The isolates were totally (100%) resistant to CTX; 8 (88.89%), 7 (77.78%), 4 (44.44%) were respectively resistant to CXM, CAZ and ATM; whereas only 3 (33.33%) were resistant to each of CRO and FOX and 2 (22.22%) were resistant to each of IMI, MEP and ETP (Figure 4). In 2013, the 6 E. coli species screened were all susceptible only to IMI, 5 of them (83.33%) were susceptible to ETP; 2 (33.33%) to each of CRO, FOX and MEP. All the isolates were 100% resistant to CXM and CTX; 83.33% of them were resistant to each of CAZ and ATM; followed by 66.67% to each of CRO, FOX and MEP; and only 16.67% was resistant to ETP; and none to IMI (Figure 4).

271

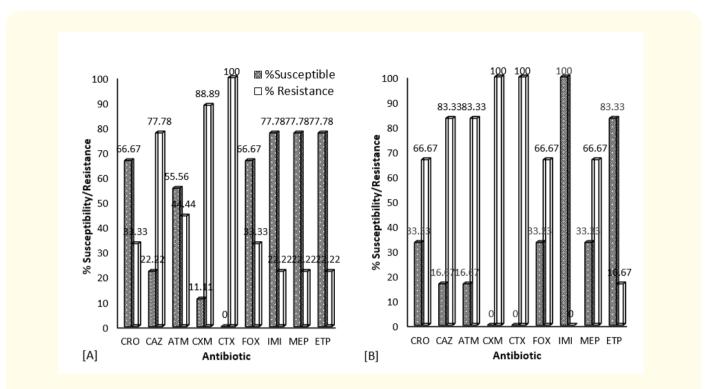


Figure 4: Antibiotic Susceptibility and Resistance Patterns of E. coli Isolated from Bat Cloacal Swabs in Abakaliki in 2012 [A] and 2013 [B].

Discussion

Escherichia coli isolates were more prevalent in pigeons (46.15%), and this was followed by ducks (23.93%), weaverbirds (17.07%) and bats (12.82%). This is in line with previous studies which reported that E. coli was the most prevalent Gram negative bacteria recovered from birds [20-23]. Wild and domestic bird species, including the long and short distance migrants occupying diverse ecological niches and adapting to varying feeding patterns, can as well host and contribute to the spread of multidrug resistant bacteria species [24,25]. Our result also showed that the *E. coli* species isolated from duck and pigeon cloacal swabs were generally more susceptible to the antibiotics used than those isolated from weaverbirds and bats. This could be associated with their closeness to humans who also use these antibiotics. Isolates from duck samples were most susceptible to imipenem (IMP), ertapenem (ETP), meropenem (MEP) and cefoxitin (FOX). Their percentage susceptibilities across the 3 years ranged from 54.55 - 100%. More than 50% of the E. coli isolates were also resistant to cefuroxime (CXM), cefotaxime(CTX) and ceftazidime (CAZ) over the 3 years under study. The susceptibility and resistance patterns of the E. coli isolated from pigeons were similar to those isolated from duck. Their percentage susceptibilities to IMP, ETP and MEP ranged from 84.21 - 100% while their resistance to CXM, CAZ and CTX ranged from 64.29 - 100%. E. coli isolated from parrots and bats were also most susceptible to IMP, ETP and MEP and most resistant to CTX, CXM and CAZ. In parrots, the E. coli susceptibility to IMP, MEP, and ETP ranged from 82.62 - 100% while their percentage resistance to CAZ, CXM and CTX ranged from 57.14 - 100%. In bats, susceptibility of *E. coli* to IMP, MEP and ETP ranged from 66.67 - 100% while their percentage resistance to CTX, CXM and CAZ was as high as 77.78 - 100%. The E. coli isolates from parrots and bats were generally more resistant to the antibiotics used when compared to the same isolates from pigeons and ducks. From this study, it is a matter of concern that E. coli isolated from birds are resistant to some commonly used antibiotics. This is most disturbing because the birds in question do not receive any direct antibiotic treatment as humans. These birds are highly migratory, feeding from diverse environments and could potentially contribute highly to the spread of

272

drug resistant microbes [1]. The high resistance of the isolates from these birds could be linked to their feeding behavior. They could feed at poorly managed hospital waste dumps; come in contact with human and animal excreta and may later visit any close-by water body to drink, making them both potential reservoirs and active spreaders of drug resistant bacteria [1]. Hassan [1] has also reported resistance to some classes of antibiotics by *E. coli* isolated from both commercial and free-range poultry birds that even have limited migration capacity and feed in less diverse environment. Similarly, antibiotic resistant *E. coli* have been previously obtained from ducks and geese [5,16], birds of prey [26,27], gulls [28-30] and feral pigeons [31]. Antibiotic resistance is a global phenomenon that bedevils the health sector and compromises our ability to effectively manage and treat some bacterial related infections. The emergence and spread of drug resistant bacteria which is very notable in the hospital environment is now being experienced in the non-hospital environment. This present study gives impetus to this rising incidence of antibiotic resistant bacteria in the non-hospital environment, and the possible damage that they cause to antimicrobial therapy.

Conclusion

This study reported the prevalence and antibiogram of *E. coli* isolates recovered from different wild and domestic bird species over a three (3) year period. And the results from this study shows that *E. coli* isolates from birds could be multidrug resistant in nature. Birds inclusive of migratory and domestic birds could serve as reservoir for antibiotic resistant bacteria due to their feeding habits or other environmental factors. It is important to be on the lookout for antibiotic resistant bacteria from birds, and prevent those circumstances that could make these birds disseminator's and reservoirs of antibiotic resistant Gram negative bacteria inclusive of *E. coli*.

Acknowledgments

Our thanks go to staff of the Applied Microbiology Department of Ebonyi State University, Abakaliki, Nigeria for helping out with isolate processing and storage.

Declaration of Interest

The authors report no declaration of interest.

Bibliography

- Hasan B. "Antimicrobial resistance and production of extended spectrum beta-lactamases in Enterobacteriaceae from birds in Bangladesh". Uppsala University, Department of Medical Sciences, Clinical Microbiology and Infectious Medicine, Akademiskasjukhuset, Uppsala, Sweden (2013): 1-75.
- Radhouani H., et al. "Potential Impact of Antimicrobial Resistance in Wildlife, Environment and Human Health". Frontier of Microbiology 5 (2014): 23.
- 3. Sato G. "Detection of conjugative R plasmids conferring chloramphenicol resistance in Escherichia coli isolated from domestic and feral pigeons and cows". *Zentralbl Bakteriol Orig A* 241.4 (1978): 407-417.
- 4. Kim T., *et al.* "Chronological study of Antibiotic Resistances and their Relevant Genes in Korean Avian Pathogenic E. coli isolates". *Journal of Clinical Microbiology* 45.10 (2007): 3309-3315.
- Tsubokura M. "Drug Resistance and Conjugative R Plasmids in Escherichia coli Strains Isolated from Migratory Waterfowl". Journal of Wildlife Diseases 31.3 (1995): 352-357.
- 6. Reed KD. "Birds, Migration and Emerging Zoonoses: West Nile Virus, LYME disease, Influenza A and Enteropathogens". *Clinical Medicine and Research* 1 (2003): 5-12.
- Heather KA. "Call of the Wild: Antibiotic Resistance Genes in Natural Environments". Nature Reviews Microbiology 8.4 (2010): 251-259.
- 8. Gordon DM. "The Distribution and Genetic Structure of E. coli in Australian Vertebrates: Host and Geographic Effects". *Microbiology* 149.12 (2003): 3575-3586.
- 9. Sorum and Sunde M. "Resistance to antibiotics in the normal flora of animals". Veterinary Research 32.3-4 (2001): 227-241.
- 10. Van den Bogaard AE. "Epidemiology of Resistance to Antibiotics: Links Between Animals and Humans". *International Journal of Antimicrobial Agents* 14.4 (2000): 335-355.
- 11. Linton AH. "The Colonization of the Human Gut by Antibiotic Resistant Escherichia coli from Chickens". *Journal of Applied Bacteriology* 43.3 (1977): 465-469.

273

- 12. Ojeniyi AA. "Direct Transmission of Escherichia coli from Poultry to Humans". *Epidemiology and Infection* 103 (1989): 513-522.
- 13. Radhouani H. "Wild Birds as Biological Indicators of Environmental Pollution: Antimicrobial Resistance Patterns of Escherichia coli and Enterococci Isolated from Common Buzzards (Buteobuteo)". *Journal of Medical Microbiology* 61.6 (2012): 837-43.
- Radhouani H. "Detection of Escherichia coli Harboring Extended-Spectrum β-Lactamases of the CTX-M Classes in Faecal Samples of Common Buzzards (Buteobuteo)". Journal of Antimicrobial Chemotherapy 65.1 (2010): 171-173.
- 15. Huba'lek Z. "Anannotated Checklist of Pathogenic Microorganisms Associated with Migratory Birds". *Journal of Wildlife Diseases* 40.4 (2004): 639-659.
- 16. Cole D. "Free-living Canada Geese and Antimicrobial Resistance". Emerging Infectious Diseases 11.6 (2005): 935-938.
- 17. Silva N. "Molecular Characterization of Vancomycin-Resistant Enterococci and Extended-Spectrum β-Lactamase-Containing Escherichia coli Isolates in Wild Birds from the Azores Archipelago". *Avian Pathology* 40.5 (2011): 473-479.
- Iroha IR. "Antimicrobial Resistance Pattern of Plasmid-Mediated Extended Spectrum Beta-Lactamase Producing Strains of Escherichia coli". Science Research Essay 3.6 (2008): 215-218.
- 19. Clinical Laboratory Standard Institute, CLSI. Performance standards for antimicrobial disk susceptibility test. Fifteenth informational supplement, CLSI document M100-S15. Wayne, PA. USA (2011).
- 20. Wallensten A. "Extended Spectrum Beta-Lactamases Detected in Escherichia coli from Gulls in Stockholm, Sweden". *Infection, Ecology and Epidemiology* 1 (2011): 7030.
- 21. Jingrang DA. US Environmental Protection Agency. Office of Microbiology, 2nd edition. MaGraw Hill Inc (2008): 417-418.
- 22. Adegunloye DV. "Microorganisms Associated with Poultry Faeces". Journal of Food, Agriculture and Environment 4 (2006): 41-42.
- 23. Guenther S. "Comparable High Rates of Germany and Mongolia". PLoS ONE 7.12 (2012): e53039.
- 24. Hernández J. "Globally Disseminated Human Pathogenic Escherichia coli of O25b-ST131 Clone, Harbouring blaCTX-M-15, found in Glaucous-winged Gull at remote Commander Islands, Russia". *Environmental Microbiology Reports* 2.2 (2010): 329-332.
- 25. Bonnedahl J. "Dissemination of Escherichia coli with CTX-M type ESBL between Humans and Yellow-Legged Gulls in the South of France". *PLoS ONE* 4.6 (2009): e5958.
- 26. Costa D. "Mechanisms of Escherichia coli Isolates Recovered from Wild Animals". Microbial Drug Resistance 14.1 (2008): 71-77.
- Blanco G. "Geographical variation in Cloacal Microflora and Bacterial Antibiotic Resistance in a Threatened Avian Scavenger in Relation to Diet and Livestock Farming Practices". Environmental Microbiology 9.7 (2007): 1738-1749.
- Gionechetti F. "Characterization of Antimicrobial Resistance and class 1 Integrons in Enterobacteriaceae isolated from Mediterranean Herring Gulls (Laruscachinnans)". Microbial Drug Resistance 14.2 (2008): 93-99.
- 29. Poeta P. "Seagulls of the Berlengas Natural Reserve of Portugal as Carriers of Fecal Escherichia coli Harboring CTX-M and TEM Extended-Spectrum Beta-Lactamases". *Applied and Environmental Microbiology* 74.23 (2008): 7439-7441.
- 30. Dolejska M. "High Prevalence of Antimicrobial-Resistant Genes and Integrons in Escherichia coli isolates from Black-Headed Gulls in the Czech Republic". *Journal of Applied Microbiology* 103.1 (2007): 11-19.
- 31. Radimersky T. "Antibiotic Resistance in Faecal Bacteria (Escherichia coli, Enterococcus spp.) in Feral Pigeons". *Journal of Applied Microbiology* 109 (2010): 1687-1695.

Volume 14 Issue 5 May 2018 ©All rights reserved by Ejikeugwu Chika., *et al.*