

Managements of Immature Apex: A Review

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Abstract

This paper reviews the rationale and techniques for treatment of the non-vital immature tooth. The importance of careful case assessment and accurate pulpal diagnosis in the treatment of immature teeth with pulpal injury cannot be overemphasized. The treatment options starts from indirect pulp capping, direct pulp capping, partial pulpotomy, full pulpotomy, apexogenesis, apexification and ending up with revitalization. Describing the techniques, indications and the drawbacks of each treatment option.

Keywords: Immature Apex; Root Development; Pulp Capping; Apexogenesis; Apexification

Root Development

Root development commences after the completion of enamel formation. The cells of inner and outer enamel epithelia unite at point forming cervical loop, begin to proliferate and form a structure known as the Hertwig epithelial sheath. This sheath determines the size and shape of root/s of the tooth [1]. Apical root closure is completed approximately 2 - 3 years after tooth eruption [2].

Causes of pulpal disease

The major irritants of pulp tissue include various bacteria, trauma, dental procedures generating thermal stimulation and chemical agent. Irritation of pulp tissue results in major changes in pulp microcirculation that can lead to pulp necrosis and arrest root formation [3].

Dental caries is the most prevalent microbial infectious disease. As the caries lesion approximates the pulp, there is acute exacerbation of the precedent chronic inflammation characterized by an influx of neutrophils [4]. In the presence of severe inflammation, focal micro-abscesses form and eventually coalesce leading to progressive pulpal necrosis.

Restorative procedures can also affect pulpal tissue. Thermal injury may occur as result of tooth preparation or finishing procedures using dry cutting, dull burs, deep periodontal curettage, orthodontic movement, or lasers and air abrasion devices [5].

Chemical irritants of pulp include cavity cleansers such as alcohol, chloroform and hydrogen peroxide as well as some substance in restorative materials and cavity liner [6]. One key requirement of a successful restorative procedure is to cause minimal additional irritation of the pulp so as not to interfere with normal pulpal healing.

Crown-fracture teeth with or without pulp exposure and associated luxation injury experience a greater frequency of pulp necrosis [7]. However immature permanent tooth has considerable capacity to heal after traumatic pulp exposure, so conservative pulp therapies are important to preserve pulp vitality and allow for continued root formation.

Clinical tests to determine pulp condition

Pulpal conditions and stage of root development are the major factors in the selection of treatment plan [8]. However with early diagnosis and intervention, pulp preservation strategies promote an environment for continued dentine apposition and root formation.

Diagnosis begins with a careful medical examination and any implications related to the anticipated treatment. A Thorough dental history including all symptoms and characteristics of associated pain is essential.

Visual examination of both soft and hard tissue for the presence or absence of swelling, crown discoloration and caries.

Mobility and periodontal probing can provide information regarding the health of status pulp.

Electric and thermal tests are of limited value due to their varied responses in permanent teeth with immature apex [9].

After traumatic injuries electric and thermal pulp tests may be unreliable, only generalized impressions may be gained from these tests [10].

Attempts have been made to use laser Doppler flowmetry (LDF) for measurement of blood flow in traumatized teeth as this would provide more accurate readings [11].

The pulse oximeter also offers accurate means of monitoring pulp vitality by recording the oxygenation of pulpal flow [12].

Radiographic examination of teeth requires good quality periapical and bitewing radiographs. These radiographs reveal the status of periapical tissues, presence and proximity of pulpal caries and stage of root development. The use of cone beam computed tomography should provide more accurate information regarding the condition of periapical tissues and root formation compared to 2-dimension conventional radiographs [13].

However it is still difficult to determine if the pulp is reversibly or irreversibly affected [14]. Moreover children may have unreliable clinical symptoms and exaggerated responses to percussion, palpation and pulp tests that do not correlate well with histopathological condition of pulp in immature teeth [15].

A recent systematic review [16] aimed to appraise the diagnostic accuracy of sign/symptoms, pulp tests to determine the condition of pulp in teeth affected by deep caries, trauma and other types of injuries showed that there is insufficient evidence to assess the value of toothache or abnormal reaction to hot/cold stimulation for determining pulp condition.

However it is hoped that by combining the result of history, examination and diagnostic tests, an accurate clinical diagnosis of pulp vitality can be made in most cases.

Treatment of teeth with vital pulp and open apices

When pulp exposure occurs in immature tooth due to caries or trauma, the exposed pulp can heal if protected from further injury.

Treatment of immature teeth has changed dramatically in recent years as new concepts and materials have developed. Vital pulp therapies are the treatment of choice for traumatized and carious teeth with vital pulps and open apices [17]. The approaches include indirect pulp capping in deep caries cavities and direct pulp capping or pulpotomy in cases of pulp exposure.

The key to the success of vital pulp therapy might partly be strict case selection and proper treatment protocol. Vital pulp therapy should not only be performed in teeth with signs and symptoms of reversible pulpitis [72]. This may be against the traditional school of thought that vital pulp therapy should only be performed in teeth reversible pulpitis [89]. Several studies included teeth with signs of irreversible pulpitis such as teeth with spontaneous pain [57,79,90], and teeth that were positive to percussion [57,79] were treated successfully.

In full pulpotomy studies [90-92], teeth with periapical radiolucency lesions were treated successfully with weighted pooled success rate of 92 % [72].

Moreover, histological studies demonstrate that cariously exposed vital pulp was not always completely infected. Occasionally the inflammation was localized adjacent to carious lesion, not spreading to the whole coronal or radicular pulp [72,93].

Apexogenesis

Apex=root end, genesis=formation.

Apexogenesis is defined as “a vital pulp therapy procedure to encourage continued physiological development and formation of the root end” [18]. Only inflamed pulp tissue should be removed and bioactive material is placed over remaining healthy pulp tissue.

The goals of apexogenesis are [19]:

- Allow continued development of root length.
- Maintain pulp vitality, thus allow continued deposition of dentin.
- Promoting root end closure, thus creating natural apical constriction.
- Generating dentine bridge at the site of pulpotomy.

Treatment of teeth with necrotic pulp and open apices

- Incomplete root development can provide a challenging clinical situation in treatments [20].
- Cleaning and shaping of blunderbuss canal is difficult.
- Necrotic debris in wide root canal is difficult to completely disinfect.
- Thin, fragile dentine walls are liable to fracture.
- Risk of extending materials beyond apex.

Three techniques were reported to obturate an immature tooth which involved the use of a root filling material without the induction of the apical closure [21].

- Placement of large gutta-percha or customized gutta-percha cone with sealer at the apex.
- Placement of gutta-percha with sealer short of apex.
- Periapical surgery.

However these techniques will not provide apical barrier. Another two techniques were reported aimed to provide apical barrier [21].

- Placement of calcium hydroxide to induce mineralized apical barrier
- Placement of biocompatible material such as dentine chips against which a root filling is placed.

Until recently, the traditional approach to treat immature apex is apexification [22].

Apexification

Apexification is histological term defined as “a method to induce a calcified barrier in a root with open apex” [18].

The infected necrotic pulp is removed up to the apex by means of mechanical debridement and anti-septic chemical irrigation [22].

Apical hard tissue barrier formation following apexification is reparative process of the dentine-pulp complex [23]. However, continued physiologic root development of immature permanent tooth with infected necrotic pulps and apical periodontitis after apexification procedure with calcium hydroxide was reported [24].

Contemporary materials

Calcium hydroxide

Calcium hydroxide is the most popular material to induce normal root development [25].

Calcium hydroxide is white odorless powder and is chemically classified as strong base with PH 12.5; in contact with aqueous fluids it dissociate into calcium and hydroxyl ions [25].

The antimicrobial activity of calcium hydroxide is related to the release of highly reactive hydroxyl ions in aqueous environment which mainly affect cytoplasmic membranes, proteins and DNA of bacterial cells [26].

The mineralizing action of calcium hydroxide is directly influenced by its high PH. The alkaline PH not only neutralize acids from osteoclasts but also activate alkaline phosphatase enzyme which plays a role in hard tissue formation [27]. However, calcium ions present in dentine bridge originated from systemic circulation [28].

Calcium hydroxide is biocompatible, but unfortunately has low compressive stress that is not compatible with condensation forces of amalgam, also solubility of calcium hydroxide in fluids is a problem as a reliable seal cannot be achieved so a good coronal seal is required in case of vital pulp therapy [29].

Mineral trioxide aggregate

Mineral trioxide aggregate is bioactive material that influences its surrounding environment.

MTA is currently marked in two forms: white and gray. MTA was introduced in gray but because of its discoloration potential, White MTA was developed [30].

Gray MTA basically consists of dicalcium, tricalcium silicate and bismuth oxide, whereas white MTA is composed of tricalcium silicate and bismuth oxide [31].

MTA is prepare by mixing its powder with sterile water in a 3:1 powder to liquid ratio that result in a colloid gel that solidifies into hard structure [32].

The PH value of MTA is 10.2 after mixing. This value rises to 12.5 at three hour [33].

The mean setting time of MTA is two hours and 45 minutes. At 24 hours MTA has the lowest compressive strength among materials (40 MPa), but increased after 21 days (67 MPa) [33]. A recent study reported that compressive strength of MTA is higher when mixed mechanically and placed by ultrasonic agitation than when mixed manually and placed without ultrasonic agitation [101].

The solubility of MTA is influenced by powder to water ratio. In fact, higher water to powder ratios increase MTA porosity and solubility [34].

MTA has an antibacterial and antifungal effect. Lowering the powder to liquid ratio might adversely affect the antibacterial and antifungal properties of MTA [35].

MTA has Cementoconductivity and cementoinductivity properties that have been confirmed when reasonable concentration of material (less than 20 mg/ml) have been used. MTA offers a biological substrate for cementoblasts attachment and growth as well as production of mineralized matrix gene (osteocalcin) and protein expression over MTA [36].

Indirect pulp capping

Indirect pulp capping is "procedure in which a material is placed on thin partition of remaining carious dentine that if removed might expose the pulp in immature permanent teeth" [18]. Also called "stepwise technique".

Indications [37]

- Permanent teeth.
- Normal response to pulp tests.
- No symptoms of pulpitis or with diagnosis of reversible pulpitis.
- Deep caries lesion radiographically with no apical pathosis.

Technique: (Figure 1)

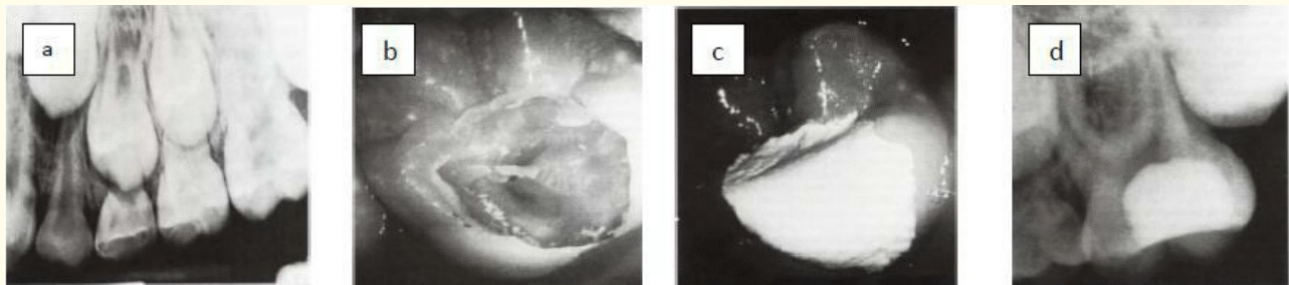


Figure 1: Indirect Pulp therapy. (a) Preoperative radiograph of maxillary first molar showing deep occlusal caries. (b) Caries are removed except a layer immediately over the pulp. (c) CH is applied to caries dentine. (d) Postoperative radiograph after 1 year, the remaining caries are removed and permanent restoration is placed.

This approach involves 2 steps process. The first step, the tooth is anesthetized and isolated with rubber dam, the carious dentine is removed completely from the dentin-enamel junction and only the softest infected dentine is excavated, leaving a carious layer over the pulp. Calcium hydroxide is then applied to the carious dentine. The objective is to change the cariogenic environment in order to decrease number of bacteria and close remaining caries from biofilm of the oral cavity. Subsequent radiographic reevaluation to ensure formation of dentine bridge and normal periapical status.

The second step, after 6 to 12 months allowing sufficient time for tertiary dentine formation, the carious lesion is reentered and the caries is removed to the formed dentine bridge and permanent restoration is placed [38]. However, what is critical to both steps of excavation is the placement of well-sealed restoration [39].

One step indirect pulp capping was also reported [40], which involves minimal removal of caries and sealed with permanent filling restoration without a return for additional excavation.

Two steps indirect pulp capping is also questioned of whether or not to have any advantage over one step treatment regimen. In fact, there may be two important disadvantages for the two steps technique. The first is the requirement of patient compliance and the assurance of return visit after 6 - 12 months. Secondly, a bacterial-tight seal is essential between visits. Thirdly, the two steps treatment is more expensive, requiring twice as much as operator and office time [41].

Mode of action

When thickness of remaining dentine including reactionary dentine between the front of bacterial penetration and the pulp is 1.1 mm or more, the pulpal inflammatory response to bacterial infection of the dentinal tubules is negligible. However, when the bacterial penetration in the dentinal tubules reaches to within 0.5 mm from the pulp, there is significant increase in the pulpal inflammation [42].

Indirect pulp capping intended to protect the primary odontoblast and the pulp from further injury. When the infected dentine is removed, the affected dentine can remineralize and promote odontoblasts to form reactionary dentine formation at the pulp-dentine junction [43].

Reactionary dentinogenesis is usually caused by milder injury of the dentin-pulp complex as slowly progressive caries, cavity preparation and biomaterial used as indirect pulp capping agents [44]. Reactionary dentinogenesis is secreted by functional upregulation of surviving primary odontoblasts, stimulated by signaling molecules such as growth factor proteins and bioactive molecules released from dentine matrix during injury to the dentin-pulp complex [44].

Treatment outcome

In a study [45] for long term the survival of indirect pulp treatment performed in primary and permanent teeth with clinically diagnosed deep caries, showed survival rate of 96% for 86 primary molars and 93% for 34 permanent molars, it was also found that glass ionomer cement may be preferred to calcium hydroxide (CH) as it prevents microleakage when compared with CH.

Many studies [46] was reported to determine whether complete excavation of caries has any advantage over stepwise treatment in an attempt to avoid pulp exposure if possible.

A critical review [47] of treatment of deep caries by complete excavation or partial removal indicate that complete removal of infected dentine approaching the pulp is not required for successful caries removal.

However, in systematic review [48] of caries and pulp treatment from Cochrane Database indicate lack of randomized clinical trials to support incomplete removal of caries.

In another systematic review [49] from Cochrane Database indicate that partial removal of caries is preferable to complete caries removal in deep caries. The review also indicated that studies used one step indirect pulp therapy did not reported adverse consequences over the two steps indirect pulp therapy.

In a recent randomized clinical trial [50] comparing complete excavation of deep caries with stepwise excavation gave less success rate (62.4%) for complete excavation than stepwise excavation (74.1%).

Direct pulp capping

Direct pulp capping is defined as “treatment of exposed vital pulp by sealing the pulpal wound with material such as mineral trioxide aggregate or calcium hydroxide to facilitate the formation of reparative dentine and maintain pulp vitality” [18].

Indications [37]

- Restorable permanent teeth.
- Carious or traumatic exposure with vital pulp.
- No symptoms of pulpitis or with diagnosis of reversible pulpitis.

Technique: (Figure 2,3)

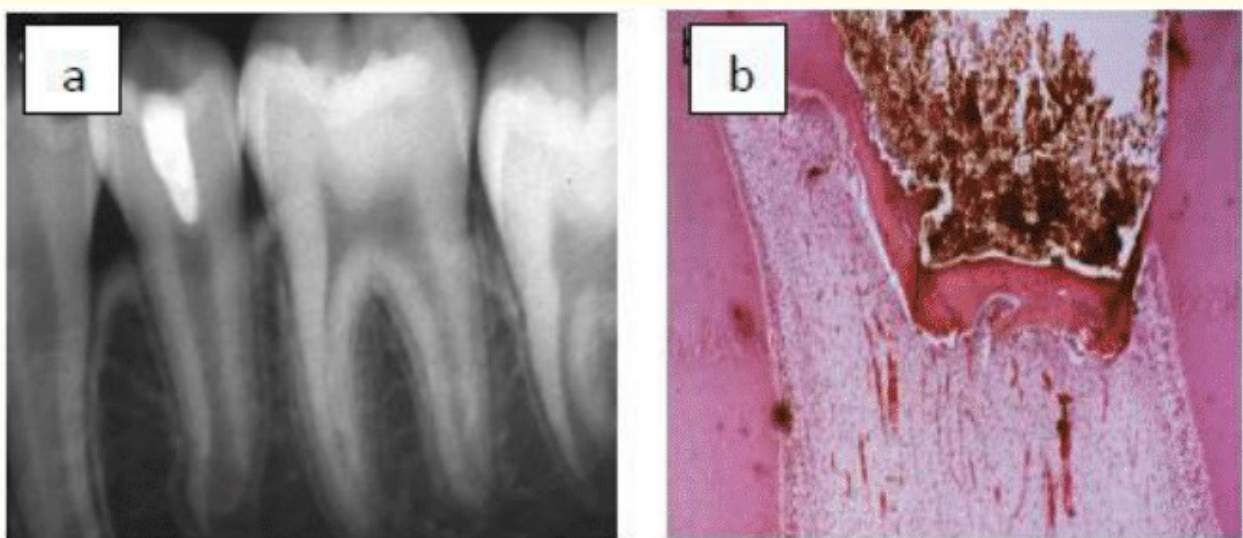


Figure 2: Direct pulp capping in lower premolar. (a) A premolar tooth capped directly with MTA. (b) Histologic section after six months shows formation of dentine bridge.



Figure 3: Direct pulp capping in lower molar. (a) Preoperative radiograph shows extensive caries. (b) The decay was removed and pulp exposure was capped by MTA.

The tooth is anaesthetized and isolated with rubber dam. All caries and undermined enamel are removed with low speed round carbide bur and excavator. Preferably, caries detector dye is applied for 10 seconds on dried caries dentine, and then all stained dentine is removed [51]. Bleeding will often occur with small exposures and can be controlled by placement of cotton pellet moisture with 3 - 6% sodium hypochlorite for 10 - 20 seconds. 2% chlorhexidine or 0.9% saline can be also used to stop bleeding. None of these agents negatively affected pulp healing at 90 days [52].

This step can be repeated for 10 minutes. If homeostasis is achieved within 10 minutes, hard setting calcium hydroxide is placed over pulpal tissue and sealed with permanent restoration [53].

If MTA will be as direct pulp capping agent, two steps approach will be taken. The first step, MTA should be mixed according to the manufacturer's instruction. MTA is placed directly over exposed pulp tissue and surrounding dentine. MTA should have at least 1.5 mm thickness.

A circumferential region of dentine and enamel approximately 1.5mm should be cleared of MTA, this will provide adequate area for efficient seal of the restoration. Flat moist cotton is placed over entire MTA and strong interim restoration is placed. The second step will be after 1 to 10 days. The tooth is examined for any signs or symptoms, and then interim restoration and cotton are removed. A definite restoration is placed. The patient is then recalled after 6 weeks when cold testing and symptoms are evaluated [54].

If bonded composite resin will be used immediately after placement of MTA, The unset MTA should be overlaid with a thin protective layer of resin modified glass ionomer or flowable resin before bonding procedure [55].

Pulp capping with adhesive resin was reported in many studies [56-58]. However these studies revealed significant better result, less pulp necrosis and thicker dentine bridge when calcium hydroxide was used as direct capping agent.

Mode of action

At the site of exposure, primary odontoblasts together with other pulpal cells are destroyed and inflammation is initiated. If infection/inflammation in coronal pulp is under control, odontoblast-like cells will regenerate.

Pulp capping materials such as MTA and calcium hydroxide induce the release of growth factor proteins and bio active molecules from dentin matrix. Post-natal stem cells in human dental pulp are capable of differentiating into odontoblast-like cells upon receiving induc-

tive signals from bioactive molecules and growth factor proteins. Reparative dentin is formed from odontoblast-like cells and not from primary odontoblasts as in case of indirect pulp capping [59].

Treatment outcome

For decade, calcium hydroxide-base materials have been the gold standard for direct pulp capping [60,61].

A retrospective study (62) reported success rate of direct pulp capping with calcium hydroxide after 1 year (80.1%), 5 years (68%) and 9 years (58.7%).

In another the long term study [63], the success rate of pulp capping with calcium hydroxide is 76.3% after 13.3 years.

These results confirm that success rates using CH diminish over time, primarily due to the dissolution of material under restorations and over dentinal bridges with tunnel defects [64].

An alternative to calcium hydroxide-based capping materials is MTA, which was developed by Torabinejad and co-workers at Loma Linda University in the 1990s [65].

High success rate (97.9%) after capping with MTA in cariously exposed permanent teeth and all teeth showed continued root formation [66].

In clinical assessment of carious permanent teeth capped with MTA and followed for 6, 12, 18, 24 months, showed success rate (93%) with evidence of continued root growth [67].

Many case reports and clinical studies compared treatment outcome of MTA and CH when used as direct pulp capping agents.

In preliminary study [68], 14 intact third maxillary molars teeth were capped with either CH or MTA. Histological examination of teeth showed dentinal bridge formation and mild chronic inflammation 2 months after pulp capping with MTA. While with CH, irregular and thin dentinal bridge was formed after 3 months with associated pulpal necrosis, hyperemia and inflammation.

Another histological study [69] revealed that all teeth capped with MTA showed dentinal bridge formation. In contrast, only 60% of CH-capped pulps showed hard tissue formation. Also MTA-capped pulps showed significantly thicker dentine bridge than those capped with CH. These results are similar to findings in another histological [70], that MTA-capped teeth were free from inflammation and showed dentine bridge formation within 3 months, while CH-capped teeth showed less consistent formation of dentine bridge that had numerous tunnel defects and persistence inflammation up to 3 months after capping.

A recent study [71] showed MTA is more effective than CH for maintaining long term pulp vitality after direct pulp capping.

A recent systematic review [72] for vital pulp therapy in vital permanent teeth with cariously exposed pulp revealed success rate of direct pulp capping was different depending on follow up time (> 6 months -1 year, 87.5%), (> 1 - 2 years, 95.4%), (> 2 - 3 years, 87.7%) and (> 3 years, 72.9%). Also showed that direct comparisons of weighed pooled success rate when using either CH or MTA as pulp capping medication revealed no statistically significant difference, while in indirect comparisons of weighted pooled success rate showed that MTA was superior to calcium hydroxide in direct pulp capping.

A recent retrospective study [55] aimed to evaluate success rate and treatment outcome of direct pulp capping in immature permanent teeth and to investigate potential factors (age, sex, tooth position, jaw, exposure site, capping material, temporary filling) contributing to the pulpal survival according to time. This study reveals survival rate (89.9%) after 1 year and (67.4%) after 3 years when MTA was used as direct capping agent. CH yielded (73.9%) and (52.5%) after 1 and 3 years respectively. Patients < 40 years age group had better survival rate than older ones, and when exposure site was limited to occlusal site there was better survival than when it was on axial side. Other potential factors not significantly affect survival rates.

Partial pulpotomy

Partial Pulpotomy is defined as “The surgical removal of coronal portion of vital as a means of preserving the vitality of the remaining coronal and radicular pulp tissues” [18].

Indications [37]

- Restorable permanent teeth.
- Deep Carious lesions or traumatic fractures longer than 24 hours with vital pulp exposure.
- When pulpal inflammation is expected to be greater than normal and pulpal bleeding cannot be controlled within several minutes.
- No symptoms of pulpitis or with diagnosis of reversible pulpitis.

Technique: (Figure 4)



Figure 4: Partial pulpotomy in upper central incisor. (a) Preoperative radiograph shows traumatic pulp exposure. (b) Partial pulpotomy capped with MTA. (c) Postoperative radiograph shows complete root formation.

The carious exposure generally occurs in the depth of a carious excavation. The cavity preparation presents convenient and effective location for the protection and seal of the pulp with pulp capping material. On the other hand, a traumatic fracture leaves the pulp exposed on the surface of the tooth and presents a problem in both retention of capping material and prevention of leakage. In addition to time of exposure, health of pulp before trauma, diameter of pulp exposure (1.5 mm diameter is the critical), age of the tooth, no concomitant luxation injury and the stage of root development [73]. To address many of these challenges, Cvek procedure [74] was developed.

The tooth is anaesthetized and isolated with rubber dam. A 1 - 2 mm deep cavity is prepared into the pulp using sterile diamond bur of appropriate size at high speed with copious coolant. If bleeding is excessive the pulp is excavated deeper and cotton pellet moisture with 3 - 6% sodium hypochlorite for 10 - 20 second is placed over the exposed pulp.

Sodium hydroxide serves as an excellent diagnostic tool to help to evaluate how far the inflammation has progressed into the pulp.

A profuse bleeding that is difficult to stop indicates severe pulpal inflammation and the decision to proceed with pulpotomy [75].

Any blood clot should be removed. If homeostasis is achieved, hard setting calcium hydroxide is placed over pulpal tissue and sealed with permanent restoration [76] or MTA is mixed according to the manufacturer’s instructions (3:1, powder to liquid), and placed directly

over exposed pulp tissue and surrounding dentine with at least 1.5 mm thickness [77]. Definitive restoration is either placed over MTA several days later or placed immediately after applying a layer flowable resin on unset MTA.

Mode of action

Similar to direct pulp capping, calcium hydroxide and mineral trioxide aggregate induce dentine bridge formation most likely by indirect mechanism through the release of growth factors from dentine matrix. Growth factors proteins stimulate the differentiation of post-natal stem cells in human dental pulp into odontoblasts-like cells that forms dentine bridge [59,78].

Treatment outcome

Studies have demonstrated that partial pulp capping of exposed permanent teeth with calcium hydroxide is an alternative treatment with good clinical outcome [79-81] and formation of reparative dentine [82].

Recently, mineral trioxide aggregate has been used successfully as pulp capping material after partial pulpotomy [83-85]. In a prospective clinical study [86], partial pulpotomy was performed by grey MTA in 28 teeth with carious exposures. The patients were followed for 24 months. 79% of these teeth had normal response to vitality tests and with no radiographic evidence of pathosis.

In another prospective clinical and radiographic study [87], comparing CH and MTA as pulp capping materials after partial pulpotomy in cariously exposed teeth, it was reported that no significant differences in radiographic and clinical success rates between CH (91%) and MTA (93%). Hard tissue barrier was noticed under CH (55%) compared with (64%) under MTA.

Long term evaluation of partial pulpotomy was (93.9%) after 49 months [88].

In a recent randomized clinical trial [50] comparing direct pulp capping and partial pulpotomy, showed no difference in pulp vitality at one year using CH.

A recent systematic review [72] for vital pulp therapy in vital permanent teeth with cariously exposed pulp revealed success rate of partial pulpotomy was different depending on follow up time (> 6 months -1 year, 97.6%), (> 1 - 2 years, 97.5%), (> 2 - 3 years, 97.6%) and (> 3 years, 99.4%). Also showed that direct comparisons of weighed pooled success rate when using either CH or MTA as pulp capping medication revealed no statistically significant difference, while in indirect comparisons of weighted pooled success rate showed that CH was superior to MTA in partial pulpotomy.

Full pulpotomy

Full pulpotomy is defined as “The removal of entire coronal pulp to the level of the root canal orifice or as much as 2-3 mm apical to the orifices” [18].

Indications [94]

- Restorable permanent teeth.
- Deep Carious lesions or traumatic fractures longer than 72 hours with vital pulp exposure.
- When pulpal bleeding cannot be controlled within 10 minutes.

The main disadvantage of full pulpotomy is that pulp testing is not possible due to the loss of coronal pulp tissue, so radiographic follow up is extremely important.

Technique [94]: (Figure 5)

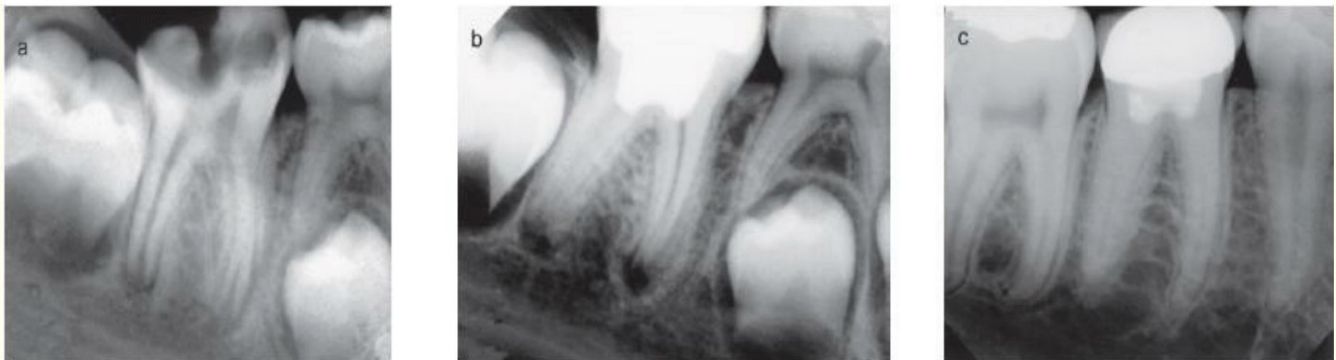


Figure 5: Pulpotomy in lower molar. (a) Preoperative radiograph with extensive decay and open apex. (b) After removal of caries and perform pulpotomy and restored with amalgam. (c) Postoperative radiograph after periapical tissue healing and apex closure.

Tooth is anaesthetized and isolated with rubber dam. The coronal pulp is removed to the level of root canal orifices. If there is bleeding, homeostasis is done by placing cotton pellet moisture with 3 - 6% sodium hypochlorite for 10 - 20 seconds. If bleeding cannot be controlled, then decision for pulpectomy should be taken. Any blood clot should be removed. If homeostasis is achieved, hard setting calcium hydroxide or MTA with bacterial tight seal and coronal restoration are carried out as with partial pulpotomy.

Mode of action

In response to pulpal dressing with calcium hydroxide or mineral trioxide aggregate apposition of hard tissue is achieved similarly as direct pulp capping and partial pulpotomy.

Dentine bridge formation following apexogenesis is a reparative process of dentine-pulp complex. However, the continued root formation is a normal physiological process [23].

Treatment outcome

- In a case report [95], pulpotomy showed successful result in treatment of dens invagination in 2 mandibular premolars.
- In histological study [96], comparing CH and MTA pulpotomy materials showed that teeth pulp-capped with MTA had significant less inflammation and more homogenous and continuous dentine bridge than those capped with CH.
- Another study [97] comparing CH and MTA as capping materials for 30 immature permanent teeth (15 with CH and 15 with MTA) and followed for 3, 6, 12 months showed that only 2 teeth that were capped with CH failed with association of pain and swelling. The authors recommend MTA as a suitable alternative to CH.
- Pulpotomy with MTA as pulp capping material was performed on symptomatic teeth with history of lingered pain [98] and histological observation showed complete dentin formation 2 months after treatment to all the treated teeth
- In another case series of pulpotomy with MTA as pulp capping material was performed on teeth with irreversible pulpitis [99] and followed for 19.7 months showed that only 1 of 19 teeth had persistence pain.
- Pulpotomy with CH as pulp capping material in symptomatic teeth [100] or with periapical involvement [90-92] showed complete resolution of symptoms, periapical healing and continued root formation.

- A recent systematic review [72] for vital pulp therapy in vital permanent teeth with cariously exposed pulp revealed success rate of full pulpotomy was different depending on follow up time (> 6 months -1 year, 94%), (> 1 - 2 years, 94.9%), (> 2 - 3 years, 96.9%) and (> 3 years, 99.3%). Also showed that direct and indirect comparisons of weighed pooled success rate when using either CH or MTA as pulp capping medication revealed that MTA was superior to CH in full pulpotomy.

Calcium hydroxide versus mineral trioxide aggregate in vital pulp therapies

The biological reactions resulting from application of bioactive materials to the pulp are confounded by several factors such as the formulation of applied material, concentration and rate of release of active ingredient, distance it has to diffuse in a chemically active form to the target cells, status of dentine, presence of infection and the health status of the pulp. It has been suggested that the type of biomaterial selected is of lesser consequence and that the quality of the seal of the cavity to prevent microbial ingress is the most important factor determining the success of the procedure [102].

Hard tissue induced by calcium hydroxide (Figure 6)

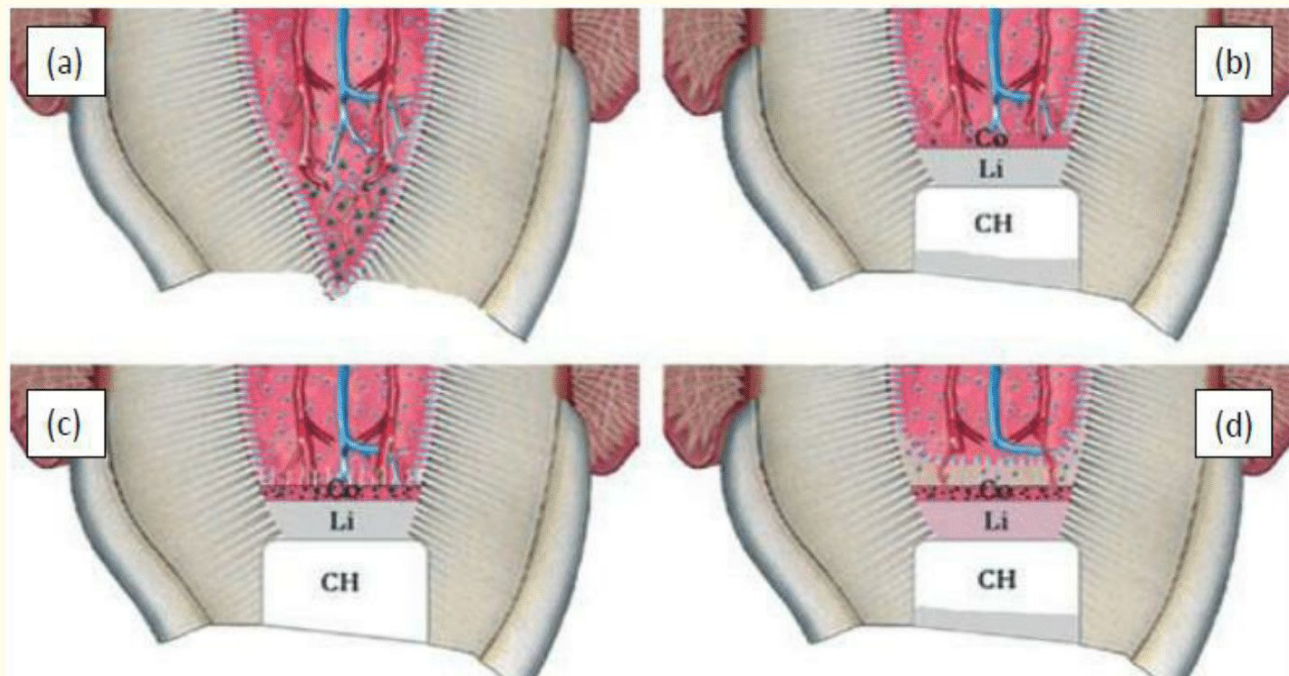


Figure 6: Calcium hydroxide and pulpal responses. (a) Exposed pulp. (b) The high PH generates a zone liquefaction necrosis and a zone of coagulation necrosis. (c) Spherical foci of calcification are formed that coalesce to form calcification zone and next to that collagen is formed. (d) Bone like tissue is deposited containing cells and vascular inclusions.

When CH is placed against the pulp, the high PH of CH results in zone of liquefaction necrosis subjacent to CH and a deeper zone of coagulation necrosis next to vital pulp within the first week (approximately 1.5 mm combined width in case of non-setting CH) [103]. Dentine matrix appeared around 30 days later once the cellular and vascular inflammatory events began to fade. Growth factor proteins, bone morphogenic proteins and bioactive molecules are released from dentin matrix and stimulate the differentiation of pulpal stem cells into odontoblasts-like cells [102]. Collagen is laid down approximating the necrotic zone and mineralized crystals are deposited in the

region. Thickening of barrier and appearance of dentine-like tissue has been reported to be observed at around 4 weeks after pulp capping procedure and then dentin deposition begins [102].

Drawbacks of calcium hydroxide

Length of time for induction of coronal hard tissue that ranges from 2 - 3 months [154].

Non-adhesive nature of the cement and its dissolution over time may lead to microleakage and entry of bacteria to exposure site [104]. It is important to ensure seal by a restoration overlying the cement [71]. Bacterial contamination may also occur through tunnel defects (imperfection in dentine bridge due to inclusion of blood vessels) [64].

Impaction of particles of capping agent, necrosing action of CH up to 1.5 mm compromising vascularity of the pulp and interference by blood clot are other factors responsible for failure of procedure Calcium hydroxide has limited solubility and diffusibility, its antimicrobial effect is limited to superficial layers of pulp, thus CH may be successful only in cases of minimal inflammation. If inflammation is severe the procedure generally fails [102].

Hard tissue induced by mineral trioxide aggregate (Figure 7)

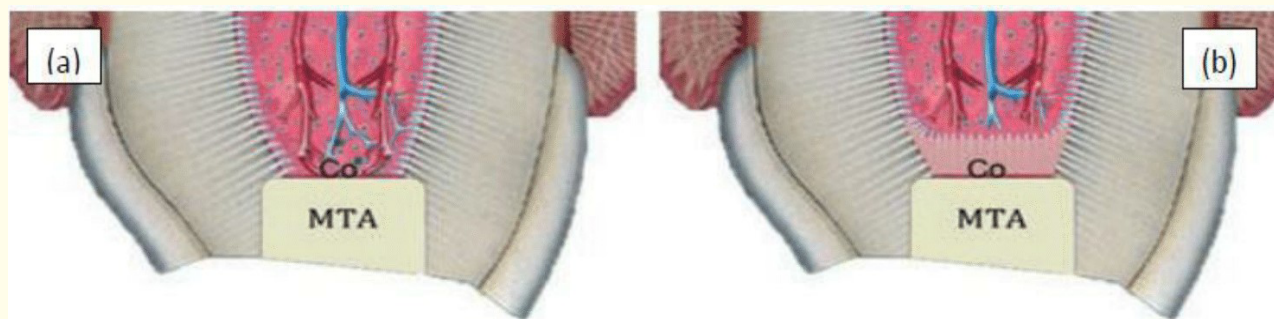


Figure 7: Mineral trioxide aggregate and pulpal responses. (a) High PH generates a very narrow zone of coagulation necrosis. (b) Next to that zone reparative dentinogenesis zone is formed.

When MTA is placed against the pulp, the high PH of MTA results in zone of coagulation necrosis next to vital pulp after one week. No zone of liquefaction necrosis is formed subjacent to MTA. High PH lasts for at least 8 weeks. Growth factor proteins, bone morphogenic proteins and bioactive molecules stimulate the differentiation of pulpal stem cells into odontoblasts-like cells which produce collagen matrix.

A regular hard tissue barrier with cellular inclusion was seen after 2 weeks [103].

The hard tissue barrier appears to be formed earlier than under CH, with larger daily dentin increments, few vascular inclusions and with less inflammation [105].

MTA forms a very tight seal where it contacts the dentin walls, most likely due to a chemical bond between MTA and dentin; a layer of hydroxyapatite is created as a link [106]. This layer prevents and reduces bacterial penetration to the pulp, enhance biocompatibility, sealing ability and dentinogenic activity of MTA. This seal might be the main cause of successful performance of this material [107].

Drawbacks of mineral trioxide aggregate Despite its many advantages, MTA has some drawbacks such as discoloration potential, presence of toxic elements in the material composition, difficult handling characteristics, long setting time, high material cost, an absence of a known solvent for this material and the difficulty of its removal after curing [108].

Internal bleaching was reported to be the treatment of discoloration due to MTA application in vital pulp therapy [109].

It should also be noted that the total amount of arsenic (that can cause toxicity) in all types of MTA is insignificant [110].

Apexification

Apexification is defined as “a method of inducing a calcified barrier in a root with open apex or the continued apical development of incompletely formed root in teeth with necrotic pulp” [18].

Indications [37]

- Permanent teeth.
- Non-vital pulp with open apex and thin dentin walls.
- Standard instrumentation technique cannot create apical stop.

Technique: (Figure 8)

The tooth should be isolated with rubber dam. The first phase of treatment is to disinfect the root canal system to ensure periapical healing [22]. The canal length is estimated with a parallel preoperative radiograph, and after access to the canals is made, a file is placed to the determined length and the working length is confirmed by radiograph. Very light filling with copious irrigation using 0.5 % sodium hypochlorite is performed. The canal is dried with paper points and creamy mix of calcium hydroxide is placed into the canal and sealed with temporary restoration for 1 to 4 weeks until there is no sign of infection [94].

Traditional apexification

Pure calcium hydroxide powder is mixed with sterile saline to a thick consistency. Ready-mixed commercial calcium hydroxide can also be used [111]. The calcium hydroxide is packed against the apical soft tissue with a plugger. This step is followed by backfilling with calcium hydroxide to completely fill the canal. The excess is removed from access cavity to level of the root orifices, and a well-sealing temporary restoration. A radiograph is taken in order to evaluate formation of hard tissue barrier and washout of CH [94].

Controversy exists as to whether or how often the calcium hydroxide should be changed.

Several studies [112,113,153] suggested placing the paste only once and waiting for radiographic evidence of barrier formation, and that there was nothing gained by repeated root filling either monthly or after 3 months. A recent study [114] reported that apical barrier was formed in short time (30 weeks) due to the minimal disturbance of apical area by avoiding repeated instrumentation and dressing changing. On the other hand, a number of studies [115-118] suggested that regular replacement of dressing allows clinical assessment of barrier formation and may increase the speed of bridge formation. This is due to the solubility of CH when comes in contact with tissue fluids [25]. Other investigators [141,142] suggested that calcium hydroxide should be replaced only when symptoms develop or the material dissolves out of the canal when assessed radiographically. However, the size of foramen opening and stage of development may be important factors to determine the time to replace the dressing [115]. Radiograph may not be a relevant to evaluate calcium hydroxide washout; because the additives (barium sulphate) that are responsible for CH radio-opacity do not washout as readily as calcium hydroxide so that if they are present in the canal, evaluation of washout is not possible [94]. When hard tissue barrier is indicated on radiograph, the calcium hydroxide is washed out with sodium hypochlorite and apical barrier is detected gently by a file or a paper point. The entire canal is then filled with thermoplasticized gutta percha [94].

One step/visit apexification (Figure 8)

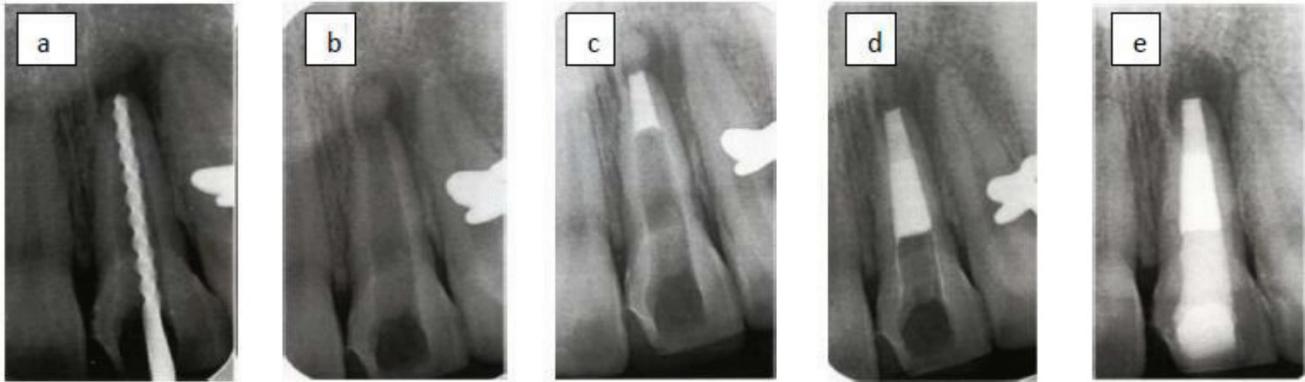


Figure 8: Apexification with MTA as apical plug. (a) The canal is disinfected with copious irrigation. (b) Calcium phosphate is placed through the apex as a barrier. (c) 4 mm of MTA is placed at the apex. (d) The canal is filled with thermoplastized gutta-percha. (e) Bonded resin restoration is placed at level below CEJ.

MTA can be used as apical plug or even as a root canal obturation [174].

If MTA will be used, calcium sulphate is pushed through the apex to provide a resorbable extraradicular matrix against which to pack the MTA [144]. Other types of matrix were also reported as collagen matrix [145]. However, MTA can be placed without matrix and care must be taken as MTA should be confined to root canal end [146]. MTA should be mixed according to manufacturer’s instructions to a thick creamy consistency and placed 1 - 1.5 mm short of working length using a carrier and condensed with minimal pressure. This is repeated until approximately 5 mm of material is deposited apically [119]. Moist cotton is placed in the canal for at least 6 hours and the entire canal is filled with thermoplasticized gutta percha [120] or the root filling is placed immediately over MTA because the tissue fluid of the open apex will probably provide enough moisture for MTA to set [121].

Soften filling technique is indicated in these canals because the apical diameter is larger than the coronal diameter. Care must be taken to avoid excessive lateral forces during obturation technique [94].

It was reported that short term canal medication with calcium hydroxide followed by an apical plug of MTA may increase the marginal adaptation of MTA and increase long-term prognosis for teeth with necrotic pulps and open apices [122,123]. Because the acidic environment due to inflammation can cause porosity in set MTA and affect its sealing ability [174,175].

Mode of action

In apexification procedures, the infected necrotic is removed up to the apex by means of mechanical debridement and antiseptic chemical irrigation [23]. Accordingly it is unlikely that reparative dentine can be formed because of the absence of pulp.

The calcified barrier at open apex has been described as cementoid-like tissue and osteoid-like tissue [124].

Calcium hydroxide and mineral trioxide aggregate stimulate release of growth factors and bioactive molecules from cementum matrix and signal progenitor/stem cells in periodontal ligament to differentiate into cementoblast-like cells that forms cementoid tissue [125].

CH and MTA also stimulate release of growth factors and bioactive molecules from alveolar bone marrow matrix and signal progenitor/stem cells in periodontal ligament and mesenchymal stem cells in alveolar bone marrow to differentiate into osteoblastic-like cells that forms osteoid tissue [126].

It was shown that MTA offers a biological substrate for cementoblasts attachment and growth as well as production of mineralized matrix gene (osteocalcin) and protein expression over MTA [36].

Continued physiological root development of immature permanent tooth with infected necrotic pulps and apical periodontitis after apexification procedure treated with calcium hydroxide was reported [24,143]. The newly formed root apical to the hard tissue barrier had normal root structure with pulp tissue in the canal space surrounded by dentine and cementum. It appears in this case that apical papilla and Hertwig's epithelial root sheath (HERS) have survived despite apical periodontitis. HERS cells are capable of signalling stem cells in apical papilla to differentiate into odontoblasts and produce root dentine [143].

Treatment outcome

- In a clinical study [127], apexification performed with calcium hydroxide in 34 teeth revealed that only 4 teeth failed to form hard tissue barrier.
- In a large retrospective clinical study [128], 328 luxated non-vital incisors were treated with CH and followed for 4 years. This study showed periapical healing in 95 % of teeth and increased frequency of fracture from 77% in least developed teeth to 28% in most developed roots.
- Two large clinical studies [129,130] involved treatment with CH and subsequent gutta-percha filling have shown average healing rate 95%.
- In a review of ten studies [111], the use of CH for apical barrier formation was successful in 74 - 100% of cases irrespective of the proprietary brand used.
- Continued physiological root development of immature permanent tooth with infected necrotic pulps and apical periodontitis after apexification procedure treated with calcium hydroxide was reported [24,143]. The newly formed root apical to the hard tissue barrier had normal root structure with pulp tissue in the canal space surrounded by dentine and cementum.
- In another retrospective study [174], 23 necrotic immature teeth treated for either long term with CH (n = 16) or short term with MTA (n = 7). All necrotic immature teeth achieved a nearly normal root development after (mean 16 months).
- In a case series study [131], a successful outcome in 10 of 11 teeth with necrotic pulps and open apices after application of MTA was observed and apical barrier formation after 24 months.
- In two studies [132,121] of necrotic pulps and open apices treated either with white MTA or grey MTA as apical barriers, radiographic examination revealed 81% success for these cases.
- In another case series study [133], used MTA as apical plug in 17 incisors and followed for 12.53 months showed that 94.1% were assessed as being successful clinically, whereas 76.5% were reported to be successful radiographically.
- In a large retrospective clinical investigation [134] on teeth with necrotic pulps and open apices that were treated with MTA as apical plug comparing 1 visit with 2 visits procedures, showed no significant difference between the two treatment protocols.
- A recent clinical and radiographic assessment [135] of two types of White MTA used as apical plug after initial dressing with CH showed overall success rate 95.5% after 23.4 months follow up.
- Another recent cohort study [136] assessing outcome of treatment of open apices with orthograde placement of MTA apical plug, reported that teeth with and without preoperative periapical radiolucencies showed healed rates of 85% and 96% respectively after 4 years follow up.
- In a comparative study [137] on permanent maxillary incisors with CH or MTA, demonstrated that mean time for CH (7 months) to form hard tissue barrier is significantly longer than time needed by MTA (3 months) to induce similar barrier.
- Another investigation [138] compared MTA with CH for treatment of teeth with immature roots and observed for 12 months. None of MTA cases showed signs of clinical and radiographic failure, whereas 2 of 15 teeth in CH group had tenderness to percussion and persistent periapical inflammation.

- In a report of three cases [147] describing treatment outcome of unintended extrusion of MTA into periapical tissue during apical barrier treatment showed that only in one case after 4 years follow up MTA had resorbed and periapical lesion had healed while in two other cases the patient remained symptomatic and scheduled for periapical surgery.
- Another two studies [148,149] revealed that complete periapical healing is possible despite extrusion of MTA periapically.
- A meta-analysis [139] of 30 article published in recent years indicated that MTA has high clinical success rate, provides best seal, shows superior biocompatibility, and the only root end filling when compared with amalgam, IRM, super EBA.
- A recent systematic review and meta-analysis [140] comparing MTA and CH as apical barriers used in apexification of immature permanent teeth showed that regarding clinical success and apical barrier formation, either CH or MTA may be used for apexification of immature teeth.

Calcium hydroxide versus mineral trioxide aggregate in non-vital pulp therapies

Apexification technique first requires elimination of bacterial infection from root canal and prevention of re-infection of this space [22]. Disinfection of the root canal space is straight forward for most cases; however, there is no natural apical construction or stop against which a suitable root filling material can be placed to prevent re-infection of this space [20]. Therefore one of the aims of treatment is to produce apical barrier against which a root canal filling can be placed and prevent extrusion of material into surrounding tissue.

Studies vary in assessment of time required for apical formation in apexification using calcium hydroxide.

In a review of ten studies [111], reported an average for apical barrier formation ranging from 5 - 20 months.

In another study [118] of 44 non-vital teeth undergoing CH apexification, found that the mean time to barrier formation was ranging from 4-16 months. The strongest predictor of rapid barrier formation was the rate of change of calcium hydroxide and also the narrower initial apical width the faster apical barrier formation.

It was reported that the infection or the presence of periapical radiolucency at the start of treatment increases the time required for barrier formation [141].

However, other studies [130,150] refuted this finding and stated that there is no relation between preoperative infection or radiolucency and rate of formation of apical barrier.

It was found [151] that in the presence of symptoms, the time required for apical closure was extended by approximately 5 months to an average of 15.9 months.

Comparative study [137] on permanent maxillary incisors with CH or MTA, demonstrated that mean time for CH (7 months) to form hard tissue barrier is significantly longer than time needed by MTA (3 months) to induce similar barrier.

Hard tissue induced by calcium hydroxide (Figure 9)

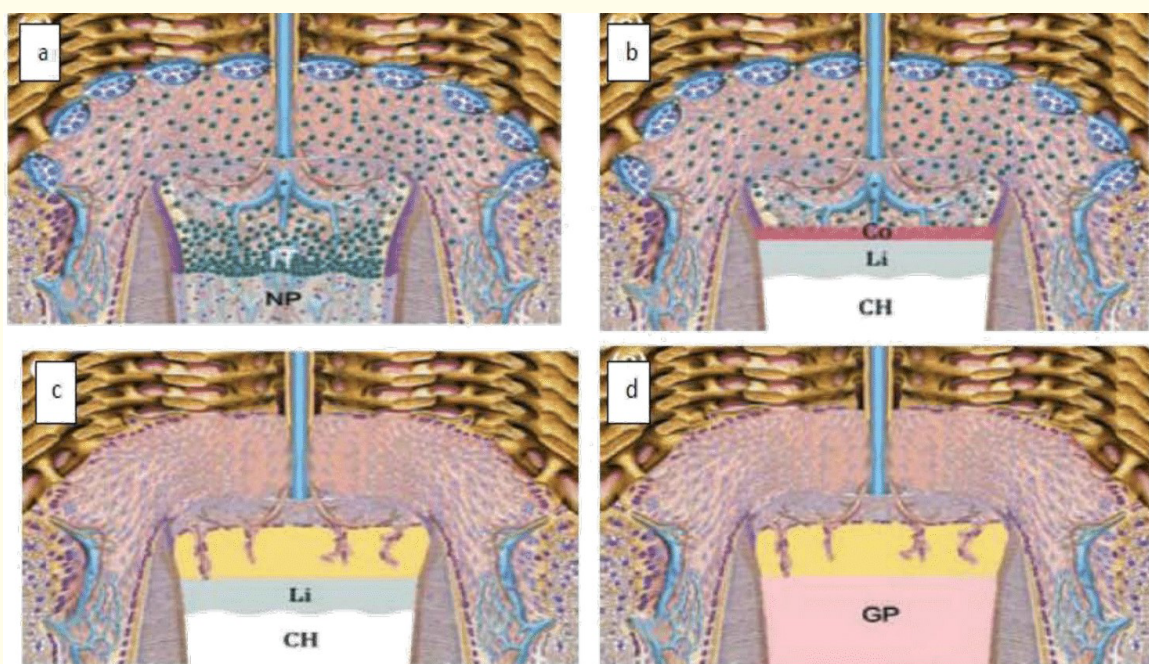


Figure 9: Calcium hydroxide and periapical responses. (a) Infected pulp and inflammation. (b) The high PH generates a zone liquefaction necrosis and a zone of coagulation necrosis. (c) New hard tissue forming cells from periapical tissue and formation of cementoid and osteoid tissue. (d) Root filling with gutta-percha, notice vascular inclusions within hard apical barrier.

When CH is placed against periapical tissues, the high PH value of CH results in zone of liquefaction necrosis subjacent to CH and a deeper zone of coagulation necrosis next to periapical tissues [103]. This coagulation necrosis zone stimulates release of growth factors (wound healing signals) and bioactive molecules from cementum matrix and alveolar bone marrow matrix. The response to these factors appear to be recruitment of new hard tissue forming from apical tissue, these are usually of cementoblastic but may also be osteoblastic origin [152]. During this process numerous vascular inclusions may occur [152].

Drawbacks of calcium hydroxide

Length of time for induction of apical hard that ranges from 6 - 18 months [111,118]. Such lengths of time may delay completion of treatment and failure to comply the patient.

Incomplete apical hard tissue barrier occurs due to vascular inclusions and may allow bacterial invasion through these defects [152].

Long term calcium hydroxide dressing cause changes in physical structure of dentine [155,156]. Such changes may weaken the dentine and lead to root fractures [128,157,158].

However, in a recent systematic review [159] of the effect of non-setting calcium hydroxide and mechanical properties of dentine showed that there are no clinical trials examining the relation between calcium hydroxide intra-canal dressing and root fracture. However, the majority of in vitro studies showed reduction in mechanical properties of radicular dentine after exposure to CH for 5 weeks or longer.

Hard tissue induced by mineral trioxide aggregate (Figure 10)

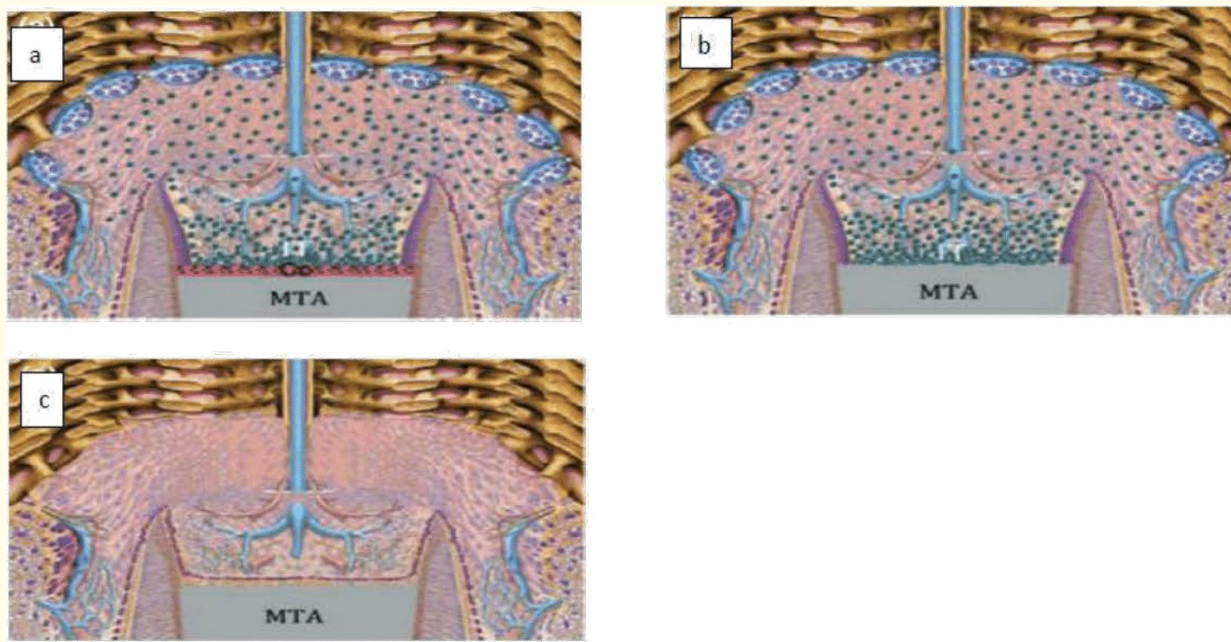


Figure 10: Mineral trioxide aggregate and pulpal responses. (a) MTA induce release of wound healing signals. (b) High PH generates a very narrow zone of coagulation necrosis. (c) Next to that zone reparative dentinogenesis zone is formed.

When MTA is placed against periapical tissues, the high PH value of MTA results in very narrow zone of coagulation necrosis next to periapical tissues [103]. MTA modulates cytokines production and encourages differentiation and migration of hard tissue producing cells, whereby hydroxyapatite is formed on MTA surface and biological seal is created [108].

Drawbacks of mineral trioxide aggregate

MTA has some drawbacks such as difficult handling characteristics, long setting time [108]. The porosity in set MTA has been identified as potential drawback for the material. It can allow potential penetration of bacteria or by-products. However, studies [107,160] have shown after MTA placement, a layer of hydroxyapatite forms over the material that fills the voids and surface defects.

Reinforcement of thin dentinal walls

Root strengthening interventions can be divided into the following four categories [161]:

Adhesive resins as root strengthening material

The use of adhesive sealers in the root canal system might possibly enhance resistance to fracture of thin weak dentinal walls as in immature permanent teeth [162].

Glass ionomer cements as root strengthening material

Glass ionomer cements have been shown to have long term adhesion effects by bonding to the hydroxyapatite component of dentine [163].

Intra-canal composite resin as strengthening root material

The use of newer dentine bonding techniques has been shown to strengthen the thin dentine walls of immature teeth [164]. Greater resistance to root fracture after placement of 4 mm thick apical plug of MTA followed by intra-canal composite resin when compared with MTA followed by gutta-percha and sealer was reported [165]. Before placement of composite, the internal dentine walls of canal are acid etched and dentine bonding agent is applied to the walls. 2 mm increment of condensable light cure composite resin is placed in the canal and cured. This step is repeated until the canal and access opening are obliterated [166]. However, due to incomplete polymerization in deeper depths of the canal because of limited transmission of light through the material, clear plastic posts have been developed to allow light transmission throughout the canal and cure entire mass of composite [167].

Resin modified glass ionomer with translucent curing post showed significant increased resistance to root fracture when compared with gutta-percha alone in open apex [168].

Fibre glass posts

Fibre-reinforced composite posts have been reported to have biomechanical properties close to those of dentine [169] and similar modulus of elasticity to that of dentine [170]. This seems to reduce stress transmission to the root canal walls by the post and thus avoiding possible root fracture [171,172].

Revitalization treatment

Revitalization is a new treatment procedure that may replace apexification procedures. Is defined as “The attempts to revive tissues in the pulp space and continue root formation in immature teeth with non-vital pulp” [175].

In this treatment, the ideal goal is to prepare an appropriate environment inside the root canal space (i.e. absence of bacteria and necrotic pulp tissue, presence of a scaffold and a tight coronal seal) that promotes repopulation of these stem cells, regeneration of pulp tissue, and continuation of root development [204].

Indications and recommendations [176]

- The traumatized tooth must be non-vital and not be suitable for apexogenesis, apexification, partial pulpotomy, or root canal obturation treatments.

- The tooth must be permanent and immature with open apex that is wide to a diameter of 1.1 mm or larger and exposed pulp.
- The patient must be aged 7 - 16 years, in good health and have parents guardians willing to take them to attend multiple appointments.
- Antibiotic paste can be used as additional disinfectant to sodium hydroxide and the patient needs to be warned about the potential for discoloration.
- An anesthetic without a vasoconstrictor should be used when attempting to induce bleeding into the root canal.
- A thin layer of white MTA or calcium hydroxide should be placed over the blood clot.
- An endodontic sealer is not compatible and cannot be used.
- The tooth should be restored with resin modified glass ionomer to help prevent microleakage.

Technique: (Figure 11)

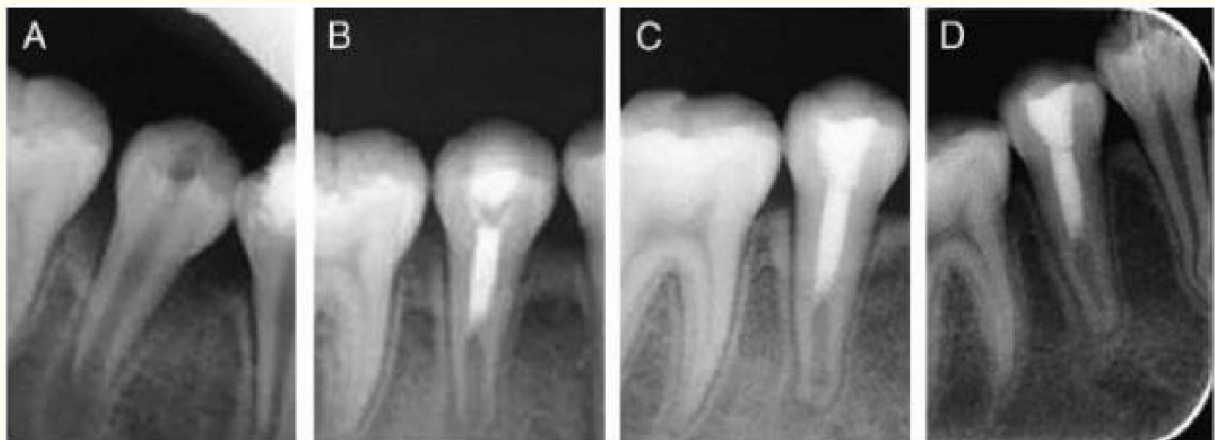


Figure 11: Revitalization procedure. (a) Preoperative radiograph with periapical lesion and open apex. (b) After 3 months elongation of root was evident. (c) After 8 months shows narrowing of apex and thickening of dentine walls. (d) After 18 months shows healing of lesion and apical closure.

Several case reports and case series have recently introduced detailed regenerative endodontic techniques applied to cases of necrotic immature permanent teeth [177-180].

The tooth is anesthetized and isolated with rubber dam. The canal is disinfected without mechanical instrumentation but with copious irrigation with 2.5% sodium hypochlorite. After drying the canal, tri-antibiotic paste, sometimes called Hoshino's paste [181]. The paste contains 200 mg ciprofloxacin, 500 mg metronidazole, and 100 mg minocycline. The triple antibiotic paste is placed in contact with necrotic pulp inside the root canal for up to 1 month before revascularization procedure. After one month, antibiotic is removed from canal by irrigation with 2.5% sodium hypochlorite and 17% ethylene diamine-tetracetic acid. Conditioning the dentin surface with ethylenediaminetetraacetic acid may enhance the adherence and differentiation of dental pulp stem cells during pulp regeneration [182,202].

Bleeding is induced till the level of canal orifice by passing a file beyond working length and causing irritation in periapical tissue. The bleeding is left for 15 minutes so that the blood would clot. Blood clot serves as a protein scaffold and permitting three-dimensional in-growth of tissue. MTA is placed over blood clot followed by moist cotton and well-sealed temporary restoration. After few days the cotton is removed and bonded restoration is placed over set MTA.

Patient should be recalled every 6 month for clinical and radiographic examination to ensure thickening of dentine walls and continued root formation.

Staining of the dentin by minocycline, a derivative of tetracycline, has been reported [183,184]. Therefore, some authors suggest eliminating the minocycline and keeping only metronidazole and ciprofloxacin in the antibiotic paste (bipaste) [185]. While others suggest that sealing dentinal tubules with dentine bonding agent may reduce the intensity of discoloration but did not prevent it [184,186].

Calcium hydroxide paste is another intracanal medicament that has been used to disinfect the canal before inducing bleeding [187,188]. However CH may not be suitable if there is remaining vital pulp tissue in the canal. The direct contact of CH paste with tissue will induce formation of layer of calcified tissue which may occlude the pulp space [189]. Another concern is that CH may damage the Hertwig's epithelial root sheath and thereby destroy its ability to induce the nearby undifferentiated cells into odontoblasts [180].

Another technique was reported [190,191], without inducing intracanal bleeding and all cases showed noticeable apical maturation with increased root length but a significant narrowing of canal space.

Mode of action

The precise nature of hard tissue formed inside the canal space and continued root development of immature permanent teeth with apical periodontitis/abscess following revascularization procedures in humans is not known, because no histological studies are available. It has been assumed that periodontal ligament tissue might grow into the canal space and deposit cementum on the canal walls after revascularization procedures [190].

HERS cells play an essential role in root development. If HERS cells survived in apical periodontitis/abscess after revascularization procedures, they can signal progenitor/stem cells in the periodontal ligament to differentiate into cementoblast-like cells and produce cementum-like tissue to promote root development [143]. If the apical papilla also survived in apical periodontitis/abscess after revascularization procedures, HERS cells will signal apical papilla cells to differentiate into root primary odontoblasts and promote root dentine formation [192]. Thus, the incompletely formed root will continue its normal physiological development after revascularization procedures.

However, no pulp-like tissue or dentine formation was present in the apical portion of the canal space of immature teeth with apical periodontitis/abscess after revascularization procedures in animal studies [193]. This indicates that the apical papilla did not survive.

Continued root development was evidenced by apical deposition of cementum without dentin after revascularization procedures in animal studies, probably because HERS cells survived in apical periodontitis [194].

When the canal dentine was treated with 17% ethylenediaminetetraacetic acid (EDTA) to expose dentine matrix, dentin-associated mineralized tissue appeared to be tightly attached to the canal dentine walls. In addition, EDTA may also cause the release of bioactive growth factors from dentine matrix [182,202].

If the tissues present inside the canal space in human immature permanent teeth with apical periodontitis/abscess after revascularization procedures are similar to that observed in animals, then revascularization is a reparative process with the loss of pulp biological function. Cementoid and osteoid tissues are not normally present as part of the pulp tissue. These tissues inside the canal space may function like periodontal tissues [175]. Histological studies are necessary to verify whether repair or regeneration of the pulp tissue occurs inside the root canal space following revascularization procedures in humans.

In immature permanent teeth with a long-standing apical periodontitis/abscess in adult patients, the apical papillae and HERS cells may be severely damaged, thus hindering the potential of continued root development. Furthermore, the functionality of stem cells is age-related decline owing to change in the intrinsic factors of stem cells and diminished signals within the extrinsic local and systemic environment that modulate the function of stem cells or their progeny [23].

The incompletely developed roots of immature permanent teeth with infected necrotic pulp and apical periodontitis are not attributable to the destruction of previously formed roots but are attributable to the inhibition or damage of HERS cells. Therefore, continued root development of human immature permanent teeth with infected necrotic pulp and apical periodontitis/abscess after revascularization procedures does not appear to be a regenerative process. It could be considered a physiological process if both apical papilla and HERS survive in apical periodontitis/abscess after periapical wound healing [23].

Regeneration of the dentine–pulp complex after complete pulp necrosis or irreversible pulpitis with apical periodontitis is likely to occur only by tissue engineering [196] or gene therapy [197].

Treatment outcome

- Most of evidence for revascularization procedures involves successful case reports and case series [177,179,180].
- The use of triple antibiotic paste was reported [181] to be effective for disinfection of infected root canals, setting the conditions for subsequent revascularization and showed healing of sinus tract [198].
- A few cases have reported positive response to cold/EPT after revascularization procedures [178,183,198,199]. However it was found that both the coronal level of regenerated tissue and the thickness of filling materials placed over this tissue affect the presence or absence of responses to EPT and cold. The authors reported positive responses to both EPT and cold in an immature maxillary premolar with a coronal MTA plug placed close to the cemento-enamel level [199].
- Cases treated with calcium hydroxide paste as intracanal medicament to disinfect the canal before inducing bleeding showed continued root development and thickening of dentin walls [187,188].
- In a recent case report [200] of revascularization treatment outcome of 2 immature necrotic mandibular first molars with symptomatic apical periodontitis and chronic apical abscess, respectively. In both teeth, the root canals received 3-week disinfection with the triple antibiotic paste. After 18 and 15 months (cases 1 and 2, respectively), the teeth showed advanced root development, thickening of the root walls, and complete peri-radicular healing.
- In a recent study [201] reporting long term outcome of three cases with necrotic immature teeth and followed for 24, 48 and 42 months, showed that all of the revascularized teeth showed satisfactory clinical and radiographical performances at long-term follow-ups. In well-selected cases, the revascularization technique may be a trustworthy alternative to conventional apexogenesis or apexification.
- A recent study [202] examining the effect of different irrigant on the survival of human stem cells of the apical papilla showed that the irrigation protocols evaluated in this study that contained EDTA promoted SCAP survival, whereas protocols in which 2% CHX was used appeared cytotoxic with no viable cells.
- Also showed that chelating effect of EDTA promotes the release of dentin-derived growth factors that were previously embedded into dentin during the process of dentinogenesis. These growth factors have been shown to foster proliferation, survival, and differentiation of dental stem cells.
- A recent *in vitro* study [203] examining the effect of medications used in regeneration techniques on chemical structure of immature human radicular dentine showed that long-term exposure of radicular dentin to $\text{Ca}(\text{OH})_2$ or antibiotic pastes might negatively affect the mechanical properties of dentin and increase the susceptibility to root fracture because of either collagen degradation in the case of $\text{Ca}(\text{OH})_2$ or excessive demineralization in the case of antibiotic pastes.

The null hypothesis that the 3 intracanal medicaments (triantibiotic, biantibiotic pastes and calcium hydroxide) have no significant effect on the chemical structure of radicular dentin was rejected.

A retrospective evaluation [205] of radiographic outcomes discovered that regenerative endodontic treatment with triple antibiotic dressing increased root wall thickness significantly more than either calcium hydroxide or formocresol. In addition, this study revealed

that in cases disinfected with calcium hydroxide, the radiographic location of calcium hydroxide inside the root canal space influenced the root development. When calcium hydroxide was radiographically limited to coronal half of the root canal space, the increase in root wall thickness was greater than when it was placed beyond coronal half.

Drawbacks of revitalization

There are several drawbacks and unfavorable outcomes of regeneration procedure [212].

Discoloration

A study [184] demonstrated that the main reason for tooth discoloration after treatment was the contact of minocycline in the triple antibiotic paste with coronal dentinal walls during treatment procedure. A recent study [186] suggested sealing dentinal walls of the access cavity by using dentin bonding agent and composite resin before placement of triple antibiotic paste inside the canal. A practical way to prevent discoloration is replacing minocycline with an antibiotic that does not stain teeth. It was reported [206] that successful regenerative endodontic treatment of a maxillary central incisor by using cefaclor instead of minocycline in the antibiotic mixture.

Another two studies [191,207] introduced novel method to shorten the treatment period and also prevent tooth discoloration by omitting the intracanal medication process.

Treatment period

The required time for disinfection of the root canal space with triple antibiotic paste or calcium hydroxide and increased number of clinical sessions (compared with one-visit MTA apical barrier technique) are other drawbacks of regenerative endodontic treatment.

The time spent on antibiotic disinfection in clinical studies varies between one [178] to eleven weeks [206].

Poor root development

Ideal root development pattern in immature teeth includes increase in root length, increase in root wall thickness, and formation of the root apex. In some studies, the outcome of regenerative endodontic treatments of necrotic immature teeth was lower than ideal, including absence of increase in root length [183,200], absence of increase in root wall thickness [183,208], or lack of formation of tooth apex [208]. Formation of a hard-tissue barrier inside the canal between the coronal MTA plug and the root apex is another reported unfavorable outcome [208].

A recent study [208] revealed that root development potential of immature necrotic teeth is related to the vitality of Hertwig epithelial root sheath. Therefore, there might be a correlation between dental history and quality of root development; the longer the duration of pulp necrosis, the lower the quality of root development after regenerative endodontic treatments.

Insufficient bleeding

A recent study [209] revealed that mesenchymal stem cells are delivered into root canal space after bleeding induction in human teeth, a phenomenon that did not happen in the absence of blood clot inside the disinfected root canal space.

An animal study [210] demonstrated that root canals that had a blood clot formation inside them after disinfection had better radiographic outcome compared with those without blood clot. To facilitate bleeding after root canal disinfection, using local anesthetics without vasoconstrictors is recommended [183]. However, lack of bleeding or insufficient bleeding after using plain local anesthetics is reported [183,208]. On the other hand, several cases with sufficient bleeding and lack of root development have been reported [183,211]. In addition, there are several reports [173,190,191] of successful regenerative endodontic treatment and continued root development without bleeding induction.

Root canal calcification

Root canal calcification/obliteration is another problem after regenerative endodontic treatment of necrotic immature teeth that is reported in cases disinfected by calcium hydroxide [190,191,208].

On the other hand, there is no report of complete root canal calcification/obliteration in cases disinfected by triple antibiotic paste.

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