Can Chorionic Villi Angiogenic Protein Expression Clarify the Vascular Abnormality Seen in Preeclampsia?

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Abstract

Objective: We have investigated the expression patterns of chorionic villi vascular endothelial growth factor $(VEGF)_{165}$, $VEGF_{165}$ b, matrix metalloproteinase (MMP)-9 and tumor necrosis factor alpha (TNF- α) proteins in normal pregnancies and preeclampsia to determine if the protein expression patterns could contribute in explaining the vascular abnormality seen in preeclampsia. Differences in the placental proteins between Black American and Hispanic groups were explored.

Methods: Term placentas from normotensive healthy women and placentas from women (37 weeks+ gestation) with preeclampsia, diagnosed by ACOG's criteria were investigated. Chorionic villi expressions of the four proteins were determined by ELISA; each ELISA kit using monoclonal antibody to the respective human proteins as capture antibody.

Results: Independent T test comparing normal pregnancy and preeclampsia showed placental VEGF₁₆₅ protein was higher in normal pregnancy (p = 0.001); VEGF₁₆₅b, MMP-9 and TNF- α proteins were higher in preeclampsia (p = 0.05, p = 0.036 and p = 0.037, respectively). Mothers' systolic and diastolic blood pressures were higher (p = 0.000), placental weight was comparable; and newborn weights were lower (p = 0.000) in preeclampsia. In normal pregnancy, the two VEGF165 isoforms were correlated (r = 0.261, p = 0.005) and VEGF₁₆₅ was negatively correlated to MMP-9 protein (r = -0.185, p = 0.047). Ethnicity was self-reported and the distribution was 23.3% of Black Americans and 69.8% of Hispanic American descent. Study variables differed between the ethnic groups.

Conclusion: Preeclampsia is a multifactorial disease. The findings suggest that placental angiogenic proteins may be contributory factors in the pathophysiology of the disease. Differences in VEGF₁₆₅ ratios, mothers' blood pressure and newborn weights were significantly different between normal pregnancy and preeclampsia, while placental weight was comparable, which suggests that placental angiogenic processes of the two groups may be different. The synergistic actions of VEGF₁₆₅ proteins in the third trimester of normal pregnancy, we suggest, may have favored sprouting angiogenesis maximizing blood flow; resulting in healthy newborns with normal weight. The imbalance in placental VEGF₁₆₅ proteins, we hypothesize, may be an angiogenic biomarker triggering angiogenesis to breakdown the extracellular matrix, to allow the capillary plexus to grow. Higher TNF- α protein in preeclampsia may be one of the compensatory mechanisms to assist the dilation of the blood vessels allowing greater amounts of blood to be transported to the fetus. The findings of this study additionally revealed that the correlations between the studied placental proteins differed significantly between the two ethnic groups.

Keywords: Pregnancy-Specific Protein Expressions; Normal Human Pregnancy; Preeclampsia, $VEGF_{165}$, $VEGF_{165}$ b, MMP-9 and TNF- α Proteins; Hispanic and Black Ethnic Groups; Ethnic Differences, Methods of Delivery, Fetal Gender

Abbreviations

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 $VEGF_{165}$: Vascular Endothelial Growth Factor (VEGF)_{165}; VEGF_{165}b: Vascular Endothelial Growth Factor (VEGF)_{165}b; MMP-9: Matrix Metalloproteinase-9; TNF- α : Tumor Necrosis Factor Alpha; GAD: Gestational Age in Days

Introduction

Preeclampsia is one of the oldest diseases on record [1], yet the etiology of preeclampsia is still unknown. Hence the quest for biomarkers continues with the hope that different biomarkers may help in understanding the underlying pathological process. Currently there is no suitable animal model to study the disease. Investigators have therefore relied on cell culture studies or have used fresh and cryogenically stored placental villous explants to identify biomarkers that might predict the syndrome [2,3].

A number of diverse theories on the causation of the disease have been put forward by many investigators over the years pertaining to oxidative stress, immunologic intolerance between the fetoplacental unit and the maternal tissue, and an angiogenic imbalance [4-7]. Regardless of the mechanisms, a two stage model of preeclampsia has been developed: in the first stage, reduced placental perfusion is thought to follow an abnormal implantation; and later failure in vascular remodeling is assumed to induce syncytiotrophoblast stress that interacts with maternal constitutional factors that trigger the maternal syndrome of preeclampsia [4-7].

Angiogenesis is one of the leading mechanisms in pregnancy that contributes to the exponential fetal growth throughout gestation; and placenta is the central offender in the pathophysiology of preeclampsia because delivery of the placenta can only reverse the syndrome. We, for several years, have investigated several placental angiogenic proteins independently in normal pregnancies and preeclampsia [8,9] in order to delineate the differences in the placental protein profiles between the two groups. In the present study we have simultaneously investigated four placental proteins: VEGF₁₆₅, VEGF₁₆₅b, MMP-9 and TNF- α protein in normal pregnancies and preeclampsia in order to understand if simultaneous quantitation of the proteins can shed additional light in the pathophysiology of the vascular defect responsible for the disease. The hospital where the study was conducted primarily serves women of Hispanic and Black American population. Hence, in the present study, we have also examined the differences in the placental proteins in the placental proteins between these two ethnic groups.

Methods

Women were not enrolled for the study. Placental tissues after clinical care following term deliveries from normotensive pregnancies that would otherwise have been discarded were collected under a protocol approved by the Institutional Human Subject Committee of the BronxCare Health System, Bronx, New York. The protocol also allowed collection of placentas from women with preeclampsia. The diagnosis of preeclampsia was made by the criteria of ACOG [10]. These women had no history of hypertension before pregnancy, their systolic and diastolic blood pressure was $\geq 140/90$ mm Hg on at least two occasions 6 hours apart, and urinary protein ≥ 300 mg/24 hr. The IRB approved protocol allowed certain clinical information to be collected at the time of tissue collection without the identification of the patients' names and their medical record numbers. These included: maternal age, race, gestational age as determined by ultrasound and/or by the initial date of the last menstrual period, and method of delivery. The excluding criteria for both the study groups included pregnancies complicated with any symptom of infection, diabetes, hypertension, chronic renal disease, chronic peripheral vascular disease, multifetal gestation or with major fetal anomalies. To keep the preeclampsia and the normotensive group comparable the gestational age of both groups was limited from 37 to 42 weeks. Placental tissues were collected within 30 minutes of vaginal or cesarean deliveries. Placentas delivered overnight or when placenta sample collection could not be done within the 30 minute window, the placentas were not collected to keep the sample collection protocol consistent.

The placental membranes were removed. Placental tissues were then thoroughly washed and dissected in saline to collect free floating chorionic villi, not anchored to the basal plate nor emerging from the chorionic plate surface vessels. Sections of chorionic villi samples

from the same placenta were placed in individual cryovials bearing identical study number and were transported to the laboratory on ice. The chorionic villi samples from the same placenta were stored in individual freezer boxes at -80°C until assay.

Commercially available Enzyme Immunoassay (EIA) kits were purchased from R&D Systems, Minneapolis, MN to determine the chorionic villi protein expressions of VEGF₁₆₅, VEGF₁₆₅b, MMP-9 and TNF- α . The four proteins were not analyzed from the same homogenate. VEGF₁₆₅ protein assays of all chorionic villi samples were carried out first. On the day of the assay, chorionic villi samples were taken out of the -80°C freezer in chronological order the placental tissues were collected. The tissues were homogenized, then centrifuged at 13,000 rpm for 2 minutes and the supernatants were used for direct-sandwich EIA. When VEGF₁₆₅ protein assays were completed, VEGF₁₆₅b protein expression was carried out following the same chronological order as stated above, followed by MMP-9 protein and then TNF- α assays. Sensitivity of the assay kit used was 31.3 pg/ml for VEGF₁₆₅, 62.5 pg/ml for VEGF₁₆₅b, 31.3 pg/ml for MMP-9, and 15.6 pg/ml for TNF α , protein, respectively. Samples were analyzed in duplicates and the intra assay and inter assay variations for all proteins were < 10%.

Statistical analyses

Statistical evaluation of the data was carried out using SPSS^R statistical package version 26 (IBM Corporation, Armonk, NY). Normal distribution of the data was first tested and was found to be skewed. Chorionic villi tissue samples were grouped based on the presence or absence of preeclampsia and Independent T test was performed to assess difference in means. Spearman's Rank correlation coefficient test was applied to estimate the strength and direction of relationships between pairs of continuous variables. P < 0.05 was considered significant.

Results and Discussion

The rationale for selecting the four proteins was because the multistep process of placental angiogenesis begins with a rise in local angiogenic growth factors. Cytotrophoblasts during human pregnancy secrete several VEGF proteins, of which VEGF₁₆₅ is considered as the key contributor of the angiogenic process [11]. VEGF₁₆₅b, the alternate spliced form of VEGF₁₆₅, is generated from the same transcript and is present in appreciable amounts in human placenta during pregnancy [9,12]. After the rise in local angiogenic growth factors, breakdown of decidual basement membrane follows, that facilitates cytotrophoblasts migration and proliferation [13] along with simultaneous remodeling of the spiral arteries. Remodeling of the extracellular matrix additionally facilitates maternal and placental tissue morphogenesis and fetal development [14] and healthy pregnancies are dependent on matrix metalloproteinases (MMPs) for trophoblast cell invasion and embryo implantation [13,14]. Placental TNF- α is a cell signaling protein that during pregnancy induces the synthesis of matrix metalloproteinases (MMPs), allows the cytotrophoblasts to reach the spiral arteries deeper within the uterine decidua and augments the apoptosis of vascular smooth muscle cells surrounding the spiral arteries [15].

In this study, a total of 146 placentas were collected, 115 placentas from women who were normotensive throughout gestation and delivered at term; and 31 from women who developed preeclampsia during their pregnancy and delivered after 37 weeks of gestation. The demographic characteristics of women from whom placental samples were collected are shown in table 1. The results show that the mothers from whom the placentas were collected were of similar age groups. Compared to normotensive women, mothers with preeclampsia had significantly shorter gestation (p = 0.001). Both the systolic and diastolic blood pressures of women in the preeclampsia group were higher (p = 0.001). The mean newborn weights in the preeclampsia group was lower (p = 0.001). However, the placental weight was comparable between the two groups $459 \pm 91g$ versus $417 \pm 138g$ (Table 1). Race/ethnicity was self-reported and the distribution pattern showed that women in the study were 23.2% of Black American origin; 69.8% were of Hispanic American origin, and the remaining were of other ethnic groups. The two stage model of preeclampsia suggests that a poorly perfused placenta that occurs in Stage 1 is not sufficient to cause the maternal syndrome of preeclampsia [4-7]. Factors that are produced in poorly perfused placenta in

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Stage 1 interact with maternal constitutional factors (genetic, behavioral or environmental) to result in Stage 2 [4-7]. Hence our study population of 69.8% Hispanic and 23.3% of Black Americans provided us with the opportunity to crudely explore if ethnic variations do contribute to the pathophysiology of preeclampsia. In the study, the number of placentas collected following spontaneous vaginal delivery and cesarean sections were 58 and 88, respectively.

Variables	Groups	N	Mean	Std. Dev	Sig. (2-tailed) Equal variance not assumed
(AD (in days)	Normotensive	115	276	7	0.02
GAD (III days)	Preeclampsia	31	259	26	0.02
	Normotensive	115	28	6	0.104
Age (years)	Preeclampsia	31	26	6	0.194
Systolic BP (mm Hg)	Normotensive	115	120	15	0.000
	Preeclampsia	31	153	19	0.000
Diostolio DD (mm Ho)	Normotensive	115	71	11	0.000
Diastolic BP (mm Hg)	Preeclampsia	31	92	13	0.000
Placental weight (grams)	Normotensive	115	459	99	0.320
	Preeclampsia	31	426	141	0.229
Newborn weight (grams)	Normotensive	115	3402	503	0.000

Table 1: Characteristics related to mothers from whom placentas were collected.

Results of independent T test that compared the protein expressions of the two groups are presented in table 2 and in figure 1. VEGF₁₆₅ and VEGF_{155} proteins were both identified in the third trimester of normotensive (n = 115) and preeclamptic (n = 31) placentas. The results showed that compared to the normotensive group the mean pro-angiogenic VEGF₁₆₅ protein expression in preeclamptic chorionic villi samples was significantly lower (p = 0.001) while the anti-angiogenic VEGF₁₆₅b protein expression was significantly higher (p = 0.001) 0.05), in contrast to the result of an earlier published report [12]. In this study a total of 115 normal and 31 preeclamptic placentas were analyzed to determine VEGF₁₆₅ and VEGF₁₆₅b protein expressions; and the ELISA method used was specific to determine human VEGF₁₆₅ and VEGF₁₆₅b protein independently. Moreover, chorionic villi samples were isolated from each placenta to determine particularly the VEGF₁₆₅ protein expressions of the cytotrophoblasts. MMP-9 and TNF- α protein expressions were both found to be significantly higher in the preeclampsia group (Table 2 and figure 1). Our findings are consistent with the report of Wang and Walsh [16] who reported a significantly higher expression of TNF- α protein in preeclamptic placentas. A higher concentration of TNF- α protein in the maternal circulation in preeclampsia was reported in another study. The investigators claimed that the higher concentration of $TNF-\alpha$ protein seen in preeclampsia was not caused by the disease but was the consequence of the disease because the higher expression of $TNF-\alpha$ only occurred after the clinical signs of disease were established [17]. Another study cited in the literature reported that TNF- α was not elevated in preeclamptic placenta despite its elevation in peripheral blood [18]. The expression of TNF- α protein in the study was determined using monoclonal antibody based ELISA kit just like ours but the earlier study differed from our study in several ways. First, the placental tissue used for ELISA included both maternal and fetal surfaces while in the present study placental chorionic villi were isolated to determine the TNF- α protein levels. Second, TNF- α protein concentrations in the previous study were expressed as TNF- α levels relative to total protein, while we had expressed our results as TNFlpha concentration per 100 mg wet tissue. The significantly higher concentrations of MMP-9 protein in preeclamptic placentas in this study (p = 0.036) is also in contrast with reports of lower concentration of MMP-9 protein in preeclamptic placentas [19]. Possible explanations for the discrepancies in the results could be due to the difference in the methods used, the type of tissue that were analyzed, serum versus placenta; or because the gestational age of the preeclampsia group did not match. Table 1 also shows that the maternal systolic and diastolic blood pressures in preeclampsia group were significantly

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higher (p = 0.0001) but the newborn weights were significantly lower (p = 0.001); consistent with a previous report [19]. It needs to be underscored that the placental weights in the study were comparable between normal pregnancy and preeclampsia groups $459 \pm 91g$ versus $417 \pm 138g$ (Table 1), and the finding is in agreement with a large Norwegian study which reported placental weight in normal pregnancies (n = 304,875) to be 672g, and in term preeclampsia (n = 9,743) with gestational age greater 37 weeks to be 675g [20].

Variables	Groups	N	Mean	Std. Dev	Sig. (2-tailed) Equal variance not assumed
VEGF ₁₆₅ (pg/100 mg	Normotensive	115	153.64	101.03	
tissue)	Preeclampsia	31	89.61	56.47	0.000
VEGF ₁₆₅ b (pg/100	Normotensive	115	341.67	231.11	
mg tissue)	Preeclampsia	31	448.03	267.79	0.05
MMP-9 (ng/100 mg	Normotensive	115	26.08	16.30	
tissue)	Preeclampsia	31	33.11	16.00	0.036
TNF-α (pg/100 mg tissue)	Normotensive	115	36.84	24.72	
	Preeclampsia	31	57.48	51.20	0.027
	Preeclampsia	31	2723	867	0.037

Table 2: Chorionic villi protein expressions.



Figure 1: Chorionic villi $VEGF_{165}$, $VEGF_{165}$, MMP-9 and TNF- α protein expression of the two study groups are shown here. Expression of each chorionic villi protein significantly differed between normal pregnancy (n = 115) and preeclampsia (n = 31) groups.

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In this study, Spearman Rank correlation coefficient test was used to determine the correlations between the ten studied variables. The correlations in the normotensive and preeclampsia groups were separately analyzed. Table 3 depicts the correlations between the studied variables in normal placentas (n = 115). The positive correlation in the expressions of VEGF₁₆₅ and VEGF₁₆₅ b placental proteins (r = 0.280, p = 0.002), suggest a synergistic action between the two proteins in the third trimester of human pregnancy, consistent with a previously published results [12]. To our knowledge placental expression of VEGF₁₆₅ and VEGF₁₆₅ b proteins in human pregnancy has not been reported elsewhere other than the one already cited in the manuscript. Table 3 also shows that in normal pregnancy the systolic and diastolic blood pressures of the mothers from whom the placentas were taken were significantly correlated (r = 0.571, p = 0.000). A significant positive correlation was also noted between placental and newborn weights (r = 0.557, p = 0.000). A relevant feature of Table 3 is the negative correlation seen between VEGF₁₆₅ and MMP-9 protein (r = -0.185, p = 0.047) (Table 3). Regulation of angiogenesis is a multiaten precess that requires activation and preliferation of and the placental cells and degradation of actracellular matrix.

is a multistep process that requires activation and proliferation of endothelial cells and degradation of extracellular matrix, enabling the proliferating cells to invade the surrounding tissue. During tumorigenesis, basement membrane destruction is an essential step for tumor invasion and metastases, and cellular invasion is dependent on MMP-9 protein [21]. A transgenic mouse model of multistage carcinogenesis showed that MMP-9 protein can explicitly render quiescent normal pancreatic islets cells to become angiogenic, by inducing the release of VEGF protein [22]. The authors confirmed that these results could not be replicated when other molecules were used. The study additionally showed that when the activity of MMP-9 protein was inhibited by MMP-9 inhibitors, the angiogenic switching was reduced, concomitant with a decrease in the number of tumors and in tumor growth. Genetic ablation of MMP-9 produced similar results [22]. To summarize, the study showed that during tumorigenesis it is MMP-9 protein that induced the release of VEGF to trigger the angiogenic process. Cytotrophoblasts during pregnancy similarly mirrors to a certain extent the behavior of malignant cells. The difference between tumorigenesis and pregnancy is that while the invasion and angiogenic processes in tumorigenesis continues, in pregnancy these processes after a certain period of time need to cease. Hence, the significant negative correlation seen in the third trimester of normal pregnancy between VEGF₁₆₅ and MMP-9 protein (r = -0.185, p = 0.047, Table 3) may reflect an inherent physiological phenomenon in human pregnancy to terminate the angiogenic process.

Variables		VEGF ₁₆₅	VEGF ₁₆₅ b	MMP-9	TNF-α	GAD	Sys Pr	Dia Pr	Pl Wt	NB Wt
VEGF (pg/100	Corr Coeff	1.000	0.261	-0.185	0.137	0.031	0.019	-0.042	0.009	-0.037
tissue)	Sig (2-Tailed)		0.005	0.047	0.143	0.744	0844	0.653	0.925	0.697
	N	115	115	115	115	115	115	115	115	115
VEGF ₁₆₅ b (pg/100 mg	Corr Coeff	0.261	1.000	-0.020	-0.002	-0.028	0.020	0.083	0.049	0.128
	Sig (2-Tailed)	0.005		0.834	0.979	0.765	0.832	0.376	0.603	0.173
tissue)	N	115	115	115	115	115	115	115	115	115
MMP-9 (ng/100	Corr Coeff	-0.185	-0.020	1.000	-0.080	-0.031	-0.064	-0.114	-0.060	-0.146
mg	Sig (2-Tailed)	0.047	0.834		0.397	0.743	0.494	0.226	0.528	0.119
tissue)	N	115	115	115	115	115	115	115	115	115
	Corr Coeff	0.137	-0.002	-0.080	1.000	-0.030	0.163	0.118	0.053	0.078
TNF-α (pg/100 mg tissue)	Sig (2-Tailed)	0.143	0.979	0.397		0.754	0.081	0.209	0.574	0.406
	N	115	115	115	115	115	115	115	115	115
	Corr Coeff	0.031	028	-0.031	-0.030	1.000	007	0.096	-0.023	.187
Gestationa age in days	Sig (2-Tailed)	0.744	0.765	0.743	0.754		0.938	0.308	0.810	0.045
	N	115	115	115	115	115	115	115	115	115
Systolic Blood	Corr Coeff	0.019	0.020	-0.064	0.163	-0.007	1.000	.571	0.008	0.116
Pressure (mm Hg)	Sig (2-Tailed)	0.844	0.832	0.494	0.081	0.938		0.000	0.930	0.217
	N	115	115	115	115	115	115	115	115	115
Diastolic Blood	Corr Coeff	-0.042	0.083	-0.144	0.118	0.096	.571	1.000	-0.110	-0.051
Pressure (mm	Sig (2-Tailed)	0.653	0.376	0.226	0.209	0.308	0.000		0.241	0.585
Hg)	N	115	115	115	115	115	115	115	115	115
	Corr Coeff	0.009	0.049	-0.060	0.053	-0.023	0.008	-0.110	1.000	0.557
P WT Weight (gms)	Sig (2-Tailed)	0.925	0.603	0.528	0.574	0.810	0.930	0.241		0.000
	N	115	115	115	115	115	115	115	115	115
	Corr Coeff	-0.037	0.128	-0.146	0.078	0.187	0.116	-0.051	0.557	1.000
NB WT Weight	Sig (2-Tailed)	0.697	0.173	0.119	0.406	0.045	0.217	0.585	0.000	
(giiis)	N	115	115	115	115	115	115	115	115	115

Table 3: Spearman's rank correlation coefficient results of studied variables in normal pregnancy.

Corr Coeff: Correlation Coefficient; GAD: Gestational Age in Days; Sys Pr: Systolic Blood Pressure;

Dia Pr: Diastolic Blood Pressure; Pl WT: Placental Weight; NB WT: Newborn Weight.

The Spearman's Rank coefficient correlation data between the 10 studied variables in preeclamptic placentas are presented in table 4. In contrast to normotensive pregnancies, the two VEGF₁₆₅ and VEGF₁₆₅b proteins in preeclamptic placentas were not correlated. This lack of correlation between VEGF₁₆₅ isoforms we interpret as an imbalance in VEGF₁₆₅ isoforms in preeclampsia that disrupts the normal angiogenic course. Since the placental weight between normal pregnancy and preeclampsia was comparable (Table 1) hence, it suggests that the angiogenic process in preeclamptic placenta continued, but the intracellular signaling mechanisms driving angiogenesis must have been different. The synergistic action between the two VEGF₁₆₅ proteins in normal pregnancy may act as a directional signal influencing capillary growth. The synergistic actions of the two VEGF₁₆₅ isoforms, we suggest, may modify the placental angiogenesis in favor of the sprouting type that allowed new branches to be formed on existing blood vessels and maximized the amount of blood that flowed through, to result in healthy newborns with mean weight of 3402g. The imbalance in the two VEGF₁₆₅ proteins in preeclampsia, we hypothesize, may be an angiogenic biomarker that triggered angiogenesis of the intussusceptive type. In contrast to sprouting angiogenesis, during intussusceptive angiogenesis each blood vessel splits into two [23]; the caliber of each blood vessels become reduced to half its size, and progressively decrease further with each subsequent division [23]. Such progressive decrease in blood vessel diameter, we hypothesize ultimately leads to ischemia and vascular dysfunction consistent with preeclampsia. In the present study, the MMP-9 protein expression is significantly higher in preeclamptic placenta (Table 2). If the concept of intussusceptive angiogenesis holds, the angiogenic process will yield capillaries that are much reduced in diameter. Hence, to intensify the amount of blood that flows to the fetus, because the fetal demand surges in the third trimester; the capillary plexus need to grow at a much faster rate in comparison to sprouting angiogenesis. The significantly higher MMP-9 protein expression seen in preeclamptic placentas (Table 1), we suggest, supports the extracellular matrix to rapidly degrade to make room for the capillary plexus to grow. A significant correlation between placental weight and placental TNF-α protein was seen in preeclampsia (r = 0.385, p = .032), underscoring the support of locally produced cytokine in placental health in preeclamptic condition. Additionally, in preeclampsia, GAD was significantly correlated to placental and newborn weights (r = 0.508, p = 0.004; r = 0.781, p = 0.001, respectively). The finding led us to suggest that perhaps the newborn outcome could be better if the gestational age of the women in the preeclampsia group is allowed to stretch.

Variab	les	VEGF ₁₆₅	VEGF ₁₆₅ b	MMP-9	TNF-α	GAD	Sys Pr	Dia Pr	Pl Wt	NB Wt
	Corr Coeff	1.000	0.291	0.097	0.304	-0.255	-0.314	-0.022	0.148	-0.138
VEGF ₁₆₅ (pg/100 mg tissue)	Sig (2-Tailed)		0.113	0.605	0.096	0.165	0.085	0.907	0.428	0.459
	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	0.291	1.000	0.027	0.036	-0.299	0.102	0.181	-0.059	-0.419
VEGF ₁₆₅ b (pg/100 mg	Sig (2-Tailed)	0.113		0.887	0.848	0.103	0.584	0.329	0.754	0.019
	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	0.097	0.027	1.000	0.049	0.071	0.175	0.269	0.303	0.022
MMP-9 (ng/100 mg	Sig (2-Tailed)	0.605	0.887		0.793	0.706	0.346	0.143	0.097	0.908
ussuej	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	0.304	0.036	0.049	1.000	-0.077	-0.313	0.066	0.385	0.033
TNF-α (pg/100 mg tissue)	Sig (2-Tailed)	0.096	0.848	0.793		0.680	0.087	0.724	0.032	0.859
	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	-0.255	-0.299	0.071	-0.077	1.000	0.009	-0.252	0.508	0.781
Gestational age in days	Sig (2-Tailed)	0.165	0.103	0.706	0.680		0.961	0.172	0.004	0.000
	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	-0.314	0.102	0.175	-0.313	0.009	1.000	.744	0.031	-0.076
Systolic Blood Pressure (mm Hg)	Sig (2-Tailed)	0.085	0.584	0.346	0.087	0.961		0.000	0.867	0.683
	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	-0.022	0.181	0.269	0.066	-0.252	0.744	1.000	-0.087	-0.257
Diastolic Blood Pressure (mm	Sig (2-Tailed)	0.907	0.329	0.143	0.724	0.172	0.000		0.643	0.163
ngj	Ν	31	31	31	31	31	31	31	31	31
P WT Weight (gms)	Corr Coeff	0.148	-0.059	0.303	0.385	0.508	0.031	0.087	1.000	0.592
	Sig (2-Tailed)	0.428	0.754	0.097	0.032	0.004	0.867	0.643		0.000
	Ν	31	31	31	31	31	31	31	31	31
	Corr Coeff	-0.138	-0.419	0.022	0.033	0.781	-0.076	-0.257	0.592	1.000
NB WT Weight (gms)	Sig (2-Tailed)	0.459	0.019	0.908	0.859	0.000	0.683	0.163	0.000	
	Ν	31	31	31	31	31	31	31	31	31

Table 4: Spearman's rank correlation coefficient results of studied variables in preeclampsia.

 Corr Coeff: Correlation Coefficient; GAD: Gestational Age in Days; Sys Pr: Systolic Blood Pressure;

 Dia Pr: Diastolic Blood Pressure; Pl WT: Placental Weight; NB WT: Newborn Weight.

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In this study, of the total 146 placentas that were collected, 101 placentas were of mothers of Hispanic American origin (69.8%), 34 were from mothers of Black American descent, and the remaining 11 were from other ethnic backgrounds. Out of 34 women from the Black American ethnic group, 15 had preeclampsia, and out of 101 women from the Hispanic American ethnic group 14 had preeclampsia. However, we need to interpret the results of this study as Black American women being at a higher risk of preeclampsia compared to Hispanic mothers with caution. The reason being, that in the study there was a limitation in our tissue collection process. We had a 30 minute window limit from the time of delivery and placental sample collection. Placentas that were delivered outside this 30 minute window or were delivered overnight were not collected. And perhaps this limitation might have affected the result. Nonetheless, higher incidence of preeclampsia among Black American women is consistent in other reports [24,25]. In a recent study that included 6000 women of racially diverse groups it was stated that race alone was unlikely to account for the higher rates of preeclampsia among Black women, and other biological, social and cultural factors may contribute to the process [26].

Previous studies examining the risk factors of preeclampsia have compared ethnic differences in maternal blood constituents [27,28]. To our knowledge this is perhaps the first study that investigated the differences in placental angiogenic proteins between two ethnic groups, Black and Hispanic population. Figure 2-4 summarize the differences in the studied variables between the two ethnic groups and reveal that ethnic differences in placental angiogenic proteins do exist. Black women who had preeclampsia (n = 15) showed a significant positive correlation between placental VEGF₁₆₅ and TNF- α protein (r = 0.571, p = 0.026). In an *in vitro* study with HUVEC cells, varying the concentration of VEGF available to the growing vessel was shown to provide the control to regulate the diameter of the vessels. The authors suggested that VEGF may dilate vessels as a result of thinning of the endothelial cells or by recruiting endothelial cells within them [29]. During pregnancy, nitric oxide synthase activity is highest in the villous trophoblast of term placenta [30]. In an in vivo model, it was shown that application of 100 ng/ml of TNF- α protein added to cerebrospinal fluid to rats caused marked progressive nitric oxide-mediated dilation of cerebral arterioles with a maximum increase in diameter at 4 hours [31]. In preeclampsia, the caliber of the blood vessels is significantly reduced; and the trophoblast invasion of the spiral arteries is incomplete, hence, the endothelial and smooth muscle cells are retained within the spiral arteries rather than being replaced by cytotrophoblasts. Based on the findings of an upregulation of TNF- α protein in preeclampsia (Table 2), along with a significant positive correlation seen between VEGF₁₆₅ and TNF- α proteins (r = 0.571, p = 0.026) we suggest, that the upregulation in TNF- α protein could be a compensatory mechanism in preeclampsia to assist the reduced caliber blood vessels to dilate; so that greater amounts of blood can be transported to the fetus, to keep up with the increase in fetal demand of the third trimester. Black women who had preeclampsia also had significant correlations between MMP-9 protein and GAD (r = 0.592, p = 0.02); MMP-9 protein and placental weight (r = 0.775, p = 0.001) and MMP-9 protein and newborn weight (r = 0.568, p = 0.027). Among Black American with preeclampsia, the chorionic villi anti-angiogenic VEGF₁₆₅b protein expression was significantly higher (p = 0.049) compared to their normotensive group. Women of Hispanic origin who had preeclampsia were significantly younger (28 ± 6 years versus 25 ± 4 years, p = 0.03) and the chorionic villi pro-angiogenic VEGF₁₆₅ protein expression was significantly lower (p = 0.01) compared to the normotensive group. Thus, the findings of this study revealed that the correlations between the studied placental proteins differed between the two ethnic groups. Overall, the data of the present study supports the notion that ethnic variations can impact the maternal constitutional factors differently in triggering the maternal syndrome of preeclampsia. We think a greater understanding of the relationship of known biomarkers and ethnicity of women with preeclampsia may provide fresh insight into the pathophysiology of the disease.



Figure 2: Differences in the mother's and diastolic blood presure; and placental weight and newborn weights of the two ethnic groups are shown. The ethnic groups from whom the placentas were collected comprised of: 19 Black women in the normotensive group and 15 Black women who had preeclampsia; and 87 Hispanic women in the normotensive group and 14 Hispanic women who had preeclampsia.



Figure 3: Differences in the mothers' systolic and diastolic blood pressure; and placental weight and newborn weights of the two ethnic groups are shown. The ethnic groups from whom the placentas were collected comprised of: 19 Black women in the normotensive group and 15 Black women who had preeclampsia; and 87 Hispanic women in the normotensive group and 14 Hispanic women who had preeclampsia.



Figure 4: Differences in Mothers' gestational age and mothers' age between the two ethnic groups are shown here. The ethnic groups from whom the placentas were collected comprised of: 19 Black women in the normotensive group and 15 Black women who had preeclampsia; and 87 Hispanic women in the normotensive group and 14 Hispanic women who had preeclampsia.

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In this study, the higher number of placentas collected following cesarean sections (n = 88) manifests the ease of collecting placental samples in a timely manner from a scheduled cesarean section than collecting placentas following spontaneous vaginal delivery (n = 58). Independent T test results revealed that women who had cesarean section were significantly older than those who delivered vaginally (29 ± 6 years versus 25 ± 6 years, p = 0.01). The systolic and diastolic blood pressures of the mothers who delivered vaginally were 131 ± 24 mm Hg and 79 ± 15 mm Hg, respectively; whereas the systolic and diastolic blood pressures of mothers who delivered by cesarean sections were 124 ± 18 mm Hg and 73 ± 13 mm Hg, respectively, the differences were statistically significant p = 0.041 and p = 0.012, respectively). Expressions of the four placental proteins were not affected by the methods of delivered male newborns. The studied variables were examined to determine if the gender of the fetus had any effect on the studied variables. Results showed that in mothers who carried a male fetus, the placental weight was 434 $\pm 105g$ and in mothers who carried a female fetus the placental weight was 470 $\pm 112g$ and the differences in placental weight was statistically significant (p = 0.05). Differences in the other studied variables were not significant.

Conclusion

In the present study we investigated whether comparison of chorionic villi angiogenic protein expression patterns between normotensive pregnancies and preeclampsia could elucidate the vascular abnormality seen in preeclampsia. Significant differences were found in: a) mean VEGF₁₆₅/ VEGF₁₆₅b ratios (p = 0.0001), b) mothers' systolic and diastolic blood pressure (p = 0.000, p = 0.000, respectively); and in newborn weights (p = 0.00) between normal pregnancies and preeclampsia. Yet, the placental weight between the two groups was comparable suggesting that the intracellular signaling mechanisms driving angiogenesis in the two studied groups may have been different. The synergistic action between the two VEGF₁₆₅ proteins in normal pregnancy may act as a directional signal influencing capillary growth modifying the placental angiogenesis in favor of the sprouting type. Formation of new branches on the existing blood vessels may have maximized the amount of blood flow, resulting in healthy newborns with mean weight of 3402g. The imbalance in the two VEGF₁₆₅ proteins in preeclampsia may be an angiogenic biomarker that triggered angiogenesis of the intussusceptive type. Since during intussusceptive angiogenesis each blood vessel splits into two, the caliber of each blood vessel progressively decrease. Such progressive decrease in blood vessel diameter, we hypothesize ultimately leads to ischemia and vascular dysfunction consistent with preeclampsia. If the concept of intussusceptive angiogenesis holds, the angiogenic process will yield capillaries that are much reduced in diameter. Hence, to intensify the amount of blood that flows to the fetus, as the fetal demand surges in the third trimester; the capillary plexus need to grow at a much faster rate in comparison to sprouting angiogenesis. It may be suggested that the higher expression of chorionic villi MMP-9 protein seen in the present study in preeclampsia is as a compensatory mechanism to support intussusceptive angiogenesis and rapid degradation the extracellular matrix to allow the capillary plexus to grow. Higher expression of TNF-α protein in preeclamptic placenta, we suggest, could also be a compensatory mechanism, to assist the spiral arteries to dilate; so that greater amounts of blood can be transported to the fetus, to keep up with the surge in fetal demand of the third trimester.

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Conflict of Interest

The authors declare no conflicts of interest with respect to the research, authorship and/or publication of this article.

Bibliography

- 1. Waite LL., et al. "Preeclampsia, an implantation disorder". Reviews in Endocrine and Metabolic Disorders 3.2 (2002): 151-158.
- 2. McNally R., *et al.* "Elucidating the pathogenesis of pre-eclampsia using in vitro models of spiral uterine artery remodeling". *Current Hypertension Reports* 19.11 (2017): 1-13.
- 3. Orendi K., *et al.* "Placental and trophoblastic in vitro models to study preventive and therapeutic agents for preeclampsia". *Placenta* 32.1 (2011): S49-S54.
- 4. Maarten TM., et al. "Oxidative stress and preeclampsia". Hypertension 44.4 (2004): 374-380.
- 5. Roberts JM and Hubel CA. "The two stage model of preeclampsia: Variations on the theme". Placenta 23 (2009): S32-S37.
- 6. Staff AC. "The two-stage placental model of preeclampsia: An update". Journal of Reproductive Immunology 134-135 (2019): 1-10.
- 7. Roberts JM and Bell MJ. "If we know so much about preeclampsia, why haven't we cured the disease?" *Journal of Reproductive Immunology* 99.0 (2013): 1-9.
- 8. Basu J., *et al.* "Correlation between placental matrix metalloproteinase -9 and tumor necrosis factor -α protein expression throughout gestation in normal human pregnancy". *Reproductive Sciences* 25.4 (2018): 621 -627.
- 9. Basu J., *et al.* "Chorionic villi expression of two vascular endothelial growth factor proteins in normal human pregnancy". *Integrative Gynecology and Obstetrics Journal* 1.3 (2018): 1-6.
- 10. American College of Obstetricians and Gynecologists. Hypertension in Pregnancy. Washington, DC (2013).
- 11. Cheung CY. "Vascular endothelial growth factor: Possible role in fetal development and placental function". *Journal of the Society for Gynecologic Investigation* 4.4 (1997): 167-177.
- Bates DO., et al. "The endogenous anti-angiogenic family of splice variants of VEGF, VEGFxxxb, are down-regulated in pre-eclamptic placentae at term". Clinical Science 110.5 (2006): 575-585.
- 13. Cohen M., et al. "Metalloproteinases and human placental invasiveness". Placenta 27.8 (2006): 783-793.
- 14. O'Connor BB., *et al.* "The role of extracellular matrix in normal and pathological pregnancy: Future applications of microphysiological systems in reproductive medicine". *Experimental Biology and Medicine* 245.13 (2020):1163-1174.
- 15. Basu J., *et al.* "Placental tumor necrosis factor-α protein expression during normal human gestation". *Journal of Maternal-Fetal and Neonatal Medicine* 29.4 (2016): 3934-3938.
- Wang Y and Walsh S. "TNF-α concentrations and mRNA expression are increased in preeclamptic placentas". Journal of Reproductive Immunology 32.2 (1996): 157-169.
- 17. Vince GS., *et al.* "Interleukin-6, tumor necrosis factor and soluble tumor necrosis factor receptors in women with pre-eclampsia". *BJOG: An International Journal of Obstetrics and Gynaecology* 102.1 (1995): 20-25.
- Hayashi M., *et al.* "Tumor necrosis factor –α in the placenta is not elevated in preeclamptic placentas despite its elevation in peripheral blood". *American Journal of Reproductive Immunology* 53.3 (2005): 113-119.
- 19. Sahay AS., *et al.* "Matrix metalloproteinases-2 (MMP-2) and metalloproteinases-9 (MMP-9) are differentially expressed in different regions of normal and preeclampsia placentae". *Journal of Cellular Biochemistry* 119.8 (2018): 6657-6664.

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- 20. Dahlstrom B., et al. "Placenta weight in preeclampsia". Act Obstetricia et Gynecologica 87.6 (2008): 608-611.
- 21. Huang Hao. "Matrix metalloproteinase-9 (MMP-9) as a cancer biomarker and MMP-9 biosensors: Recent advances". *Sensors* 18.10 (2018): 3249.
- 22. Bergers G., *et al.* "Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis". *Nature Cell Biology* 2.10 (2000): 737-744.
- 23. Djonov V Baum O and Burri PH. "Vascular remodeling by intussusceptive angiogenesis". *Cell and Tissue Research* 314.1 (2003): 107-117.
- 24. Breathett K., *et al.* "Differences in preeclampsia rates between African American and Caucasian women: trends from the national hospital discharge survey". *Journal of Women's Health* 23.11 (2014): 886-893.
- 25. Franki R. "Preeclampsia/eclampsia rate highest in black women". News from the FDA/CDC (2017).
- 26. Boakye E., *et al.* "Nativity-related disparities in preeclampsia and cardiovascular risk among a racially diverse cohort of US women". *JAMA Network Open* 4.12 (2021): e2139564.
- 27. Patrick TR., *et al.* "Homocysteine and folic acid are inversely related in Black women with preeclampsia". *Hypertension* 43.6 (2004): 1279-1282.
- 28. Yang J., et al. "Racial differences in midtrimester maternal serum levels of angiogenic and antiangiogenic factors". American Journal of Obstetrics and Gynecology 215.3 (2016): 359.e1-e9.
- 29. Nakatsu MN., *et al.* "VEGF121 and VEGF165 regulate blood vessel diameter through vascular endothelial growth factor receptor2 in a in vitro angiogenesis model". *Laboratory Investigation* 83.12 (2003): 1873-1885.
- 30. Al-Hijji J., et al. "Nitric acid synthase activity in human trophoblast, term placenta and pregnant myometrium". *Reproductive Biology* and Endocrinology 1.51 (2003): 1-7.
- 31. Brian JEJr and Faraci FM. "Tumor necrosis factor- α -induced dilation of cerebral arterioles". Stroke 29.2 (1998): 509-515.

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