

Photorhabdus Insect-Related Proteins (*PirAB*) an Insecticidal Toxin against Dengue Vectors, *Aedes aegypti*

Ihsan Ulla*

Department of Biological Sciences, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia

***Corresponding Author:** Ihsan Ulla, Department of Biological Sciences, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia.

Received: November 09, 2017; **Published:** December 18, 2017

A number of vector-borne diseases posed genuine problems, including health and economic problems. Dengue fever is among the most hazardous vector-borne disease, which concern the public health issue [1]. The major cause of the fever is dengue virus (DENV), is a mosquito-borne RNA⁽⁺⁾ (positive stranded) virus, belongs to *Flaviviridae* family and *Flavivirus* genus [2]. Mosquito *Aedes aegypti* is the major vector and source of transmission of dengue virus. Toxins from *Bacillus thuringiensis* had been successfully used against the larvae of *A. aegypti*, but a gradual resistance was noticed in the insects against the toxins over the years [3].

Photorhabdus bacteria such as *P. luminescens*, *P. temperata* and *P. asymbiotica* produce an array of new toxins and virulence factors, against the insects including *A. aegypti*, and no resistance has been reported in the insect vectors against these toxins to date [3,4]. All three species of *Photorhabdus* are nematodes symbiont of the genus *Heterorhabditis* [5].

Photorhabdus sp. are now well known entomopathogenic bacteria against a wide range of insects when released in to the gut of insects [2]. Complete genome analysis of *Photorhabdus* sp. revealed that genes encoding hemolysins, and proteases are critical of insecticidal activities [6]. The *lic*; plu4092-4093 and plu4436-4437 encoding polypeptide chains in *Photorhabdus* sp., have been termed as *PirA* and *PirB*, respectively, for "*Photorhabdus* insect-related proteins A and B," reveal sequence similarity with endotoxins of *B. thuringiensis* as well as with developmentally regulated protein from *Leptinotarsa decemlineata* [7].

Moreover, it has been reported that the *PirAB* toxin from *Photorhabdus* sp. had the potential to kill larvae of many insects including *A. aegypti*. The *PirAB* toxin have been determined to equally effective, either expressed in *E. coli* expression system or/and purified from the culture of *Photorhabdus* sp [8]. The *Pir* toxins function as binary proteins and both are necessary for insecticidal activity [9]. All the two components of *Pir* (*PirA* and *PirB*) are encoded by genes, located at plu4093-4092 (*pirA*) and plu4437-4436 (*pirB*) loci in *Photorhabdus* genome. Both *Pir* components exhibit a strong functional similarity to the δ -endotoxins of *B. thuringiensis*, which make them a substitute of *Bt* toxin [9]. *PirA* component of the binary protein exhibits a bit sequence similarities with known toxic proteins, however, its counterpart (*PirB*) shows strong sequence similarity with the N-terminal side of the pore-forming domain of the Cry2A insecticidal toxin [10]. These similarities suggest the presence of a similar motif in *PirAB*. Moreover, *PirB* has also a strong similarity with the developmentally regulated protein (DRP) of *L. decemlineata* [11]. The DRP has been thought to have a putative juvenile hormone esterase (JHE) property because of its pattern of expression, that matches insect development profile and the levels of JH produced [12]. It is therefore, assumed that *PirB* may exhibit the same kind of activity. However, further study is need to elucidate a the *PirB* activity [6,13].

It has been evaluated in a comparative study of the rate of mortality of *PirAB*, *PirA* and *PirB*, when applied to *A. aegypti*. The *PirAB* was most effective against the larvae of *A. aegypti* as compared to *PirA* or *PirB* exclusively [14]. The *PirB* activity was improved gradually and was tended to be stable after 2 days. However, *PirA* caused mortality in larvae of *A. aegypti* and tended to be stable after 4 days of treatment [14].

It is concluded that chemical control of *A. aegypti* has more hazardous environmental consequences and biological control is a best solution against the dengue virus. However, there are consistent raise of resistance against *Bt* and other biocontrol system. *Photorhabdus* toxins are comparatively new and more efficient and the level of the resistance in the insects is still at very primitive. The *Photorhabdus* toxins PirAB is more efficient *A. aegypti*, and can to used against *A. aegypti* to control the dengue fever.

Bibliography

1. NE Boemare., *et al.* "DNA relatedness between *Xenorhabdus* spp. (Enterobacteriaceae), symbiotic bacteria of entomopathogenic nematodes, and a proposal to transfer *Xenorhabdus luminescens* to a new genus, *Photorhabdus* gen". *International Journal of Systematic Bacteriology* 43.2 (1993): 249-255.
2. NEA Boemare., *et al.* "Symbiosis and pathogenicity of nematode-bacterium complexes". *Symbiosis* 22 (1997): 21-45.
3. ANB Chattopadhyay., *et al.* "Bacterial insecticidal toxins". *Critical Review in Microbiology* 30.1 (2004): 33-54.
4. PJ Daborn., *et al.* "A single *Photorhabdus* gene, makes caterpillars floppy (mcf), allows *Escherichia coli* to persist within and kill insects". *Proceedings of the National Academy of Sciences USA* 99.16 (2002): 10742-10747.
5. EG De-Gregorio., *et al.* "Enterobacterial repetitive intergenic consensus sequence repeats in yersiniae: genomic organization and functional properties". *Journal of Bacteriology* 187 (2005): 7945-7954.
6. E Duchaud., *et al.* "The genome sequence of the entomopathogenic bacterium *Photorhabdus luminescens*". *Nature Biotechnology* 21.11 (2003): 1307-1313.
7. SM Fischer-Le., *et al.* "Polyphasic classification of the genus *Photorhabdus* and proposal of new taxa: *P. luminescens* subsp. *luminescens* subsp. nov., *P. luminescens* subsp. *akurstii* subsp. nov., *P. luminescens* subsp. *laumondii* subsp. nov., *P. temperata* sp. nov., *P. temperata* subsp. *P. temperata* subsp. nov., and *P. asymbiotica* sp. Nov". *International Journal of Systematic Bacteriology* 49.4 (1999): 1645-1656.
8. JG Gerrard., *et al.* "Nematode symbiont for *Photorhabdus asymbiotica*". *Emerging Infectious Diseases* 12.10 (2006): 1562-1564.
9. RH ffrench-Constant. "Insecticidal toxins from *Photorhabdus* bacteria and their potential use in agriculture". *Toxicon* 49.4 (2007): 436-451.
10. J Gerrard., *et al.* "Human infection with *Photorhabdus asymbiotica*: An emerging bacterial pathogen". *Microbes Infections* 6.2 (2004): 229-237.
11. JG Gerrard., *et al.* "*Photorhabdus* Species: bioluminescent bacteria as emerging human Pathogens". *Emerging Infectious Diseases* 9.2 (2003): 251-254.
12. J Parkhill., *et al.* "Genome sequence of *Yersinia pestis*, the causative agent of plague". *Nature* 413.6855 (2001): 523-527.
13. NR Waterfield., *et al.* "Genomic islands in *Photorhabdus*". *Trends in Microbiology* 10.12 (2002): 541-545.
14. A Arunee., *et al.* "PirAB Toxin from *Photorhabdus asymbiotica* as a larvicide against dengue Vectors". *Applied and Environmental Microbiology* 75.13 (2009): 4627-4629.

Volume 13 Issue 4 December 2017

© All rights reserved by Ihsan Ulla.